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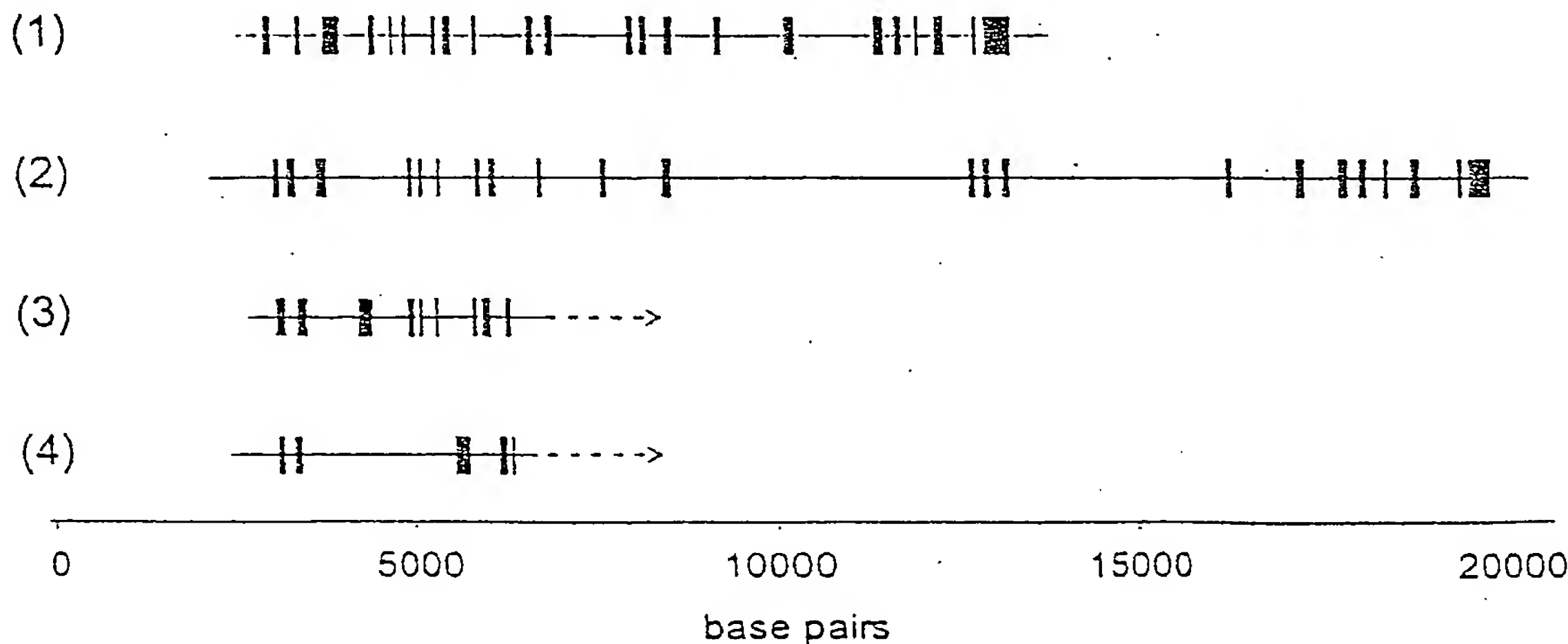
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(54) Title: STARCH BRANCHING ENZYME



(57) Abstract: This invention relates to a new starch branching enzyme, and to the gene encoding the enzyme. In particular, the invention provides a new starch branching enzyme type II from wheat, the nucleic acid encoding the enzyme, and constructs comprising the nucleic acid. The invention also relates to a novel method for identification of branching enzyme type II proteins, which is useful for screening wheat germplasm for null or altered alleles of wheat branching enzyme IIb. The novel gene, protein and methods of the invention are useful in production of plants which produce grain with novel properties, for example wheat grain containing high amylose or low amylopectin starch. Applicants: Zhongyi Li et al.

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STARCH BRANCHING ENZYME

This invention relates to a new starch branching enzyme, and to the gene encoding the enzyme. In particular, the invention relates to a new starch branching enzyme type II from wheat. The invention also relates to a novel method for identification of such branching enzyme type II proteins, which is useful for screening wheat germplasm for null or altered alleles of wheat branching enzyme IIb. The novel gene, protein and methods of the invention are useful in production of wheat plants which produce grain with novel properties for food and industrial applications, for example wheat grain containing high amylose or low amylopectin starch.

15 Background of the Invention

In plants, two classes of genes encode starch branching enzymes, known respectively as BEI, and BEII. In the monocotyledonous cereals, there is strong evidence demonstrating that the BEII class contains two independent types of genes, known in maize as BEIIa and BEIIb (Gao et al., 1996; Fisher et al., 1996). In barley, two types of genes have been reported, and shown to be differentially expressed (Sun et al., 1998). An additional class of branching enzyme (50/51 kD) from barley has also been described (Sun et al., 1996).

In dicotyledonous plants, loss of BEII activity through either mutation (Bhattacharyya et al., 1990) or gene suppression technologies gives rise to starches containing high amylose levels (Safford, 1998, Jobling 1999).

In monocotyledonous plants, mutations giving rise to high amylose contents are known in maize, rice and barley. In neither rice (Mizuno et al., 1993) nor barley (Schondelmaier et al., 1992) have the known high amylose phenotypes been associated with the BEIIa or BEIIb mutations respectively. However, in maize it is firmly

established that the high amylose phenotype is associated with down regulation of the BEIIb gene (Boyer et al., 1980; Boyer and Preiss, 1981, Fisher et al, 1996).

5 The impact of down-regulation of BEI has been investigated through antisense inhibition in potato tuber; the down-regulation has been found to alter the properties of the starch, but not its gross structural features, such as the amylose content (Filipse et al., 1996). In wheat, antisense down-regulation of BEI activity has small but
10 significant effects on starch structure (Baga et al, 1999). The branching enzyme I gene from maize has been cloned (Kim et al., 1998), but mutants affecting branching enzyme I activity in maize are not known.

No mutations specifically reducing BEIIa activity have
15 been reported, and no gene suppression experiments in plants have succeeded in reducing BEIIa activity, although the *dul* mutation in maize is known to reduce the expression of both BEIIa and starch synthase III. However, the *dul* mutation is now known to be due to mutation of the
20 structural gene for starch synthase III (Gao 1998, Cao 1999).

In our previous patent application No. PCT/AU98/00743 (WO99/14314), we have described the structure of a BEII gene from wheat, which we have subsequently designated the
25 BEIIa gene.

In the present application we describe the isolation of a second BEII gene from wheat, which we have designated the BEIIb gene, and discuss the uses to which this gene sequence can be applied. We have surprisingly found that
30 in wheat the expression level of the various branching enzymes is very different to that in maize and barley. In this specification we show that BEIIb in wheat is expressed at low levels in the soluble fraction of the wheat endosperm, and is predominantly found within the starch
35 granule. This indicates that there are important differences in the regulation of gene expression in wheat compared to other cereals, suggesting that the manipulation

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of the amylose to amylopectin ratio in wheat will involve the manipulation of more than just the BEIIb gene.

We have also surprisingly found that the BEIIa and BEIIb gene structures are highly conserved with respect to
5 exon size and position, allowing us to prepare DNA-based diagnostics which they can distinguish not only the BEIIa and BEIIb classes of genes, but also the forms of these genes encoded on the A, B and D genomes of wheat, and to
10 identify the BEIIb proteins expressed by the wheat A, B and D genomes, providing an essential tool for the screening of wheat germplasm for null or altered alleles of wheat branching enzyme IIa.

Summary of the Invention

15 In a first aspect, the invention provides an isolated nucleic acid molecule encoding wheat starch branching enzyme IIb (BEIIb).

Preferably the nucleic acid sequence is a DNA sequence, and may be genomic DNA or cDNA.

20 Preferably the nucleic acid molecule has the sequence depicted in Figure 8 (SEQ ID NO:5), Figure 9 (SEQ ID NO:6), or SEQ ID NO:10. It will be clearly understood that the invention also encompasses nucleic acid molecules capable of hybridising to these sequences under at least low
25 stringency hybridization conditions, or a nucleic acid molecule with at least 70% sequence identity to at least one of these sequences. Methods for assessing ability to hybridize and % sequence identity are well known in the art. Even more preferably the nucleic acid molecule is
30 capable of hybridizing thereto under high stringency conditions, or has at least 80%, most preferably at least 90% sequence identity. A nucleic acid molecule having at least 70%, preferably at least 90%, more preferably at least 95% sequence identity to one or more of these
35 sequences is also within the scope of the invention.

Biologically-active untranslated control sequences of genomic DNA are also within the scope of the invention.

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Thus the invention also provides the promoter of BEIIb.

In a second aspect of the invention, there is provided a genetic construct comprising a nucleic acid sequence of the invention, a biologically-active fragment thereof, or a
5 fragment thereof encoding a biologically-active fragment of BEIIb operably linked to one or more nucleic acid sequences which are capable of facilitating expression of BEIIb in a plant, preferably a cereal plant. The construct may be a plasmid or a vector, preferably one suitable for use in
10 transformation of a plant. Such a suitable vector is a bacterium of the genus *Agrobacterium*, preferably *Agrobacterium tumefaciens*. Methods of transforming cereal plants using *Agrobacterium tumefaciens* are known; see for example Australian Patent No. 667939 by Japan Tobacco
15 Inc.; Australian Patent No. 687863 by Japan Tobacco Inc.; International Patent Application No. PCT/US97/10621 by Monsanto Company; and Tingay et al (1997).

In a third aspect, the invention provides a genetic construct for targeting of a desired gene to endosperm of a
20 cereal plant, and/or for modulating the time of expression of a desired gene in endosperm of a cereal plant, comprising a BEIIb promoter, operatively linked to a nucleic acid sequence encoding a desired protein, and optionally also operatively linked to one or more
25 additional targeting sequences and/or one or more 3' untranslated sequences.

The nucleic acid encoding the desired protein may be in either the sense orientation or in the anti-sense orientation. Alternatively it may be a duplex construct,
30 comprising a portion of the nucleic acid sequence encoding the desired protein in both the sense and anti-sense orientations, operably linked by a spacer sequence. It is contemplated that any desired protein which is encoded by a gene which is capable of being expressed in the endosperm
35 of a cereal plant is suitable for use in the invention. Preferably the desired protein is an enzyme of the starch biosynthetic pathway. For example, the antisense sequences

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of GBSS, starch debranching enzyme, SBE II, low molecular weight glutenin, or grain softness protein I, may be used. Preferred sequences for use in sense orientation include those of bacterial isoamylase, bacterial glycogen synthase, or wheat high molecular weight glutenin Bx17.

In a fourth aspect, the invention provides a wheat BEIIb polypeptide, comprising an amino acid sequence encoded by a nucleic acid molecule according to the invention, or a polypeptide having at least 70%, more preferably 80%, even more preferably 90% amino acid sequence identity thereto, and having the biological activity of BEIIb.

The polypeptide may be designed on the basis of amino acid sequences deduced from the nucleic acid sequences of the invention, or may be generated by expression of the wheat BEIIb nucleic acid molecule in a heterologous system. Suitable heterologous systems are very well known in the art, and the skilled person will readily be able to select a system suitable for the particular purpose desired.

In a fifth aspect, the invention provides an antibody directed against BEII polypeptide. The antibody may be polyclonal or monoclonal. It will be clearly understood that the invention also encompasses biologically-active antibody fragments, such as Fab, (Fab)₂, and ScFv. Methods for production of antibodies and fragments thereof are very well known in the art.

The antibodies of the invention may be used for identification and separation of BEIIb proteins, for example by affinity electrophoresis. This greatly facilitates the identification and combination of altered forms of BEIIb via analysis of germplasm, and greatly assists plant breeding. The antibodies of the invention are suitable for use in any affinity-based separation system, preferably using methods which overcome interference by amylases. Suitable methods are known in the art.

In a sixth aspect, the invention provides a plant cell

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transformed by a genetic construct according to the invention, or a plant derived from such a cell. Additionally, a transformed plant cell may also comprise one or more null alleles for a gene selected from the group consisting of GBSS, BEIIa, and SSII. Preferably the plant is a cereal plant, more preferably wheat or barley.

In a seventh aspect, the invention provides a method of modifying the characteristics of starch produced by a plant, comprising the steps of:

- a) increasing the level of expression of BEIIb in the plant, for example by introducing a nucleic acid molecule encoding BEIIb into a host plant, or
- b) decreasing the level of expression of BEIIb in the plant, for example by introducing an anti-sense nucleic acid sequence directed to a nucleic acid molecule encoding BEIIb into a host plant.

As is well known in the art, over-expression of a gene can be achieved by introduction of additional copies of the gene, and anti-sense sequences can be used to suppress expression of the protein to which the anti-sense sequence is complementary. Other methods of suppressing expression of genes are known in the art, for example co-suppression, RNA duplex formation, or homologous recombination. It would be evident to the person skilled in the art that sense and anti-sense sequences may be chosen depending on the host plant, so as to effect a variety of different modifications of the characteristics of the starch produced by the plant.

Preferably the plant is a cereal plant, more preferably wheat or barley.

Preferably the branching of the amylopectin component of starch is modified by either of these procedures. More preferably a plant with high amylose content is produced.

In an eighth aspect, the invention provides a method of targeting expression of a desired gene to the endosperm of a cereal plant, comprising the step of transforming the plant with a construct according to the invention.

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In a ninth aspect, the invention provides a method of identifying a null or altered allele encoding an enzyme of the starch biosynthetic pathway, comprising the step of subjecting DNA from a plant suspected to possess such an allele to a DNA fingerprinting or amplification assay, which utilizes at least one DNA probe comprising one or more of the nucleic acid molecules of the invention. The nucleic acid molecule may be a genomic DNA or a cDNA, and may comprise the full-length coding sequence or a fragment thereof. Any suitable method for identification of BEIIb sequences may be used, including but not limited to PCR, rolling circle amplification, RFLP, and AFLP. Such methods are well known in the art, and any suitable technique may be used.

In a tenth aspect, the invention provides a plant comprising one or more BEIIb null alleles, in combination with one or more other null alleles selected from the group consisting of BEIIa, GBSS, SSII and BEI. Optionally the plant may also comprise a BEIIa or BEIIb gene expressed in either the sense or the anti-sense orientation. The null alleles for BEIIa, GBSS SSII and BEI may be identified using methods described in PCT/AU97/00743.

It will clearly understood that the invention also encompasses products produced from plants according to the invention, including but not limited to whole grain, part grain, flour or starch.

Because of the very close relationship between *Aegilops tauschii*, formerly known as *Triticum tauschii*, and wheat, as discussed in PCT/AU97/00743, results obtained with *A. tauschii* can be directly applied to wheat with little if any modification. Such modification as may be required represents routine trial and error experimentation. Sequences from these genes can be used as probes to identify null or altered alleles in wheat, which can then be used in plant breeding programmes to provide modifications of starch characteristics. The novel sequences of the invention can be used in genetic

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engineering strategies or to introduce a desired gene into a host plant, or to provide anti-sense sequences for suppression of expression of the BEIIb gene in a host plant, in order to modify the characteristics of starch
5 produced by the plant.

While the invention is described in detail in relation to wheat, it will be clearly understood that it is also applicable to other cereal plants of the family Gramineae, such as maize, barley and rice.

10 Methods for transformation of monocotyledonous plants such as wheat, maize, barley and rice and for regeneration of plants from protoplasts or immature plant embryos are well known in the art. See for example Lazzeri et al, 1991; Jahne et al, 1991 and Wan and Lemaux, 1994 for
15 barley; Wirtzens et al, 1997; Tingay et al, 1997; Canadian Patent Application No. 2092588 by Nehra; Australian Patent Application No. 61781/94 by National Research Council of Canada, and Australian Patents No. 667939 and No. 687863 by Japan Tobacco Co.

20 The sequences of ADP glucose pyrophosphorylase from barley (Australian Patent Application No. 65392/94), starch debranching enzyme and its promoter from rice (Japanese Patent Publication No. Kokai 6261787 and
25 Japanese Patent Publication No. Kokai 5317057), and starch debranching enzyme from spinach and potato (Australian Patent Application No. 44333/96) are all known.

Brief Description of the Figures

Figure 1 shows the sequence of the SBE9 branching
30 enzyme cDNA encodes SBE IIa, cloned from a wheat cv Rosella cDNA library (SEQ ID NO:1).

Figure 2 shows the sequence of the branching enzyme BEIIa gene (SEQ ID NO:2) contained within the F2 lambda clone isolated from an *Aegilops tauschii* genomic DNA
35 library.

Figure 3 shows the results of hybridisation of *Aegilops tauschii* DNA with probes derived from wSBE II-DA1

type sequences. A. Hybridisation with a probe from SBE9 consisting of exons 5-9. B. Hybridisation with fragment F2.2 (consisting of exons 4-9 and introns 4-8 and part of introns 3 and 9). Enzymes used for the digest were:

- 5 1. *Bam* HI, 2. *Dra* I, 3. *EcoR* I, 4. *EcoR* V. Molecular size markers are indicated.

Figure 4 shows the alignment of sequences of Intron 5 fragments from the A, B and D genomes of wheat

Figure 5 shows the PCR analysis of *A. tauschii* genomic clones using Intron V sequences.

Figure 6 shows the alignment of a 262bp PCR fragment amplified from hexaploid wheat using the primers sr913F and WBE2E6R, and a region from the wheat branching enzyme IIb gene *WSBE II-DB1*.

15 Figure 7 shows the alignment of barley branching enzyme IIb cDNA, wheat branching enzyme IIb cDNA, and SBE9 sequences with the sequence of the wheat (*A. tauschii*) branching enzyme IIb gene.

Figure 8 shows the partial genomic sequence of a branching enzyme IIb gene from *A. tauschii* (SEQ ID NO:5).

Figure 9 shows the sequence of a cDNA for branching enzyme IIb gene from hexaploid wheat (SEQ ID NO:6).

Figure 10 shows the sequence alignment of branching enzyme genes. The cDNA sequences used for this analysis were SBE9 (SEQ ID NO:1; Figure 1), wheat BEIIb cDNA (SEQ ID NO:6; Figure 9), Y11282, a wheat branching enzyme sequence (Nair et al. 1997), barley BEIIa (Sun et al. 1998), barley BEIIb (Sun et al. 1998), rice BEIII (Mizuno et al. 1993), rice BEIV (Genbank Accession No. E14723) maize BEIIa (Gao et al. 1997) and maize BEIIb (Gao et al. 1997). The observed N-terminal of wheat (Morell et al., 1997; Y11282) is shown in bold. Figure 11 shows the dendrogram of BE sequences. The sequences analysed were for wheat Y11282 (Nair et al., 1997), SBE 9 (SEQ ID NO:1; (Figure 1), wheat BEIIb (SEQ ID NO:9; Figure 9), barley IIa and IIb (Sun et al. 1998), maize IIa (Gao et al. 1997), maize IIb (Fisher et al. 1993), rice III (Mizuno et al. 1993), rice IV

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(Genbank accession E14723), potato BEI (Khoshnoodi et al. 1997), potato BE II (Cangiano et al 1993), pea BEI and BEII (Burton et al. 1995), E.coli BE (Baecker et al. 1986) and bacillus (Kiel et al 1992). Note that pea BE I and pea BE II sequences correspond to maize BE II and BE I respectively because of differences in nomenclature conventions.

Figure 12 shows the comparison of exon/intron structure for the BEIIa and BEIIb genes. (1) wheat branching enzyme IIa gene, wSBE II DA1 (2) maize amylose extender BEIIb gene (3) partial wheat branching enzyme IIb gene, wSBE II DB1 (4) partial barley branching enzyme IIb gene.

Figure 13 shows the results of analysis of the expression of mRNA for the BEIIa and BEIIb genes in wheat. Panel (A): Hybridisation of SBE9 probe to lanes 1 to 3 and hybridisation of wheat BEIIb cDNA probe to lanes 4 to 6. Panel (B): mRNA loading for each lane.

Lanes 1 and 4 contain leaf mRNA; lanes 2 and 5 contain pre-anthesis floret mRNA; lanes 3 and 6 contain mRNA from wheat endosperm collected 15 days after anthesis.

Figure 14 shows the results of analysis of wheat endosperm branching enzyme IIa by affinity electrophoresis.

Samples: Lanes 1, 4 and 7 contained 20 µg endosperm soluble protein, lanes 2, 5 and 8 contained 30 µg endosperm soluble protein and lanes 3 and 6 contained 10 µg endosperm soluble protein.

Figure 15 shows the results of non-denaturing gel electrophoresis analysis of branching enzymes in the soluble fraction of wheat endosperm.

Samples were: Lane 1, R6 pre-immune, 1:100; Lane 2, R6 pre-immune, 1:3000; Lane 3, R6, 1:100; Lane 4, R6, 1:1000; Lane 5, R6, 1:3000; Lane 6, 3KLH, 1:2000; Lane 7, 3KLH, 1:5000; Lane 8, R7 pre-immune, 1:1000; Lane 9, R7 pre-immune 1:5000; Lane 10, R7, 1:1000; Lane 11, R7, 1:3000; Lane 12, R7, 1:5000

Figure 16 shows the results of affinity

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electrophoresis separation of branching enzyme IIa forms from diverse wheat germplasm using the gel conditions described in Figure 11 (Panel C). Panel A. Lane 1, Durati, *T. durum*; Lane 2 *A. tauschii*, Accession No. 24242; Lane 3, *A. tauschii*, Accession No. 24095; Lane 4, *A. tauschii*, Accession No. 24092; Lane 5, Hartog, *Triticum aestivum*; Lane 6, Rosella, *T. aestivum*; Lane 7, Corrigin, *T. aestivum*; Lane 8, Bodallin, *T. aestivum*; Lane 9, Beulah, *T. aestivum*; Lane 10 Bindawarra, *T. aestivum*; Lane 11, Barley (*Hordeum vulgare*). Panel B. Lane 1: Afghanistan 006, *Triticum durum*; Lane 2, Persia 20, *T. aestivum*; Lane 3, Afghanistan 8, *T. aestivum*; Lane 4, Kashmir 4, *T. aestivum*; Lane 5, Gandum Sockhak, *T. aestivum*; Lane 6, Warbler, *T. aestivum*; Lane 7, Bayles, *T. aestivum*; Lane 8, Kometa; Lane 9, Kashmir 14, *T. aestivum*; Lane 10, Rosella, *T. aestivum*; Lane 11, Kashmir 8, *T. aestivum*; Lane 12, Beijing 10, *T. aestivum*; Lane 13, Savannah, *T. aestivum*; Lane 14, Afghanistan 55-623, *T. aestivum*; Lane 15, Karizik, *T. aestivum*; Lane 16, Indore E98, *T. durum*; Lane 17, Iraq 17, *T. durum*; Lane 18, Seri 82, *T. aestivum*; Lane 19, Indore 19, *T. aestivum*.

Figure 17 shows the results of two-dimensional separation of the components of the wheat starch granule 88 kD band. The wheat starch granule 88 kDa band was electrophoresed in the first dimension through an SDS-PAGE gel. Lanes were excised, renatured, and placed on top of a non-denaturing PAGE gel and electrophoresed in a second dimension. Two lanes were placed on top of each non-denaturing PAGE gel. (A) protein staining with Coomassie blue reagent (B) Immunoblotting of gels with either 3KLH or R6 antibodies, as indicated on the figure.

Figure 18 is a diagrammatic representation of the BEII genes from various species, showing the exon/intron structure. The dark rectangles represent exons.

Figure 19 shows the results of PCR amplification of SBE IIb gene from CS nullisomic lines, using the primers ARA 12F and ARA 10R.

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Figure 20 shows the results of PCR amplification of SBE IIb gene, using the primers ARA 6F and ARA 8R from *Triticum* spp. Lanes: 1) *T. monococcum*, 2) *T. durum*, 3) *T. urartu*, 4) *T. tauschii*, 5) CSDT2DS, 6) CSDT2BL-9, 7) CSDT2AS and 8) CS.

Figure 21 shows the alignment of the exon 1 - intron 1 - exon 2 region of the SBE IIb gene from the A, B and D genomes. * indicates that the sequence could not be specifically assigned to the A or B genome.

Figure 22 shows the alignment of the BEIIb sequences from each genome.

Figure 23 shows the results of PCR amplification of the SBE IIb gene was carried out using the primers ARA 19F and ARA 15R, followed by restriction digestion using *Rsa*I. Lanes 1) CS, 2) *T. monococcum*, 3) *T. tauschii*, 4) CSDT2BL-9, which is missing part of the long arm of chromosome 2B, and 6) CSDT2AS, which is missing the long of chromosome 2A.

Figure 24 shows the results of PCR amplification of intron 3 region of SBE IIb from wheat lines, using the primers ARA 19F and ARA 23R followed by *Rsa* I digestion. Lane 12 is the null mutant for the D genome

Figure 25 is a schematic representation showing the development of the SBE IIa construct. A) Biogemma vector, pDV03000; B) pBluescript carrying the full length cDNA of SBE IIa; C) SBE IIa construct in pDV03000; D) Sense IIa construct and E) Antisense IIa construct.

Figure 26 is a schematic representation of the development of the SBE IIb construct. A) Biogemma vector, pDV03000; B) pGEM-T carrying a 1046bp fragment of SBE IIb; C) SBE IIb construct in pDV03000; D) Sense IIb construct and E) Antisense IIb construct.

Figure 27 is a schematic representation of a SBE II duplex construct. A) SBE sequence inserted in between the promoter and the terminator in its linear form; B) Duplex formation of mRNA within the transgenic plant.

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Example 1 *Isolation of BEII genes from an A. tauschii genomic library and their characterisation by PCR*

Plant material

5 *Aegilops tauschii*, CPI 110799, was used for the construction of the genomic library. Previously this accession has been shown to be most like the ancestral D genome donor of wheat, on the basis of the conservation of order of genetic markers (Lagudah et al. 1991). The
10 *Triticum aestivum* cultivars Rosella, Wyuna and Chinese Spring were used for the construction of different cDNA libraries.

cDNA and genomic libraries

15 The construction of the cDNA and genomic libraries used in this example was as described in Rahman et al., (1997,1999) and in Li et al. (1999). Conditions for library screening were hybridisation at 25% formamide, 5XSSC, 0.1% SDS, 10X Denhardts, 100µg/ml salmon sperm DNA
20 at 42°C for 16h, followed by washing at 2XSSC, 0.1% SDS at 65°C for 3X1h.

Screening of a wheat cDNA library

 Screening of a wheat cv Rosella cDNA library prepared
25 from endosperm (mid-stage of development) with the maize SBE I clone (Baba et al., 1991) at low hybridisation stringency led to the isolation of two classes of positive plaques. One class hybridised strongly to the probe, and encoded wheat SBE I (Rahman et al., 1997,1999). The second
30 class was weakly hybridising. The clone with the longest insert from this second class was called SBE 9, and its sequence showed greater identity to SBE II than to SBE I type sequences. This was designated SBE IIa. The sequence of SBE 9 (SEQ ID NO:1) is set out in Figure 1.

35

Screening of *A. tauschii* genomic library

A genomic library constructed from *A. tauschii* was screened by DNA hybridisation with SBE9, and four positive clones were purified. These were designated F1 to F4. The
5 sequence from positions 537 to 890 of SBE9 was amplified by PCR, and used to screen the *A. tauschii* library again. Clones isolated from this screening are referred to as G1 and G2 and H1 to H8

10 (1) Number of *BEII* type genes in wheat

The sequence of a branching enzyme gene, designated F2, from *Aegilops tauschii* was described in WO99/14314, and is given in Figure 2 (SEQ ID NO:2). A probe generated from F2, designated F2.2, contained sequences from 2704 to
15 4456 bp of SEQ ID NO:2, and contained exons 4-9, introns 4-8, and parts of intron 3 and 9. Hybridisation of *A. tauschii* DNA (cut with four different restriction enzymes) with F2.2 revealed only one strongly hybridising band and several very faint bands (Figure 3, panel B), consistent
20 with the presence of a single *BEII* type gene in the *A. tauschii* genome. The cDNA clone, SBE9 (SEQ ID NO:1) has >95% identity to the exon regions of the F2 branching enzyme gene. A region of SBE9 from nucleotides 537 to 890, including exons 5 to 9, was used as a hybridisation probe,
25 and gave a much more complex pattern (Figure 3, panel A), strongly indicating that there is more than one *BEII* gene type in the *A. tauschii* genome.

Example 2:

PCR analysis of BEIIa - Intron 5

30 PCR primers, sr913F (5' ATC ACT TAC CGA GAA TGG G 3', SEQ ID NO:3) and WBE2E6R (5' CTG CAT TTG GAT TTC AAT TG 3', SEQ ID NO:4) were designed to anneal to Exon 5 and Exon 6 respectively of the wheat F2 gene in order to amplify the intron region (Intron 5) between these exons. Analysis of
35 the products of PCR reactions using these primers shows that the primers amplify fragments of 228 bp from the A-genome of wheat, 226 bp from the D genome and 217 bp from

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the B genome. These fragments were shown to be amplified from chromosome 2A, 2D and 2B of wheat respectively by analysis of nullisomic/tetrasomic chromosome-engineered lines of wheat. In addition to these fragments, a 262 bp fragment was amplified, and this fragment (designated the 262 bp Universal fragment) was not polymorphic among the chromosome engineered lines tested. The 262 bp Universal fragment and the A, B and D regions from the F2 gene were cloned and sequenced, and the sequence comparison is shown in Figure 4.

Example 3: *Classification of the G1-G2 and H1-H10 genes*

PCR analysis using PCR primers sr913F (5' ATC ACT TAC CGA GAA TGG G 3') and WBE2E6R (5' CTG CAT TTG GAT TTC AAT TG 3') showed that the H1 to H10 lambda clones yielded an approximately 200 bp fragment, and the G1 and G2 clones yielded an approximately 260 bp fragment (Figure 5). Partial sequencing of G1 and G2 showed that the parts of the sequence analysed had 80% identity with the exons 4 and 5 of *WSBE II-DA1*, but the intervening intron contained a sequence that showed no homology to any sequence contained within F2.

However, the G1 and G2 clones from *A. tauschii* showed 92.7% identity to the sequence of the 262 bp universal fragment amplified and cloned from hexaploid wheat, and an alignment of these sequences is shown in Figure 6. Figure 7 shows an alignment of a region corresponding to the 537 to 890 bp region of the SBE9 clone from the cDNAs for barley BEIIb (Sun et al., 1995, Sun et al., 1998), SBE9, wheat BEIIb cDNA with the sequence from clone G1. Further sequencing of G1 led to the isolation of a sequence, shown in Figure 8 (SEQ ID NO:5), which showed high identity with the sequence reported by Sun et al. (1998) for the 5' end of barley IIb cDNA and the partial sequence for the cognate gene. G1 and G2 therefore contain a gene which is distinct from F2, and which has high homology to barley BEIIb. We have designated this gene *WSBE II-DB1*.

Example 4: *Isolation of a wheat BEIIb cDNA and an additional genomic fragment*

A barley cDNA library was constructed using 5 µg of
5 polyA⁺ mRNA (1.67 µg of polyA⁺ mRNA from 10, 12 and 15 DPA
endosperm tissues were pooled). cDNA was synthesised using
the cDNA synthesis system marketed by Life Technology,
except that the NotI-(dT)₁₈ primer (Pharmacia Biotech) was
used to synthesise the first strand of cDNA. Pfu polymerase
10 was added to the reaction after second strand synthesis to
flush the ends of cDNAs. SalI-XhoI adapter (Stratagene) was
then added to the cDNAs. cDNAs were ligated to SalI-NotI
arms of λZipLox (Life Technology) after digestion of cDNAs
with NotI followed by size fractionation (SizeSep 400 spun
15 Column of Pharmacia Biotech). The entire ligation reaction
(5 µl) was packaged using Gigapack III Gold packaging
extract (Stratagene). The titre of the library was tested
by transfecting either the Y1090(ZL) or the LE392 strain of
E.coli.

20 Primers 1 and 2 (Sun et al. 1998)), were used for PCR
amplification of a fragment from a barley cDNA library (Ali
et al., 2000) using conditions described in Sun et al.
(1998). The identity of this fragment was confirmed by
sequence analysis, and the fragment was used as a probe to
25 isolate a cDNA by hybridisation, cDNA from a cDNA library
constructed from Chinese Spring (Li et al. 1999).

This cDNA was designed wBEIIb, and its sequence is
shown in Figure 9 (SEQ ID NO:6). This probe was also used
to reprobe the genomic library from *A.tauschii* referred to
30 above, and a clone, designated G5, was recovered from this
screen. Analysis showed that the wBEIIb cDNA sequence
showed 98.5% identity and the G5 sequence showed 100%
identity to sequences already recovered from G1 and G2. G5
therefore represented the same wSBE II-DB1 gene, and the
35 wBEIIb cDNA is a product of the orthologous gene in
hexaploid wheat.

Example 5: *Relationships between BEII sequences*

Deduced amino acid sequences for branching enzymes from various cereals were analysed using the PILEUP program from the GCG suite of programs (Devereux 1984), and an alignment of these sequences is shown in Figure 10. The PILEUP analysis used a penalty of 12 for insertion of a gap and 0.1 for extending the gap per residue. The cDNA sequences used for this analysis were SBE9 (SEQ ID NO:1; Figure 1), wheat BEIIb cDNA (SEQ ID NO:6; Figure 9), Y11282, a wheat branching enzyme sequence (Nair et al. 1997), barley BEIIa (Sun et al. 1998), barley BEIIb (Sun et al. 1998), rice BEIII (Mizuno et al. 1993), rice BEIV (Genbank Accession No. E14723) maize BEIIa (Gao et al. 1997) and maize BEIIb (Fisher et al., 1993). The observed N-terminal of wheat (Morell et al., 1997; Y11282) is shown in bold.

The relationships between branching enzyme sequences are illustrated in Figure 11, using a dendrogram generated by the PILEUP program. The sequences analysed were for wheat Y11282 (Nair et al., 1997), SBE 9 (Figure 1), wheat BEIIb (Figure 9), barley IIa and IIb (Sun et al. 1998), maize BEI (Kim et al, 1998), maize IIa (Gao et al. 1997), maize IIb (Fisher et al. 1993), Arabidopsis BEII (U22428, Fisher et al., 1996), Arabidopsis BEII (U18817, Fisher et al., 1996), rice I (Kawasaki et al., 1993), rice III (Mizuno et al. 1993), rice IV (Genbank accession E14723), potato BEI (Khoshnoodi et al. 1997), potato BE II (Cangiano et al 1993), pea BEI and BEII (Burton et al. 1995), E. coli BE (Baecker et al. 1986) and bacillus (Kiel et al 1992). Note that pea BE I and pea BE II sequences correspond to maize BE II and BE I respectively because of differences in nomenclature conventions.

On the basis of this comparison, the branching enzyme gene contained on clone F2 was classified as a BEIIa type gene and designated *WSBE II-DA1*.

Example 6: *Structure of the wSBE II-DA1 and wSBE II-DB1 genes*

Figure 12 shows a comparison of the exon/intron structures of the wheat wSBE II-DA1 and wSBE II-DB1 genes. The structure of the wSBE II-DB1 gene is shown from the beginning of the wheat BEIIb cDNA through to exon 5. Hybridisation results suggest that regions at the 3' end of the wheat BEIIb cDNA are not contained within any of the clones G1, G2 and G5. This is not surprising, as the maize SBE II b gene extends over 16.5kb and required the isolation of two genomic clones (Kim et al 1998). The positions of the intron/exon boundaries for the first five introns of the wheat BEIIa and BEIIb genes are conserved, as shown in Table 1. The size of the first five introns in wSBE II-DB1 vary considerably in size from the first five introns in wSBE II-DA1.

Table 1
Exon/Intron Structures of Cereal branching Enzyme Genes

Exons					Introns				
	Wheat WSBE DA1	Maize BEIIB	Wheat WSBE II-DB1	Barley BEIIB		Wheat WSBE II-DA1	Maize BEIIB	Wheat WSBE II-DB1	Barley BEIIB
1	123 ^a	112 ^a	148 ^a	121 ^a	1	327	106	148	105
2	98	146	146	152	2	276	244	663	2064
3	242	155	230	230	3	401	1086	465	388
4	99	99	99	99	4	169	76	74	74
5	43	43	43	43 ^b	5	152	196	181	
6	60	60	60		6	335	499	442	
7	81	81	81		7	83	81	79	
8	117	117	117		8	288	567	178	
9	81	84	84		9	629	775		
10	122	122			10	175	751		
11	120	120			11	974	4020		
12	130	130			12	88	86		
13	111	111			13	201	148		
14	129	129			14	579	3051		
15	104	104			15	841	872		
16	145	145			16	1019	457		
17	148	148			17	135	144		
18	105	101			18	176	226		
19	74	78			19	201	266		
20	156	156			20	377	448		
21	75	75			21	89	96		
22	384	84							

^a. Exon 1 numbering begins from ATG of translation start codon

^b. Partial sequence for exon or intron

Example 7:*Expression analysis at the mRNA level*

RNA from endosperm at different developmental stages was obtained from wheat grown in the glasshouse as described in Li et al. (1999). RNA was extracted by the method of Higgins et al. (1976), separated on denaturing formamide gels and blotted onto Hybond N+ paper, essentially as described in Maniatis et al. (1992). Probes were prepared from the extreme 3' ends of SBE9 (bases 2450 to 2640 of SEQ ID NO:1) and wBEIIb cDNA (bases 2700 to 2890 of SEQ ID NO:6) by PCR using the following scheme: 94°C, 2min, 1 cycle, 94°C, 30s, 55°C, 30s, 72°C, 30s, 36 cycles, 72°C 5min, 1cycle, 25°C, 1min, 1cycle. The probes were from the 3' untranslated region, and were specific for either wSBE II-DA1 or wSBE II-DB1 type sequences. An RNA species of about 2.9kb hybridised to each probe (Figure 13 Panel B). However, the intensity of hybridisation determined by densitometry, and normalised for differences in RNA loading), indicated that RNA hybridising to the wSBE II-DB1 gene was present at 2.5 to 3 fold lower concentration than RNA hybridising to the wSBE II-DA1 gene.

Example 8:*Analysis of branching enzymes by affinity electrophoresis demonstrates that only BEIIa is predominant in the soluble fraction*

In Morell et al., (1997), we reported that only a single form of branching enzyme II could be identified in the wheat developing endosperm soluble fraction. However, this was on the basis of anion-exchange chromatography, and it remained possible that there were multiple forms, even though they could not be separated by this technique. Matsumoto has developed an affinity electrophoresis method for measuring the interaction of branching enzymes with polysaccharide substrates (Matsumoto et al., 1990), and we have further developed this technique specifically to allow the separation of the branching enzyme IIa forms encoded by each of the three wheat genomes. Figure 14 shows an

immunoblot of a non-denaturing polyacrylamide gel electrophoresis experiment in which the gel matrix contained the β -limit dextrin of maize amylopectin alone (Figure 14, lanes 1 and 2), showing separation of three forms of branching enzyme IIa. Resolution is slightly enhanced by the addition of the α -amylase inhibitor acarbose (Figure 14, lanes 3, 4 and 5), and substantially enhanced by the addition of α -cyclodextrin (Figure 14 lanes 6, 7 and 8).

A non-denaturing gel was prepared, containing a stacking gel composed of 0.125 M Tris-HCl buffer (pH 6.8), 6% acrylamide, 0.06% ammonium persulphate and 0.1% TEMED. The separating gel was composed of three panels. The basic non-denaturing gel mix contained 0.34 M Tris-HCl buffer (pH 8.8), CHAPS (0.05%), glycerol (10.3%), acrylamide (6.2%), 0.06% ammonium persulphate, 0.1% TEMED and the β -limit dextrin of maize amylopectin (0.155%). Panel A (lanes 1 and 2) contained only the basic non-denaturing gel reagents. Panel B (Lanes 3, 4 and 5) contained the basic non-denaturing gel reagents and 0.066 mM acarbose. Panel C (lanes 6, 7 and 8) contained the basic non-denaturing gel reagents and 0.067 mM α -cyclodextrin.

Following electrophoresis at 100 V for 16 hours at 4 °C, the proteins in the separating gel were transferred to nitrocellulose membrane according to Morell et al (1997) and immunoreacted with 1:5000 dilution of 3KLH antibodies (raised against the synthetic peptide AASPGKVLVPDESDDLGC (SEQ ID NO:7) coupled to the keyhole limpet hemocyanin via the heterobifunctional reagent m-Maleimidobenzoyl-N-hydroxysuccinimide ester).

The use of a β -limit dextrin provides a superior separation because it prevents interference with the separation by endogenous β -amylases in the wheat endosperm tissue, and the use of α -cyclodextrin in the assay further enhances the separation. Without wishing to limit the invention by any proposed mechanism, we believe that this enhancement may result from the inhibition of endogenous

wheat endosperm α -amylases.

The analysis shows that three branching enzyme II proteins are present, and that each of these proteins cross-reacts with antibodies to a synthetic oligopeptide designed from the N-terminal region of the BEIIa protein in a region that shares no homology with the wheat BEIIb protein.

The soluble fraction of the wheat endosperm was reacted with various antibodies raised against peptides designed on the basis of the sequences of the wheat BEIIa (see Figure 12) or the wheat BEIIb cDNA. Figure 15 shows that only 3KLH, raised against the N-terminus of BEIIa, cross-reacted with proteins (marked by arrows) in the soluble fraction which show a specific shift in mobility in the presence of the β -limit dextrin of amylopectin and α -cyclodextrin. Gels were prepared as described in Figure 14, except that the gel used in Panel A contained the non-denaturing gel mix without the β -limit dextrin of maize amylopectin. Panel B contained the non-denaturing gel mix plus α -cyclodextrin. An extract of developing wheat endosperm was prepared using 3 volumes of extraction buffer per g of tissue, and 140 μ l of sample applied per gel. Following electrophoresis at 100 V for 16 hours at 4 °C, the proteins in the separating gel were transferred to nitrocellulose membrane according to Morell et al (1997) which was cut into 1 cm strips. The antibodies prepared were 3KLH (see Figure 11), R6 (raised in rabbit against the synthetic peptide AGGPSGEVMIGC (SEQ ID NO:8) coupled to the keyhole limpet hemocyanin via the heterobifunctional reagent m-Maleimidobenzoyl-N-hydroxysuccinimide ester); pre-immune serum from the R6 rabbit; R7 (raised in rabbit against the synthetic peptide GGTPPSIDGPVQDSDGC (SEQ ID NO:9) coupled to the keyhole limpet hemocyanin via the heterobifunctional reagent m-maleimidobenzoyl-N-hydroxysuccinimide ester) and pre-immune serum from the R7 rabbit.

As in Figure 14, the BEIIa protein is separated into three forms (indicated by arrows in Figure 15, Panel B), by affinity electrophoresis in the presence of β -limit dextrin. In barley (Sun et al., 1997) and maize (Bayer and Preiss 1981) both branching enzymes IIa and IIb are present in the soluble fraction. In some subsequent experiments we have detected low levels of BE IIb in the soluble fraction.

The separation of the forms of BEIIa encoded by each wheat genome is demonstrated in Figure 16. In Panel (A) the diploid *A. tauschii* (lanes 2,3 and 4) and barley line (lane 11) yields a single band. However, the tetraploid *T. durum* lines (Panel A lane 1, Panel B, lanes 1, 16, and 17) and hexaploid *T. aestivum* lines (Panel A lanes 5-10, Panel B lanes 2-15, 18-19) give at least 2 bands. Some hexaploid lines (panel A, lane 7 and 9, Panel B lanes 2-6, lanes 8-9, lane 13) yield 2 bands, indicating either that they are null for one genome or that the products of two genomes migrate with identical mobility in the gel system.

The use of the separation system as a means of screening for wheat genomes with altered or null alleles of branching enzyme IIa is demonstrated by Figure 14 (Panel B), where different lines are shown to have different numbers and mobilities of branching enzyme IIa proteins.

Example 9: Presence of two classes of proteins in the starch granule at 88 kDa and their differential antibody binding.

The wheat starch granule contains a number of proteins that have been analysed by SDS-PAGE (Rahman et al., 1995, Denyer et al., 1995, Takaoka et al., Li et al., 1999a, Li et al., 1999b) and two-dimensional gel electrophoresis (Yamamori and Endo, 1996). The following bands have been identified: 60 kDa, GBSS; 75 kDa, SSI; 100 kDa, 108 kDa and 115 kDa, SSII). An 88 kDa band is also observed, and has been shown to be associated with branching enzyme activity (Denyer et al., 1995) and to react to antibodies to maize BEII (Rahman et al., 1995). This protein band was shown to

contain at least two protein components.

This analysis has been extended by purification and analysis of the individual granule proteins. The granule proteins were isolated from 4.7g of wheat starch granules by boiling in 24 ml of SDS buffer (50 mM Tris-HCl buffer pH 6.8, 10% SDS and 6.25% 2-mercaptoethanol) as described by Rahman et al., (1995). Residual granular starch was removed by centrifugation, and granule proteins were separated by applying the supernatant to a 9% SDS-PAGE gel prepared in a Biorad Model 491 Prep Cell apparatus. The SDS gel contained a stacking gel composed of 0.125 M Tris-HCl buffer (pH 6.8), 0.25% SDS, 6% acrylamide, 0.06% ammonium persulphate and 0.1% TEMED and a separating gel containing 0.34 M Tris-HCl buffer (pH 8.8), 0.25% SDS, acrylamide (9 %), 0.06% ammonium persulphate, and 0.1% TEMED. The samples were electrophoresed at 60 mA constant current for 16 hours, and fractions of reactions (5 ml) collected by a pump operating at 0.5 ml/min. Fractions were analysed by SDS-PAGE, and fractions containing an 88 kDa band precipitated by the addition of 3 volumes of acetone. The precipitate from each 5 ml fraction was collected by centrifugation, the sample dissolved in SDS buffer, and electrophoresed through a standard 8% SDS-PAGE gel. The lane was excised from the gel and renatured in 0.04 M Tris for 2 hours. To generate a two-dimensional separation, the gel was then laid across the top of a second non-denaturing PAGE gel and electrophoresed. Proteins were identified by staining with Coomassie blue (a 50:50 mixture of 2.5% Coomassie Blue R-250 and Coomassie Blue G250 solutions).

Figure 17, Panel (A) shows that two proteins were visible after staining, and these were designated 88 kD (U) and 88 kD (L), as indicated by the arrows. Immunoblotting of the two-dimensional gel with peptide antibodies to the N-terminal of BEIIa (3KLH) and to the N-terminus of the wheat BEIIb cDNA sequence (R6; see Figures 12 and 13 for details of the antibodies are set out in Example 8)

indicated preferential binding of the R6 antibody to 88 kD (U) and preferential binding of 3KLH to 88 kD (L) (Figure 17, Panel B), providing a provisional assignment of these proteins as BEIIb and BEIIa respectively.

5 The proteins were further analysed by digestion with trypsin, and the peptides released were identified by MALDI-TOF analysis at the Australian Proteome Analysis Facility, Macquarie University, Sydney. The results of this analysis, shown in Table 2, demonstrated that 88 kD
10 (U) was the product of the wheat BEIIb gene, and that while the assignment of 88 kD (L) was inconclusive, the results were consistent with the protein being a branching enzyme encoded by either SBE9 or the wheat BEIIb cDNA.

Table 2

(a) Comparison of 88 kD (U) and the predicted protein encoded by the wheat BEIIb cDNA.

5

Matches: 6

MOWSE Score: 4.97e+001

Coverage: 8.85%

Matching Peptides:

10

MW	Delta	Start	End	Sequence
755.4766	-0.13	320	325	(K) RPKSLR (I)
1337.7092	0.01	453	463	(R) VFNYGNKEVIR (F)
1337.6728	-0.03	703	713	(R) RFDLGDAEFLR (Y)
1508.7623	-0.12	785	799	(K) VVLDS DAGLFGGFGR (I)
1589.6933	-0.08	731	743	(K) YGFMTSDHQYVSR (K)
1692.7049	-0.17	184	198	(R) SDIDEHEGGMDVFSR (G)
1706.8740	-0.04	340	353	(K) INTYANFRDEVLP (I)

(b) Comparison of 88 kD (L) and the predicted proteins encoded by the wheat BEIIb cDNA and SBE9 cDNA.

15

Matches to wheat BEIIb cDNA

Matches: 8

MOWSE Score: 1.32e+003

Likelihood: 2.053+003

20

Coverage: 11.72%

Matching Peptides:

MW	Delta	Start	End	Sequence
819.4603	11.23	464	470	(R) FLLSNAR (W)
1210.5090	-105.27	444	452	(R) GHHWMWDSR (V)
1337.7092	10.53	453	463	(R) VFNYGNKEVIR (F)
1337.6728	-16.68	703	713	(R) RFDLGDAEFLR (Y)
1508.7623	-44.33	785	799	(K) VVLDS DAGLFGGFGR (I)
1573.7446	-16.81	326	339	(R) IYETHVGMSSPEPK (I)
1589.6933	-23.46	731	743	(K) YGFMTSDHQYVSR (K)
1692.7049	-95.07	184	198	(R) SDIDEHEGGMDVFSR (G)
1706.8740	-15.57	340	353	(K) INTYANFRDEVLP (I)

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Matches to wheat SBE9

Matches: 6

MOWSE Score: 1.04e+001

5 Coverage: 8.63%

Matching Peptides:

MW	Delta	Start	End	Sequence
819.4603	11.23	451	457	(R)FLLSNAR (W)
1210.5090	-105.27	431	439	(R) GHHWMWDSR (V)
1508.7875	-27.64	145	156	(K) IYEIDPTLKDFR (S)
1573.7446	-16.81	313	326	(R) IYESHIGMSSPEPK (I)
1599.7641	-9.93	171	185	(R) AAIDQHEGGLEAFSR (G)
1692.8583	-4.45	327	340	(K) INSYANFRDEVLP (I)

10 Example 10: *Sequencing of the SBE IIb gene*

A partial genomic sequence of the SBEIIb gene was obtained, using clone G5 described in Example 4. So far approximately 8.4kb of sequence has been obtained. This includes approximately 500bp upstream of the start codon, presumably comprising the promoter region, and exons 1 to 14 in full. This partial sequence is set out in SEQ ID NO:10. From the sequences of the corresponding maize and Arabidopsis BEII genes, we would expect the gene to contain 22 exons. A comparison between the exon/intron structures of various BEII genes and the wheat BEIIb gene is shown in Figure 18, and the sizes of the exons in various SBEII genes are compared in Table 3. In this table "Arab" represents Arabidopsis.

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Table 3
 Sizes of exons in various SBE IIb genes

Exon no	Arab21	Arab22	Wheat BEIIa	Maize BEIIb	Barley BEIIb	Wheat BEIIb
1	42	124	279	212	121	148
2	253	120	98	146	152	146
3	236	182	243	155	230	230
4	99	99	99	99	99	99
5	43	43	43	43	43	43
6	60	60	60	60		60
7	81	81	81	81		81
8	117	117	117	117		117
9	84	84	84	84		84
10	122	122	122	122		122
11	120	120	120	120		120
12	130	130	130	130		130
13	111	111	111	111		111
14	129	129	129	129		129
15	104	104	104	104		
16	145	145	145	145		
17		148	148	148		
18		101	101	101		
19		78	78	78		
20		156	156	156		
21		75	75	75		
22		90	384	304		
17	558					
18	164					

5 Using a probe specific for the 3' end of SBE IIb, three clones designated G7, G8 and G9 respectively, have

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now been isolated from the *T. tauschii* genomic library, and are being subjected to sequence analysis to provide the 3' region of the gene.

5 Example 11: *Development of PCR Primer Sets for the Discrimination of the BEIb Genes from each genome*

A number of primer sets, designed on the basis of comparisons between SBE IIa and SBE IIb genes, were tested
10 on wheat genomic DNA. The sequences of these primers were as follows:

ARA 12F:	5' CCG TCC TAC ATG ACA CCT GGC CG 3'	SEQ ID NO:11
ARA 10R:	5' CCG CCG GAT CGA GGA GCC GAC GG 3'	SEQ ID NO:12
ARA 6F:	5' GGC GGC GGC GAC GGG ATG GCT GC 3'	SEQ ID NO:13
15 ARA 8R:	5' CGC CGT CAG GGA TCA TCA CCT CC 3'	SEQ ID NO:14
ARA 19F:	5' CAC CCA TTG TAA TTG GGT ACA CTG 3'	SEQ ID NO:15
ARA 15R	5' TCC ATG CCT CCT TCG TGT TCA TCA 3'	SEQ ID NO:16
ARA 23R	5' CTG CGC ATA AAT CCA AAC TTC TCG 3'	SEQ ID NO:17

20 Targeting the promoter region of SBE IIb using the primers ARA 12F and ARA 13R resulted in the specific amplification of only the D genome gene. Aneuploid analysis using this pair of primers showed that the SBE IIb was located on the long arm of chromosome 2 in wheat, as
25 illustrated in Figure 19.

The primers ARA6F and ARA8R, which amplify the exon 1-intron 1-exon 2 region of SBE IIb, could distinguish the D genome from the A and B genomes, as shown in Figure 20. Sequence analysis of this region indicated that the genes
30 from the A and B genomes completely lack intron 1. This is illustrated in Figure 21.

Example 12: *Identification of SBE IIb in Genomes A, B and D*

35 Sequence analysis of the intron 3 region of SBE IIb, amplified by PCR using the primers ARA 19F and ARA 15R, followed by digestion using the restriction enzyme *Rsa*I,

revealed significant polymorphism amongst the three genomes. This polymorphism, illustrated in the sequence alignment set out in Figure 22, was utilised to develop genome specific markers which can distinguish between the A, B and D genomes.

PCR amplification of the SBE IIb gene was carried out using the primers ARA 19F and ARA 15R, followed by restriction digestion using *Rsa*I. The results of the PCR analysis, shown in Figure 23, indicate that these primers can distinguish between the three genomes.

Screening of approximately 600 wheat lines using the genome specific primer pair, ARA 19F and ARA 23R, which amplifies the same region as ARA 19F and ARA 15R, identified one null mutant of the wheat genome. The amplification was performed as described for Figure 23, and the results are shown in Figure 24.

Example 13: *Constructs for Expression of BEII genes*

Recombinant DNA technology may be used to inhibit or abolish expression of either or both of BE IIa and BE IIb. Three general approaches are used, using transformation of the target plant cells with one of the following types of construct:

a) 'Antisense' constructs of SBE IIa and SBE IIb, in which the desired nucleic acid sequence is inserted into the construct in the opposite direction to the functional gene.

b) 'Sense' constructs of SBE IIa and SBE IIb, in which the desired nucleic acid is inserted in the same direction as the functional gene; this utilises co-suppression events to inhibit the expression of the target gene;

c) Duplex constructs of SBE IIa and SBE IIb, in which the desired nucleic acid in both the sense and antisense orientations is co-located in the construct on either side of a "spacer" loop formed by an intron sequence.

In all three cases, the desired nucleic acid is operably linked to a promoter sequence in the construct.

Sense and antisense constructs have been widely used to modulate gene expression in plants. More recently, it has been shown that the delivery of RNAs with potential to form duplexes is a particularly efficient strategy for generating post-transcriptional gene silencing in transgenic plants (Waterhouse et al., 1998; Smith et al., 2000).

Transformation of the target wheat cells, or cells of other plants, using these constructs is effected using methods known in the art, such as transformation with *Agrobacterium tumefaciens*. Once transgenic plants are obtained, they are assessed for the effects of the transgenes on BE IIa and BE IIb expression. For example, in both maize and potato it has been shown that crossing BE II mutations or BE II transgenes into BE I-deficient backgrounds greatly increases amylose content. Wheat BE I null lines, identified using the methods described in WO99/14314, provide a ready source of BE I-deficient genetic material into which BE IIa and BE IIb transgenics can be crossed, in order to extend further the range of starches which can be produced.

Sense, antisense and duplex constructs of SBE IIa and SBE IIb were generated in the vector pDV03000 (Biogemma Ltd, UK) which carries the high molecular weight gluten promoter (pHMGW) and the Nopaline synthase (Nos) terminator. These constructs are schematically represented in Figures 25, 26 and 27. The Biogemma vectors are based on the well-known plasmid pBR322, and comprise a number of restriction sites, as illustrated in Figures 25 and 26, for incorporation of desired DNA sequences. The entire desired DNA, plus the promoter and terminator sequences referred to above, can then be excised as a *Xho* fragment and cloned into a suitable vector, such as *Agrobacterium tumefaciens*. Those skilled in the art will be aware of other suitable vectors which could be used.

SBE IIa constructs

A sense construct of SB IIa was prepared by inserting a 2143bp fragment of SBE IIa coding sequence in the sense orientation at the *EcoR1/Sma1* site of pDV03000. An SBE IIa antisense construct was prepared by inserting 1913bp of SBE IIa coding sequence in the antisense orientation at the *EcoR1/BamH1* site of pDV03000. This is also illustrated in Figure 25.

SBE IIb constructs

A sense construct of SBE IIb was generated by inserting a 1008bp fragment of the SBE IIb coding sequence in the sense orientation at the *EcoR1/Sma1* site of pDV03000. An antisense SBE IIb construct was prepared by inserting a 955bp sequence of the coding region for SBE IIb at the *BamH1/Pst1* site of pDV03000 in the antisense orientation. This is illustrated in Figure 26.

Duplex constructs

A schematic model of a duplex construct is set out in Figure 27. The duplex construct was prepared using the following protocol, in which all the amplification steps were performed using PCR under conventional conditions.

SBE IIa duplex

- 1) a 468bp sequence of SBE IIa, which includes the whole of exons 1 and 2 and part of exon 3, with *EcoR1* and *Kpn1* restriction sites on either side, was amplified to obtain a first fragment (fragment 1);
- 2) a second fragment, 512bp in length, consisting of part of exons 3 and 4, and the whole of intron 3 of SBE IIa, with *Kpn1* and *Sac1* sites on either side, was amplified to provide fragment 2;
- 3) a 528bp fragment consisting of the complete exons 1, 2 and 3 of SBE IIa, with *BamH1* and *Sac1* sites on either side, was amplified to provide fragment 3;
- 4) fragments 1, 2 and 3 were ligated so that the sequence of fragment 3 was ligated to fragment 2 in the antisense orientation to fragment 1.

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SBE IIb duplex

1) a 471bp sequence consisting of the whole of exons 1 and 2 and part of exon 3 of SBE IIb was amplified with
5 *Eco*R1 and *Kpn*I restriction sites on either side to generate fragment 1;

2) a 589bp fragment consisting of part of exons 3 and 4 and the whole of intron 3 of SBE IIb, with *Kpn*I and *Sac*I sites on either side, was amplified to provide
10 fragment 2;

3) a 528bp fragment consisting of the complete exons 1, 2 and 3, with *Bam*HI and *Sac*I sites on either side was amplified to provide fragment 3;

4) fragments 1, 2 and 3 were ligated so that
15 fragment 3 was in the antisense orientation to fragment 1 when ligated to fragment 2.

The start and end points of the sequences used for making the constructs were as follows:

20 a) *SBE IIa sense construct*

Start: 461bp of Sbe 9 (SBE IIa) cDNA

End: 2603bp of Sbe 9 (SBE IIa) cDNA

25 b) *SBE IIa anti-sense construct*

Start: 691bp of Sbe 9 (SBE IIa) cDNA

End: 2603bp of Sbe 9 (SBE IIa) cDNA

This fragment was ligated in the anti-sense orientation.

30

c) *SBE IIb sense construct*

Start: 85bp of SBE IIb cDNA

End: 1085bp of SBE IIb cDNA

35

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d) *SBE IIb anti-sense construct*

Start: 153bp of SBE IIb cDNA

End: 1085bp of SBE IIb cDNA

5 This fragment was ligated in the anti-sense orientation.

e) *SBE IIa duplex construct*

i) Fragment 1

10 Full exon 1: 1151bp - 1336bp of SBE IIa genomic sequence

Full exon 2: 1664bp - 1761bp of SBE IIa genomic
sequence

Partial exon 3: 2038bp - 2219bp of SBE IIa genomic
sequence

15 This fragment had an *EcoRI* site (GAATTC) introduced at the
start of the exon 1 sequence and a *KpnI* site (GGTACC)
introduced at the end of the partial exon 3 sequence.

ii) Fragment 2

20 Partial exon 3: 2220bp - 2279bp of SBE IIa genomic
sequence

Full intron 3: 2280bp - 2680bp of SBE IIa genomic
sequence

25 Partial exon 4: 2681bp - 2731bp of SBE IIa genomic
sequence

This fragment had a *KpnI* site (GGTACC) introduced at the
start of the partial exon 3 and a *SacI* site (GAGCTC)
introduced at the end of the partial exon 4 sequence.

30 iii) Fragment 3

Full exon 1: 1151bp - 1336bp of SBE IIa genomic
sequence

Full exon 2: 1664bp - 1761bp of SBE IIa genomic
sequence

35 Full exon 3: 2038bp - 2279bp of SBE IIa genomic
sequence

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This fragment had a BamH1 site (GGATCC) introduced at the start of the complete exon 1 sequence and a Sac1 site (GAGCTC) introduced at the end of the complete exon 3 sequence.

5

f) *SBE IIb duplex construct*

i) Fragment 1

Full exon 1: 489bp - 640bp of SBE IIb genomic sequence

10 Full exon 2: 789bp - 934bp of SBE IIb genomic sequence

Partial exon 3: 1598bp - 1770bp of SBE IIb genomic sequence

This fragment had an EcoR1 site (GAATTC) introduced at the start of exon 1 and a Kpn1 site (GGTACC) introduced at the
15 end of the partial exon 3 sequence.

ii) Fragment 2

Partial exon 3: 1771bp - 1827bp of SBE IIb genomic sequence

20 Full intron 3: 1828bp - 2292bp of SBE IIb genomic sequence

Partial exon 4: 2293bp - 2359bp of SBE IIb genomic sequence

This fragment had a Kpn1 site (GGTACC) introduced at the
25 start of the partial exon 3 sequence and a Sac1 site (GAGCTC) introduced at the end of the partial exon 4 sequence.

iii) Fragment 3

30 Full exon 1: 489bp - 640bp of SBE IIb genomic sequence

Full exon 2: 789bp - 934bp of SBE IIb genomic sequence

Full exon 3: 1598bp - 1827bp of SBE IIb genomic sequence

This fragment had a BamH1 site (GGATCC) introduced at the
35 start of exon 1 and a Sac1 site (GAGCTC) introduced at the end of exon 3.

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The SBE IIa and SBE IIb duplexes thus formed were respectively inserted at the *EcoRI/BamHI* site of pDV03000.

Samples of λ phage clones G5 and G9 have been deposited in the Australian Government Analytical
5 Laboratories, acting as an International Depository Authority under the Budapest Treaty on 20 February 2001, under accession numbers NM01/19255 and NM01/19256 respectively.

It will be apparent to the person skilled in the art
10 that while the invention has been described in some detail for the purposes of clarity and understanding, various modifications and alterations to the embodiments and methods described herein may be made without departing from the scope of the inventive concept disclosed in this
15 specification.

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CLAIMS:

1. An isolated nucleic acid molecule encoding wheat starch branching enzyme IIb (BEIIb).
2. An isolated nucleic acid molecule according to claim 5 1, in which the nucleic acid is a DNA.
3. An isolated nucleic acid molecule according to claim 1 or claim 2, in which the nucleic acid is a genomic DNA.
4. An isolated nucleic acid molecule according to claim 10 3, in which the nucleic acid is present in any one of clones G1, G2, G7 to G9, or H1 to H10.
5. An isolated nucleic acid molecule according to claim 1 or claim 2, in which the nucleic acid is a cDNA.
6. An isolated nucleic acid molecule according to claim 5, which has
 - 15 (a) the sequence depicted in any one of Figure 8 (SEQ ID NO 5), Figure 9 (SEQ ID NO 6), or SEQ ID NO 10;
 - (b) a nucleic acid molecule capable of hybridising to at least one of the sequences in (a) under at least low stringency hybridization conditions; or
 - 20 (c) a nucleic acid molecule with at least 70% sequence identity to at least one of the sequences in (a).
7. An isolated nucleic acid molecule according to claim 6, which has
 - (a) the sequence depicted in SEQ ID NO 10;
 - 25 (b) a nucleic acid molecule capable of hybridising to SEQ ID NO:10 under at least low stringency hybridization conditions; or
 - (c) a nucleic acid molecule with at least 70% sequence identity to SEQ ID NO:10.

8. An isolated nucleic acid molecule according to claim 6 or claim 7, in which the nucleic acid molecule is capable of hybridizing to at least one of the sequences in (a) under high stringency conditions, or has at least 80% sequence identity thereto.
9. An isolated nucleic acid molecule according to any one of claims 6 to 8, in which the nucleic acid molecule has at least 90% sequence identity to at least one of the sequences in (a).
10. A promoter sequence of a genomic DNA according to any one of claims 1 to 3.
11. A genetic construct comprising a nucleic acid sequence according to any one of claims 1 to 9, a biologically-active fragment thereof, or a fragment thereof encoding a biologically-active fragment of BEIIb operably linked to one or more nucleic acid sequences which are capable of facilitating expression of BEIIb in a plant.
12. A genetic construct according to claim 11, in which the plant is a cereal plant.
13. A genetic construct according to claim 11 or claim 12, in which the construct is a plasmid or a vector.
14. A genetic construct according to any one of claims 11 to 13, in which the construct is one suitable for use in transformation of a plant.
15. A genetic construct according to claim 13 or claim 14, in which the vector is a bacterium of the genus *Agrobacterium*.
16. A genetic construct according to claim 15, in which the bacterium is *Agrobacterium tumefaciens*.
17. A genetic construct for targeting of a desired gene to endosperm of a cereal plant, and/or for modulating the time of expression of a desired gene in endosperm of a cereal

- plant, comprising a BEIIb promoter, operatively linked to a nucleic acid sequence encoding a desired protein, and optionally also operatively linked to one or more additional targeting sequences and/or one or more 3' untranslated sequences.
18. A genetic construct according to claim 17, in which the desired protein is encoded by a gene which is capable of being expressed in the endosperm of a cereal plant.
19. A genetic construct according to any one of claims 16 to 18, in which the desired protein is an enzyme of the starch biosynthetic pathway.
20. A genetic construct according to any one of claims 16 to 19, in which the nucleic acid encoding the desired protein is in the sense orientation.
21. A genetic construct according to claim 20, in which the sense sequence is selected from the group consisting of bacterial isoamylase, bacterial glycogen synthase, and wheat high molecular weight glutenin Bx17.
22. A genetic construct according to any one of claims 16 to 19, in which the nucleic acid encoding the desired protein is in the anti-sense orientation.
23. A genetic construct according to claim 22, in which the antisense sequence is selected from the group consisting of GBSS, starch debranching enzyme, SBE II, low molecular weight glutenin, and grain softness protein I.
24. A wheat BEIIb polypeptide, comprising an amino acid sequence encoded by a nucleic acid molecule according to any one of claims 1 to 9, or a polypeptide having at least 70% amino acid sequence identity thereto, and having the biological activity of BEIIb.

25. A wheat BEIIb polypeptide according to claim 24, having at least 80% amino acid sequence identity to an amino acid sequence encoded by a nucleic acid molecule according to any one of claims 1 to 9.
- 5 26. A wheat BEIIb polypeptide according to claim 26, having at least 90% amino acid sequence identity to an amino acid sequence encoded by a nucleic acid molecule according to any one of claims 1 to 9.
- 10 27. An antibody directed against a BEII polypeptide according to any one of claims 24 to 26.
28. An antibody according to claim 27, which is polyclonal.
29. An antibody according to claim 27, which is monoclonal.
- 15 30. An antibody according to any one of claims 27 to 29, which is raised against a sequence as set out in SEQ ID NO 7, SEQ ID NO 8, or SEQ ID NO 9.
31. A plant cell transformed by a genetic construct according to any one of claims 11 to 23.
- 20 32. A plant cell according to claim 31, which also comprises a null allele for a gene selected from the group consisting of GBSS, BEIIa, and SSII.
33. A plant derived from a cell according to claim 31 or claim 32.
- 25 34. A plant comprising one or more BEIIb null alleles, in combination with one or more other null alleles selected from the group consisting of BEIIa, GBSS, SSII and BEI, and optionally also comprising a BEIIa or BEIIb gene expressed in either the sense or the anti-sense orientation.
- 30 35. A plant according to claim 33 or claim 34, which is a cereal plant.

36. A plant according to claim 35, which is wheat or barley.
37. A product produced from a plant according to any one of claims 33 to 36.
- 5 38. A product according to claim 37, selected from the group consisting of whole grain, part grain, flour and starch.
39. A product according to claim 37 or claim 38, which is a food.
- 10 40. A food product according to claim 39, selected from the group consisting of unleavened breads, pasta, noodles, breakfast cereals, snack foods, cakes, pastries, and foods containing flour- or starch-based sauces.
- 15 41. A product according to claim 37 or claim 38, which is not a food.
42. A non-food product according to claim 41, selected from the group consisting of films, coating, adhesives, building materials, disposable goods, and packaging materials.
- 20 43. A method of modifying the characteristics of starch produced by a plant, comprising the steps of:
- a) increasing the level of expression of BEIIb in the plant, or
- b) decreasing the level of expression of BEIIb in
- 25 the plant.
44. A method according to claim 43, in which the plant is a cereal plant.
45. A method according to claim 44, in which the plant is wheat or barley.

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46. A method according to any one of claims 43 to 45, in which the branching of the amylopectin component of starch is modified.

47. A method according to any one of claims 43 to 46, in which a plant with high amylose content is produced.

48. A method according to any one of claims 43 to 46, in which a plant with high amylose content is produced.

49. A method according to any one of claims 43 to 46, in which a plant with low amylopectin content is produced.

50. A method of targeting expression of a desired gene to the endosperm of a cereal plant, comprising the step of transforming the plant with a genetic construct according to any one of claims 11 to 23.

51. A method of identifying a null or altered allele encoding an enzyme of the starch biosynthetic pathway, comprising the step of subjecting DNA from a plant suspected to possess such an allele to a DNA fingerprinting or amplification assay which utilises at least one DNA probe comprising a nucleic acid molecule according to any one of claims 1 to 10.

52. An oligonucleotide probe selected from the group consisting of SEQ ID NOS:11 to 17.

Sequence of the wheat SBE9 (BEIIa) cDNA

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1  ACGTTGCTCC CCCTTCTCAT CGCTTCTCAA TTAATATCTC CATCACTCGG
51  TTCCGCGCTG CATTTCGGCC GGC GG GTTGA GTGAGATCTG GGCCACTGAC
101 CGACTCACTC GCTCGCTGCG GGGATGGCGA CGTTCGCGGT GTCCGGCGCG
151 ACCCTCGGTG TGGCGCGGCC GCCGGCGGCG GCGCAACCTG AAGAATTACA
201 GATACCTGAA GACATCGAGG AGCAAACGGC TGAAGTAAAC ATGACAGGGG
251 GGACTGCAGA AAAACTTGAA TCTTCAGAAC CGACTCAAGG CATTGTGGAA
301 ACAATCACTG ATGGTGTAAC CAAAGGAGTT AAGGAACTAG TCGTGGGGGA
351 GAAACCGCGA GTTGTCCCAA AACCAGGAGA TGGGCAGAAA ATATACGAGA
401 TTGACCCAAC GCTGAAAGAT TTTCGGAGCC ATCTTGACTA CCGATACAGC
451 GAATACAGGA GAATTCGTGC TGCTATTGAC CAACATGAAG GTGGATTGGA
501 AGCATTTTCT CGTGGTTATG AAAAGCTTGG ATTTACCCGC AGTGCTGAAG
551 GTATCACTTA CCGAGAATGG GCTCCTGGAG CGCATTCTGC AGCATTAGTA
601 GGTGACTTCA ACAATTGGAA TCCGAATGCA GATACTATGA CCAGAGATGA
651 TTATGGTGTT TGGGAGATTT TCCTCCCTAA CAATGCTGAT GGATCCCCAG
701 CTATTCCTCA TGGCTCACGT GTAAAGATAC GGATGGATAC TCCATCTGGT
751 GTGAAGGATT CAATTTCTGC TTGGATCAAG TTCTCTGTGC AGGCTCCAGG
801 TGAAATACCA TTCAATGGCA TATATTATGA TCCACCTGAA GAGGAGAAAGT
851 ATGTCTTCCA ACATCCTCAA CCTAAACGAC CAGAGTCACT GAGGATTTAT
901 GAATCACACA TTGGAATGAG CAGCCCAGAA CCGAAGATAA ATTCATATGC
951 TAATTTTAGG GATGAGGTGC TGCCAAGAAT TAAAAGGCTT GGATACAATG
1001 CAGTGCAGAT AATGGCAATC CAGGAGCATT CATACTATGC GAGCTTTGGG
1051 TACCATGTTA CTAATTTTTT TGCACCAAGT AGCCGTTTTG GAACTCCAGA
1101 GGACTTAAAA TCCCTGATCG ATAGAGCACA TGAGCTTGGT TTGCTTGTTT
1151 TTATGGATAT TGTTCATAGT CATTTCATCAA ATAATACCCT TGACGGCTTG
1201 AATGGTTTCG ATGGCACTGA TACACATTAC TTCCACGGTG GTCCACGTGG
1251 CCATCATTGG ATGTGGGATT CTCGTCTATT CAACTATGGG AGTTGGGAAG
1301 TATTGAGATT CTTACTGTCA AACGCGAGAT GGTGGCTTGA AGAATATAAG
1351 TTTGATGGAT TTCCGATTGA TGGGGTGACC TCCATGATGT ATACTCACCA
1401 TGGATTACAA ATGACATTTA CTGGGAACTA TGGCGAGTAT TTTGGATTTG
1451 CTACTGATGT TGATGCGGTA GTTTACTTGA TGCTGGTCAA CGATCTAATT
1501 CATGGACTTC ATCCTGATGC TGTATCCATT GGTGAAGATG TCAGTGGAAT
1551 GCCCACATTT TGCATCCCTG TTCCAGATGG TGGTGTGGT TTTGACTATC
1601 GCTTGCAATG GGCTGTAGCA GATAAATGGA TTGAACTCCT CAAGCAAAGT
1651 GACGAATCTT GGAAAATGGG TGATATTGTG CACACCCTAA CAAATAGAAG
1701 GTGGCTTGAG AAGTGTGTAA CTTATGCAGA AAGTCATGAT CAAGCACTAG
1751 TTGGTGACAA GACTATTGCA TTCTGGTTGA TGGATAAGGA TATGTATGAT
1801 TTCATGGCTC TGGATAGGCC TTCAACTCCT CGCATTGATC GTGGCATAGC
1851 ATTACATAAA ATGATCAGGC TTGTCAACAT GGGTTTAGGT GGTGAAGGCT
1901 ATCTTAACTT CATGGGAAAT GAGTTTGGGC ATCCTGAATG GATAGATTTT
1951 CCAAGAGGTC CGCAAACCTC TCCAACCGGC AAAGTTCTCC CTGGAAATAA
2001 CAATAGTTAT GATAAATGCC GCCGTAGATT TGATCTTGGA GATGCAGATT
2051 TTCTTAGATA TCATGGTATG CAAGAGTTCG ATCAGGCAAT GCAGCATCTT
2101 GAGGAAAAAT ATGGGTTTAT GACATCTGAG CACCAGTATG TTTCACGGAA
2151 ACATGAGGAA GATAAGGTGA TCATCTTCGA AAGAGGAGAT TTGGTATTTG
2201 TTTTCAACTT CCACTGGAGC AATAGCTTTT TTGACTACCG TGTTGGGTGT
2251 TCCAGGCCTG GGAAGTACAA GGTGGCCTTA GACTCCGACG ATGCACTCTT
2301 TGGTGGATTG AGCAGGCTTG ATCATGATGT CGACTACTTC ACAACCGAAC
2351 ATCCGCATGA CAACAGGCCG CGCTCTTTCT CCGTGTACAC TCCGAGCAGA
2401 ACTGCGGTCG TGTATGCCCT TACAGAGTAA GAACCAGCAG CTGCTTGTTA
2451 CAAGGCAAAG AGAGAACTCC AGAGAGCTCG TGGATCGTGA CCGAAGCGAC
2501 GGGCAACGGC GCGAGGCTGC TCTAAGCGCC ATGACTGGGA GGGGATCGTG
2551 CCTCTTCCCC AGATGCCAGG AGGAGCAGAT GGATAGGTAG CTTGTTGGTG
2601 AGCGCTCGAA AGAAAATGGA CGGGCCTGGG TGTGTGTCGT GCTGCACTAC
2651 CCTCCTCCTA TCTTGACAT TCCCGGTTGT TTTTGTACAT ATAACATAA
2701 ATTGCCCGTG CGCTCAACGT GAACAA

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Figure 1

Sequence of the Starch Branching Enzyme II gene (wSBE II-D1)
from *A. tauschii*

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1  AGAAACACCT CCATTTTAGA TTTTTTTTTT GTTCTTTTCG GACGGTGGGT
51  CGTGGAGAGA TTAGCGTCTA GTTTTCTTAA AAGAACAGGC CATTTAGGCC
101 CTGCTTTACA AAAGGCTCAA CCAGTCCAAA ACGTCTGCTA GGATCACCAG
151 CTGCAAAGTT AAGCGCGAGA CCACCAAAAC AGGCGCATTC GAACTGGACA
201 GACGCTCACG CAGGAGCCCA GCACCACAGG CTTGAGCCTG ACAGCGGACG
251 TGAGTGCGTG ACACATGGGG TCATCTATGG GCGTCGGAGC AAGGAAGAGA
301 GACGCACATG AACACCATGA TGATGCTATC AGGCCTGATG GAGGGAGCAA
351 CCATGCACCT TTTCCCCTCT GGAAATTCAT AGCTCACACT TTTTTTTAAT
401 GGAAGCAAGA GTTGGCAAAC ACATGCATTT TCAAACAAGG AAAATTAATT
451 CTCAAACCAC CATGACATGC AATTCTCAA CCATGCACCG ACGAGTCCAT
501 GCGAGGTGGA AACGAAGAAC TGAAAATCAA CATCCCAGTT GTCGAGTCGA
551 GAAGAGGATG AACTGAAAG TATGCGTATT ACGATTTTCAT TTACATACAT
601 GTACAAATAC ATAATGTACC CTACAATTTG TTTTTTGGAG CAGAGTGGTG
651 TGGTCTTTTT TTTTACACG AAAATGCCAT AGCTGGCCCG CATGCGTGCA
701 GATCGGATGA TCGGTCGGAG ACGACGGACA ATCAGACACT CACCAACTGC
751 TTTTGTCTGG GACACAATAA ATGTTTTTGT AAACAAAATA AATACTTATA
801 AACGAGGGTA CTAGAGGCCG CTAACGGCAT GGCCAGGTAA ACGCGCTCCC
851 AGCCGTTGGT TTGCGATCTC GTCCTCCCGC ACGCAGCGTC GCCTCCACCG
901 TCCGTCCGTC GCTGCCACCT CTGCTGTGCG CGCGCACGAA GGGAGGAAGA
951 ACGAACGCCG CACACACACT CACACACGGC ACACTCCCCG TGGGTCCCCT
1001 TTCCGGCTTG GCGTCTATCT CCTCTCCCCC GCCCATCCCC ATGCACTGCA
1051 CCGTACCCGC CAGCTTCCAC CCCCGCCGCA CACGTTGCTC CCCCTTCTCA
1101 TCGCTTCTCA ATTAATATCT CCATCACTCG GGTTCCGCGC TGCATTTCGG
1151 CCGGCGGGTT GAGTGAGATC TGGGCGACTG GCTGACTCAA TCACTACGCG
1201 GGGATGGCGA CGTTCGCGGT GTCCGGCGCG ACTCTCGGTG TGGCGCGGGC
1251 CGGCGTCGGA GTGGCGCGGG CCGGCTCGGA GCGGAGGGGC GGGGCGGACT
1301 TGCCGTCGCT GCTCCTCAGG AAGAAGGACT CCTCTCGTAC GCCTCGCTCT
1351 CTCGAATCTC CCCCCTCTGG CTTTGGCTCC CCTTCTCTCT CCTCTGCGCG
1401 CGCATGGCCT GTTCGATGCT GTTCCCCAAT TGATCTCCAT GAGTGAGAGA
1451 GATAGCTGGA TTAGGCGATC GCGCTTCCTG AACCTGTATT TTTTCCCCCG
1501 CGGGGAAATG CGTTAGTGTC ACCCAGGCCC TGGTGTTACC ACGGCTTTGA
1551 TCATTCTCTG TTTCAATCTG ATATATATTT TCTCATTTCT TTTCTTCCTG
1601 TTCTTGCTGT AACTGCAAGT TGTGGCGTTT TTTCATATT TTTTCCCTG
1651 TTGCATTTTG CAGGCGCCGT CCTGAGCCGC GCGGCCTCTC CAGGGAAGGT
1701 CCTGGTGCCT GACGGCGAGA GAGACGACTT GGCAAGTCCG GCGCAACCTG
1751 AAGAATTACA GGTACACACA CTCGTGCCGG TAAATCTTCA TACAATCGTT
1801 ATTCACTTAC CAAATGCCGG ATGAAACCAA CCACGGATGC GTCAGGTTTC
1851 GAGCTTCTTC TATCAGCATT GTGCAGTACT GCACTGCCTT GTTCATTTTG
1901 FTAGCCTTGG CCCCCTGCTG GCTCTTGGGC CACTGAAAAA ATCAGATGGA
1951 TGTGCATTCT AGCAAGAAGT TCACAACATA ATGCACCGTT TGGGGTTTCG
2001 TCAGTCTGCT CTACAATTGC TATTTTTTCG GCTGTAGATA CCTGAAGATA
2051 TCGAGGAGCA AACGGCGGAA GTGAACATGA CAGGGGGGAC TGCAGAGAAA
2101 CTTCAATCTT CAGAACCGAC TCAGGGCATT GTGGAAACAA TCACTGATGG
2151 TGTAACCAAA GGAGTTAAGG AACTAGTCGT GGGGGAGAAA CCGCGAGTTG
2201 TCCCAAACCC AGGAGATGGG CAGAAAATAT ACGAGATTGA CCCAACACTG
2251 AAAGATTTTC GGAGCCATCT TGACTACCGG TAATGCCTAC CCGCTGCTTT
2301 CGCTCATTTT GAATTAAGGT CCTTTCATCA TGCAAATTTG GGAACATCA
2351 AAGAGACAAA GACTAGGGAC CACCATTTC TACAGATCCC TTCGTGGTCT
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2451 AATTGCAATA CGAATAACTG TCTCCGATCA TTACAATTAA AGAGTGGCAA
2501 ACTGATGAAA ATGTGGTGGA TGGGTATAG ATTTTACTTT GCTAATTCCT
2551 CTACCAAATT CCTAGGGGGG AAATCTACCA GTTGGGAAAC TTAGTTTCTT
2601 ATCTTTGTGG CCTTTTTGTT TTGGGGAAAA CACATTGCTA AATTCGAATG
2651 ATTTTGGGTA TACCTCGGTG GATTCAACAG ATACAGCGAA TACAAGAGAA
2701 TTCGTGCTGC TATTGACCAA CATGAAGGTG GATTGGAAGC ATTTTCTCGT
2751 GGTATGAAA AGCTTGGATT TACCCGCAGG TAAATTTAAA GCTTTATTAT
2801 TATGAAACGC CTCCACTAGT CTAATTCAT ATCTTATAAG AAAATTTATA
2851 ATTCCTGTTT TCCCCTCTCT TTTTTCAGT GCTGAAGGTA TCGTCTAATT
2901 GCATATCTTA TAAGAAAATT TATATTCCTG TTTTCCCCTA TTTTCCAGTG
2951 CTGAAGGTAT CACTTACCGA GAATGGGCTC CCTGGAGCGC ATGTTATGTT
3001 CTTTAAAGTT CCTTAACGAG ACACCTTCCA ATTTATTGTT AATGGTCACT
3051 ATTCACCAAC TAGCTTACTG GACTTACAAA TTAGCTTACT GAATACTGAC
3101 CAGTTACTAT AAATTTATGA TCTGGCTTTT GCACCCTGTT ACAGTCTGCA
3151 GCATTAGTAG GTGACTTCAA CAATTGGAAT CCAAATGCAG ATACTATGAC
3201 CAGAGTATGT CTACAGCTTG GCAATTTTCC ACCTTTGCTT CATACTACT
3251 GATACATCTA TTTGTATTTA TTTAGCTGTT TGCACATTCC TTAAAGTTGA

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Figure 2

3301	GCCTCAACTA	CATCATATCA	AAATGGTATA	ATTTGTCAGT	GTCTTAAGCT
3351	TCAGCCCAAA	GATTCTACTG	AATTTAGTCC	ATCTTTTGA	GATTGAAAAT
3401	GAGTATATTA	AGGATGAATG	AATACGTGCA	ACACTCCCAT	CTGCATTATG
3451	TGTGCTTTTC	CATCTACAAT	GAGCATATTT	CCATGCTATC	AGTGAAGGTT
3501	TGCTCCTATT	GATGCAGATA	TTTGATATGG	TCTTTTCAGG	ATGATTATGG
3551	TGTTTGGGAG	ATTTTCCTCC	CTAACAACGC	TGATGGATCC	TCAGCTATTC
3601	CTCATGGCTC	ACGTGTAAAG	GTAAGCTGGC	CAATTATTTA	GTCGAGGATG
3651	TAGCATTTTC	GAACCTCTGCC	TACTAAGGGT	CCCTTTTCCT	CTCTGTTTTT
3701	TAGATACGGA	TGGATACTCC	ATCCGGTGTG	AAGGATTCAA	TTTCTGCTTG
3751	GATCAAGTTC	TCTGTGCAGG	CTCCAGGTGA	AATACCTTTC	AATGGCATAT
3801	ATTATGATCC	ACCTGAAGAG	GTAAGTATCG	ATCTACATTA	CATTATTAAA
3851	TGAAATTTCC	AGTGTTACAG	TTTTTTAATA	CCCCTTCTT	ACTGACATGT
3901	GAGTCAAGAC	AATACTTTTG	AATTTGGAAG	TGACATATGC	ATTAATTCAC
3951	CTTCTAAGGG	CTAAGGGGCA	ACCAACCTTG	GTGATGTGTG	TATGCTTGTG
4001	TGTGACATAA	GATCTTATAG	CTCTTTTATG	TGTTCTCTGT	TGGTTAGGAT
4051	ATTCCATTTT	GGCCTTTTGT	GACCATTTAC	TAAGGATATT	TACATGCAAA
4101	TGCAGGAGAA	GTATGTCTTC	CAACATCTCA	ACTAAACGAC	CAGAGTCACT
4151	AAGGATTTAT	GAATCACACA	TTGGAATGAG	CAGCCCGGTA	TGTCAATAAG
4201	TTATTTTACC	TGTTTCTGGT	CTGATGGTTT	ATTCTATGGA	TTTTCTAGTT
4251	CTGTTATGTA	CTGTTAACAT	ATTACATGGT	GCATTCACCT	GACAACCTCG
4301	ATTTTATTTT	CTAATGTCTT	CATATTTGGCA	AGTGCAAAAC	TTTGCTTCCT
4351	CTTTGTCTGC	TTGTTCTTTT	GTCTTCTGTA	AGATTTCCAT	TGCATTTGGA
4401	GGCAGTGGGC	ATGTGAAAGT	CATATCTATT	TTTTTTTTGT	CAGAGCATAG
4451	TTATATGAAT	TCCATTGTTG	TTGCAATAGC	TCGGTATAAT	GTAACCATGT
4501	TACTAGCTTA	AGATTTCCCA	CTTAGGATGT	AAGAAATATT	GCATTGGAGC
4551	GTCTCCAGCA	AGCCATTTCC	TACCTTATTA	ATGAGAGAGA	GACAAGGGGG
4601	GGGGGGGGGG	GGGGGTTCCT	TTCATTATTC	TGCGAGCGAT	TCAAAAACCTT
4651	CCATTGTTCT	GAGGTGTACG	TACTGCAGGG	ATCTCCCAT	ATGAAGAGGA
4701	TATAGTTAAT	TCTTTGTAAC	CTACTTGGAA	ACTTGAGTCT	TGAGGCATCG
4751	CTAATATATA	CTATCATCAC	AATACTTAGA	GGATGCATCT	GAATATTTTA
4801	GTGTGATCTT	GCACAGGAAC	CGAAGATAAA	TTCATATGCT	AATTTTAGGG
4851	ATGAGGTGTT	GCCAAGAATT	AAAAGGCTTG	GATACAATGC	AGTGCAGATA
4901	ATGGCAATCC	AGGAGCATTC	ATACTATGCA	AGCTTTGGGT	ATTCACACAA
4951	TCCATTTTTT	TCTGTATACA	CNTCTTCACC	CATTTGGAGC	TATTACATCC
5001	TAATGCTTCA	TGCACATAAA	ATATTTGGAT	ATAATCCTTT	ATTAGATATA
5051	TAGTACAACT	ACACTTAGTA	TTCTGAAATA	AAAGATCATT	TTATTGTTGT
5101	TGGCTTGTTT	CAGGTACCAT	GTTACTAATT	TTTTTGCACC	AAGTAGCCGT
5151	TTTGGAATCT	CAGAGGACTT	AAAATCCTTG	ATCGATAGAG	CACATGAGCT
5201	TGGTTTGCTT	GTTCTTATGG	ATATTGTTCA	TAGGTAATTA	GTCCAATTTA
5251	ATTTTAGCTG	TTTTACTGTT	TATCTGGTAT	TCTAAAGGGA	AATTCAGGCA
5301	ATTATGATAC	ATTGTCAAAA	GCTAAGAGTG	GCGAAAGTGA	AATGTCAAAA
5351	TCTAGAGTGG	CATAAGGAAA	ATTGGCAAAA	ACTAGAGTGG	CAAAAATAAA
5401	ATTTTCCCAT	CCTAAATGGC	AGGGCCCTAT	CGCCGAATAT	TTTTCCATTC
5451	TATATAATTG	TGCTACGTGA	CTTCTTTTTT	CTCAGATGTA	TTAAACCAGT
5501	TGGACATGAA	ATGTATTTGG	TACATGTAGT	AAACTGACAG	TTCCATAGAA
5551	TATCGTTTTG	TAATGGCAAC	ACAATTTGAT	GCCATAGATG	TGGATTGAGA
5601	AGTTCAGATG	CTATCAATAG	AATTAATCAA	CTGGCCATGT	ACTCGTGGCA
5651	CTACATATAG	TTTGCAAGTT	GGAAAACCTGA	CAGCAATACC	TCACTGATAA
5701	GTGGCCAGGC	CCCCTTGCC	AGCTTCATAC	TAGATGTTAC	TTCCCTGTTG
5751	AATTCATTTG	AACATATTAC	TTAAAGTTCT	TCATTTGTCC	TAAGTCAAAC
5801	TTCTTTAAGT	TTGACCAAGT	CTATTGGAAA	ATATATCAAC	ATCTACAACA
5851	CCAAATTACT	TTGATCAGAT	TAACAATTTT	TATTTTATTA	TATTAGCACA
5901	TCTTTGATGT	TGTAGATATC	AGCACATTTT	TCTATAGACT	TGGTCAAATA
5951	TAGAGAAGTT	TGACTTAGGA	CAAATCTAGA	ACTTCAATCA	ATTTGGATCA
6001	GAGGGAACAT	CAAATAATAT	AGATAGATGT	CAACACTTCA	ACAAAAAAT
6051	CAGACCTTGT	CACCATATAT	GCATCAGACC	ATCTGTTTGC	TTTAGCCACT
6101	TGCTTTCATA	TTTATGTGTT	TGTACCTAAT	CTACTTTTCC	TTCTACTTGG
6151	TTTGGTTGAT	TCTATTTTCT	TTGCATTGCT	TCATCAATGA	TTTTGTGTAC
6201	CCTGCAGTCA	TTCGTCAAAT	AATACCCTTG	ACGGTTTGAA	TGGTTTCGAT
6251	GGCACTGATA	CACATTACTT	CCACGGTGGT	CCACGCGGCC	ATCATTGGAT
6301	GTGGGATTCT	CGTCTATTCA	ACTATGGGAG	TTGGGAAGTA	TGTAGCTCTG
6351	ACTTCTGTCA	CCATATTTGG	CTAACTGTTC	CTGTTAATCT	GTTCTTACAC
6401	ATGTTGATAT	TCTATTCTTA	TGCAGGTATT	GAGATTCTTA	CTGTCAAACG
6451	CGAGATGGTG	GCTTGAAGAA	TATAAGTTTG	ATGGATTTTC	ATTTGATCGG
6501	GTGACCTCCA	TGATGTATAC	TCACCATGGA	TTACAAGTAA	GTCATCAAGT
6551	GGTTTCAGTA	ACTTTTTTAG	GGCACTGAAA	CAATTGCTAT	GCATCATAAC
6601	ATGTATCATG	ATCAGGACTT	GTGCTACGGA	GTCTTAGATA	GTTCCCTAGT
6651	ATGCTTGATC	AATTTTACCT	GATGAGATCA	TGGAAGATTG	GAAGTGATTA
6701	TTATTTATTT	TCTTTCTAAG	TTTGTTCCTT	GTTCTAGATG	ACATTTACTG

Figure 2 (cont'd)


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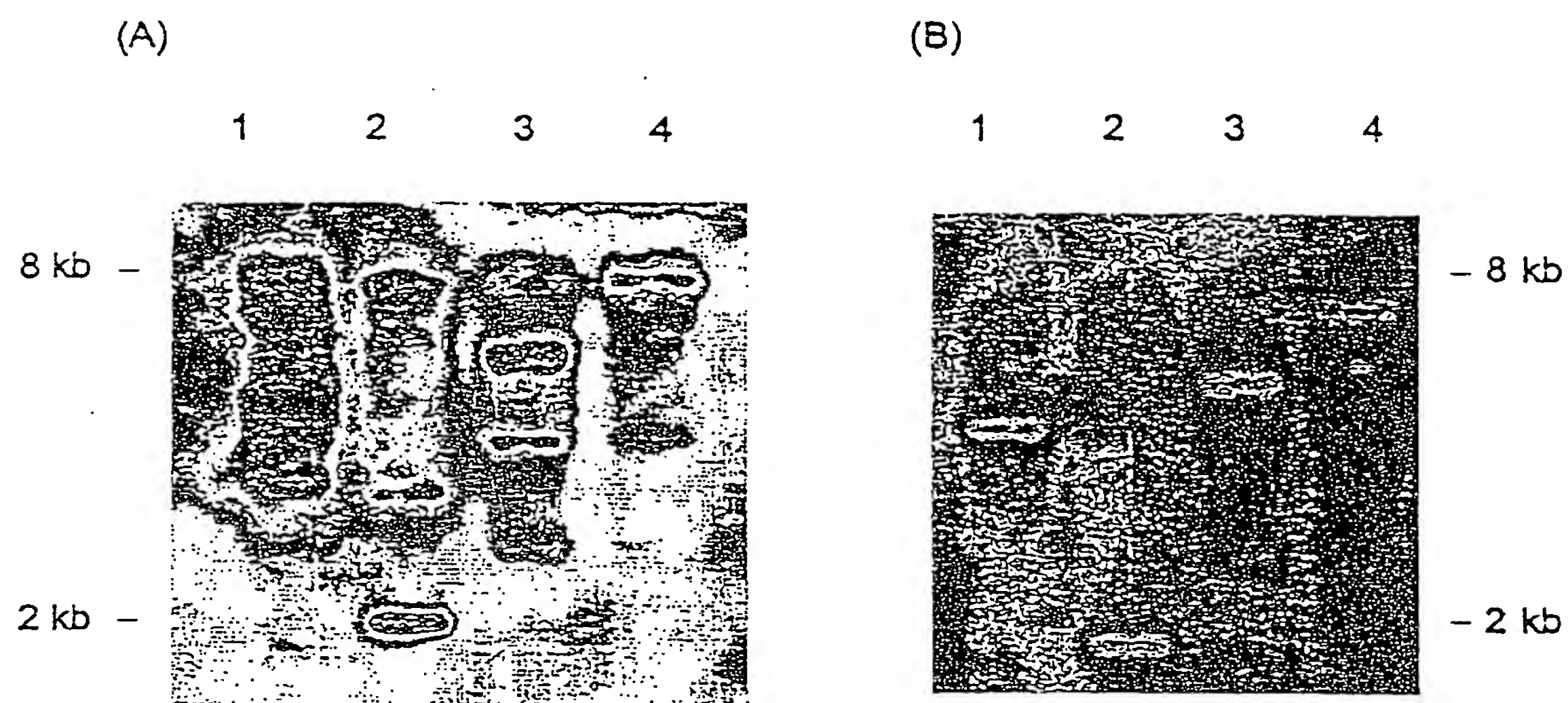
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6801 TACTTGATGC TGGTCAACGA TCTAATTCAT GGACTTTATC CTGATGCTGT
6851 ATCCATTGGT GAAGATGTAA GTGCTTACAG TATTTATGAT TTTTAACTAG
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7001 ATCACTTAAG TAATTTGAAA AGTGCAAGGG CATTCAAGCT TACGAGCATA
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7351 ATCCTGAGCT TTCAACTCAT GAGAAAATAn GAnGTCCGCT TCTGCCAGCA
7401 TTAAGTGTTT ACAGTTCTAA TTTGTGTAACT TGTGAAATTG TTCAGGTCAG
7451 TGGAAATGCCT ACATTTTGCA TCCCTGTTCC AGATGGTGGT GTTGGTTTTG
7501 ACTACCGCCT GCATATGGCT GTAGCAGATA AATGGATTGA ACTCCTCAAG
7551 TAAGTGCAGG AATATTGGTG ATTACATGCG CACAATGATC TAGATTACAT
7601 TTTCTAAATG GTAAAAAGGA AAATATGTAT GTGAATATCT AGACATTTGC
7651 CTGTTATCAG CTTGAATACG AGAAGTCAAA TACATGATTT AAATAGCAAA
7701 TCTCGGAAAT GTAATGGCTA GTGTCTTTAT GCTGGGCAGT GTACATTGCC
7751 CTGTAGCAGG CCAGTCAACA CAGTTAGCAA TATTTTCAGA AACAAATATTA
7801 TTTATATCCG TATATGAnGA AAGTTAGTAT ATAAACTGTG GTCATTAATT
7851 GTGTTACCT TTTGTCTCTG TTAAGGATGG GCAGTAGGTA ATAAATTTAG
7901 CCAGATAAAA TAAATCGTTA TTAGGTTTAC AAAAGGAATA TACAGGGTCA
7951 TGTAGCATAT CTAGTTGTAA TTAATGAAAA GGCTGACAAA AGGCTCGGTA
8001 AAAAAAACTT TATGATGATC CAGATAGATA TGCAGGAACG CGACTAAAGC
8051 TCAAATACTT ATTGCTACTA CACAGCTGCC AATCTGTCAT GATCTGTGTT
8101 CTGCTTTGTG CTATTTTAGAT TTAAATACTA ACTCGATACA TTGGCAATAA
8151 TAAACTTAAC TATTC AACCA ATTTGGTGGA TACCAGAnAT TTCTGCCCTC
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8751 TAGTGTTTTT TTGTGATAAA GATTGGCTGC CTCACCCATC ACCAGCTATT
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8851 GTGGCGGCTT GTGAACCTTG ACAGTTATGT TGCAATTTTC TGTTCTTATT
8901 TATTTGATTG CTTATGTTAC CGTTCATTTG CTCATTCCTT TCCGAGACCA
8951 GCCAAAGTCA CGTGTTAGCT GTGTGATCTG TTATCTGAAT CTTGAGCAAA
9001 TTTTATTAAT AGGCTAAAAT CCAACGAATT ATTTGCTTGA ATTTAAATAT
9051 ACAGACGTAT AGTCACCTGG CTCTTTCTTA GATGATTACC ATAGTCCCTG
9101 AAGGCTGAAA TAGTTTTGGT GTTCTTTGGA TGCCGCCTAA AGGAGTGATT
9151 TTTATTGGAT AGATTCCTGG CCGAGTCTTC GTTACAACAT AACATTTTGG
9201 AGATATGCTT AGTAACAGCT CTGGGAAGTT TGGTCACAAG TCTGCATCTA
9251 CACGCTCCTT GAGGTTTTAT TATGGCGCCA TCTTTGTAAC TAGTGGCACC
9301 TGTAAGGAAA CACATTCAAA AGGAAACGGT CACATCATTC TAATCAGGAC
9351 CACCATACTA AGAGCAAGAT TCTGTTCCAA TTTTATGAGT TTTTGGGACT
9401 CCAAAGGGAA CAAAAGTGTC TCATATTGTG CTTATAACTA CAGTTGTTTT
9451 TATACCAGTG TAGTTTTATT CCAGGACAGT TGATACTTGG TACTGTGCTG
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9701 TGGTCAGTCT TTACAACATT ATTGCATTCT GCATGATTGT GATTTACTGT
9751 AATTTGAACC ATGCTTTTCT TTCACATTGT ATGTATTATG TAATCTGTTG
9801 CTTCCAAGGA GGAAGTTAAC TTCTATTTAC TTGGCAGAAT GGATAGATTT
9851 TCCAAGAGGC CCACAACTC TTCCAACCGG CAAAGTTCTC CCCTGGAAAT
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10101 CATGATTTTT TGCAGGGAGA TGCAGATTTT CTTAGATATC GTGGTATGCA
10151 AGAGTTCGAT CAGGCAATGC AGCATCTTGA GGAAAAATAT GGGGTATGTC
10201 ACTGGTTTGT CTTTGTTGCA TAACAAGTCA CAGTTTAACG TCAGTCTCTT

```

Figure 2 (cont'd)

10251	CAAGTGGTAA	AAAAAGTGTA	GAATTAATTC	CTGTAATGAG	ATGAAAACCTG
10301	TGCAAAGGCG	GAGCTGGAAT	TGCTTTTCAC	CAAAACTATT	TTCTTAAGTG
10351	CTTGTGTATT	GATACATATA	CCAGCACTGA	CAATGTAACT	GCAGTTTATG
10401	ACATCTGAGC	ACCAGTATGT	TTCACGGAAA	CATGAGGAAG	ATAAGGTGAT
10451	CATCCTCnAA	AAGAGGAGAT	TTGGTATTTG	TTTTCAACTT	CCACTGGAGC
10501	AATAGCTTTT	TTGACTACCG	TGTTGGGTGT	TCCAAGCCTG	GGAAGTACAA
10551	GGTATGCTTG	CCTTTTCATT	GTCCACCCTT	CACCAGTAGG	GTTAGTGGGG
10601	GCTTCTACAA	CTTTTAATTC	CACATGGATA	GAGTTTGTTG	GTCGTGCAGC
10651	TATCAATATA	AAGAATAGGG	TAATTTGTAA	AGAAAAGAAT	TTGCTCGAGC
10701	TGTTGTAGCC	ATAGGAAGGT	TGTTCTTAAC	AGCCCCGAAG	CACATACCAT
10751	TCATTCATAT	tATCTACTTA	AGTGTTTGTT	TCAATCTTTA	TGCTCAGTTG
10801	GACTCGGTCT	AATACTAGAA	CTATTTTCCG	AATCTACCCT	AACCATCCTA
10851	GCAGTTTtag	AGCAGCCCCA	TTTGGACAAT	TGGCTGGGTT	TTTGTtagTT
10901	GTGACAGTTT	CTGCTATTTT	TTAATCAGGT	GGCCTTGGAC	TCTGACGATG
10951	CACTCTTTGG	TGGATTcAGC	AGGCTTGATC	ATGATGTcGA	CTACTTCACA
11001	ACCGTAAGTC	TGGGCTCAAG	CGTCACTTGA	CTCGTCTTGA	CTCAACTGCT
11051	TACAAATCTG	AATCAACTTC	CCAATTGCTG	ATGCCCTTGC	AGGAACATCC
11101	GCATGACAAC	AGGCCGCGCT	CTTTCTCGGT	GTACACTCCG	AGCAGAACTG
11151	CGGTcGTGTA	TGCCCTTACA	GAGTAAGAAC	CAGCAGCGGC	TTGTTACAAG
11201	GCAAAGAGAG	AACTCCAGAG	AGCTCGTGGA	TCGTGAGCGA	AGCGACGGGC
11251	AACGGCGCGA	GGCTGCTCCA	AGCGCCATGA	CTGGGAGGGG	ATCGTGcCTC
11301	TTCCCCAGAT	GCCAGGAGGA	GCAGATGGAT	AGGTAGCTTG	TTGGTGAGCG
11351	CTCGAAAGAA	AATGGACGGG	CCTGGGTGTT	TGTTGTGCTG	CACTGAACCC
11401	TCCTCCTATC	TTGCACATTC	CCGGTTGTTT	TTGTACATAT	AACTAATAAT
11451	TGCCCCGTGCG	CTCAACGTGA	AAATCC		

6/34



- Figure 3

DNA sequence of INTRON 5 PCR Fragments

	1		50
D genome	ATCACTTACC GAGAATGGGC TCCT.GGAGC GCATGTATGT CTTT.....		
A genome	ATCACTTACC GAGAATGGGC TCCT.GGAGC GCATGTACGT CTTT.....		
B genome	ATCACTTACC GAGAATGGGC TCCT.GGAGC GCATGTAC..		
262bp	ATCACTTACC GAGAATGGGC TCCTGNGAGC ANATGTATGT TCTTCTGACT		
	51		100
D genome	...TAAGTCT TAACAGACAC CTTCCAATTT ATTGTTAATG GT..CACTAT		
A genome	...TAAGTCT TAACAGACAC CTTCCAATTC ATTGTTAATG GTCACACTAT		
B genomeGTCT TAACAGACAC CTTCTAATTT ATTGTTAATG GT..CACTAT		
262bp	GTCTGATCGT TTACCTGACT ATACTAATTC TATCTTTCAA CTGCTTGTTGA		
	101		150
D genome	TCACCAACTA GCTTACTGGA CTTACAAATT AGCTTACTGA ATACTGACCA		
A genome	TCACCAACTA GCTTACTGGA CTTACAACTT AGCTTACTGA ATACTGACCA		
B genome	TCACCAACTA GCTTACTGGA CTTACAAAAT AGCTTACTGA ATACTGACCA		
262bp	ATAATTAGTG CTCATCTGCT ATCCTAAGGT TGGGGATTTT GCACTTCCCA		
	151		200
D genome	GTTA.....	CT ATAAATTTAT GATCTGGCTT	
A genome	GTTG.....	CT CTAAATTTAT GATCTGGCTT	
B genome	GTTA.....	CT CTAAATTTAT GATCTGGCTT	
262bp	GATGAACAGC ATATTAAGTT GCACAACTAN CTTTATTTAA GAACTAACTC		
	201		250
D genome	TTGCACCCTG TTACAGTCTG CAGCATTAGT AGGTGACTTC AACAATTGGG		
A genome	TTGCATCCTG TTACAGTCTG CAGCATTAGT AGGTGACTTC AACAATTGGA		
B genome	TTGGATCCTG TTACAGTCTG CAGCATTAGT AGGTGACTTC AACAATTGGA		
262bp	TTGCTTCCAA TTGCAGTCTG CAACATTAGT TGGCGACTTC AACAATTGGA		
	251	262	
D genome	ATCCAAATGC AG		
A genome	ATCCAAATGC AG		
B genome	ATCCAAATGC AG		
262bp	ATCCAAATGC AG		

- Figure 4

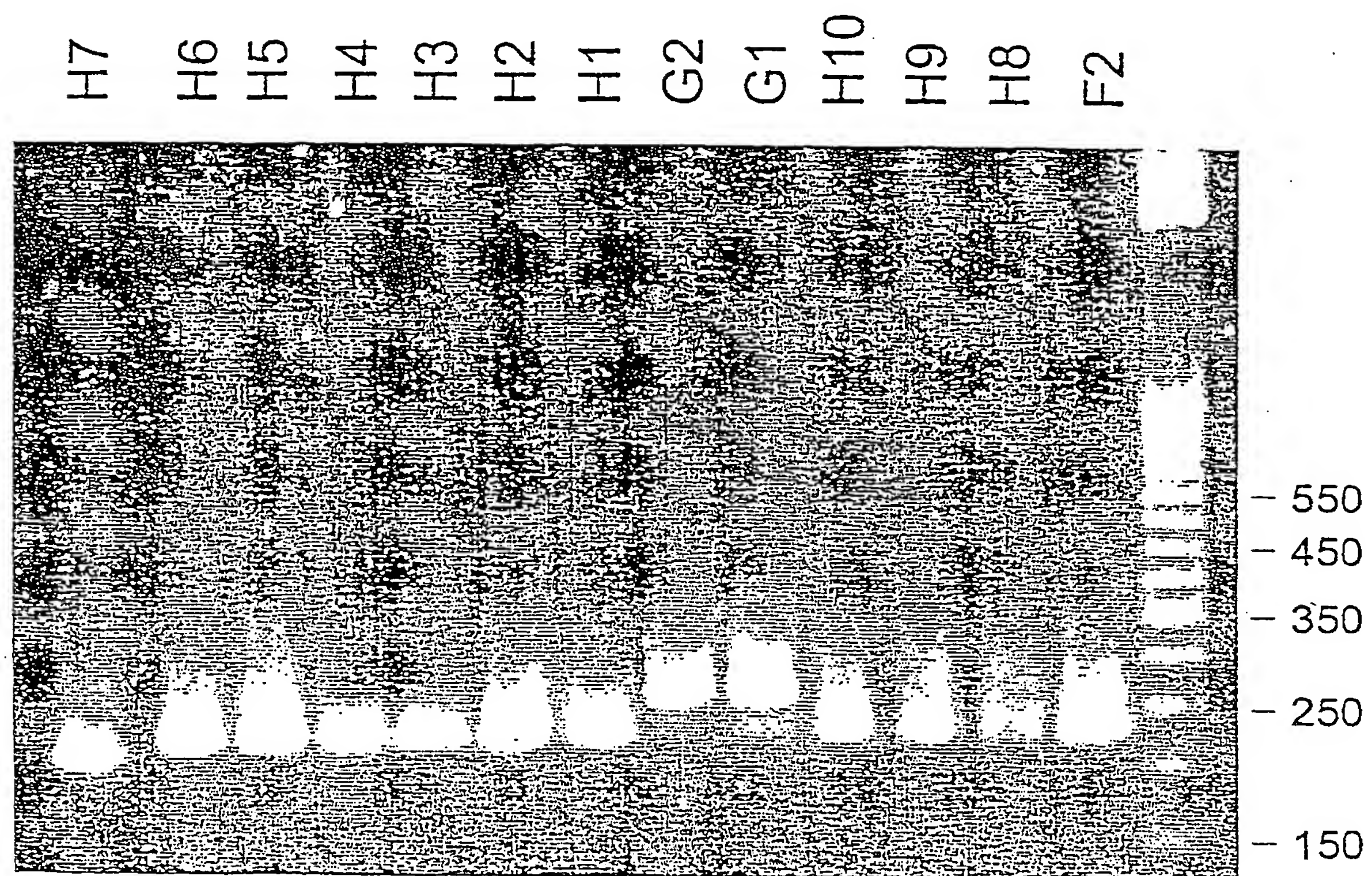


Figure 5

Comparison of Universal 262 bp Sequence with the Wheat Branching Enzyme Iib Gene

FILE NAME	-5	4	14	24	34	44	54
262bpATCACTTACCGAGAATGGGCTCCTGNGAGCANATGTATGTTCTTCTGACTGTCT						
WBEIIB	GAAGGTATCACTTACCGAGAATGGGCTCCTGG-AGCAGATGTACGTTCTTCTAACCATCT						
	2010	2019	2029	2039	2049	2059	2069
FILE NAME	55	64	74	84	94	104	114
262bp	GATCGTTTACCTGACTATACTAATTCTATCTTTCAACTGCTTGTGAATAATTAGTGCTCA						
WBEIIB	GATCGTTTACCTGACTATACTAATTCTATCTTTCAACTAATTGTGAATAATTACTGCTCA						
	2070	2079	2089	2099	2109	2119	2129
FILE NAME	115	124	134	144	154	164	174
262bp	TCTGCTATCCTAAGGTTGGGGATTTTGCACCTCCCAGATGAACAGCATATTAAGTTGCAC						
WBEIIB	TCAGCTATCCTAAGGTTGGGGATTTTGCACCTCCCAGATGAACAGCATATTAAGTCGCAC						
	2130	2139	2149	2159	2169	2179	2189
FILE NAME	175	184	194	204	214	224	234
262bp	AACTANCTTTATTTAAGAACTAACTCTTGCTTCCAATTGCAGTCTGCAACATTAGTTGGC						
WBEIIB	AACTAGCATTATT-AAGAACTAACTCCTGCTTCCAATTGCAGTCTGCAGCATTAGTTGGC						
	2190	2199	2209	2219	2229	2239	2249
FILE NAME	235	244	254	264	274		
262bp	GACTTCAACAATTGGAATCCAAATGCAG.....						
WBEIIB	GACTTCAACAATTGGGATCCAAATGCAGACCATATGAGCAAAG						
	2250	2259	2269	2279	2289		

Figure 6

Comparison of BARLEY BEIIB CDNA, WHEAT BEIIB CDNA, wheat BEIa cDNA sequences with the wSBE II-DB1 gene

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BARLEY BEIIB CDNA      603 TCGCAGCGCT GAAGGTATCA CTTACCGAGA ATGGGCTCCT GGAGCAGAT- 10 20 30 40 50 60 70 80 90
692
WHEAT BEIIB CDNA      802 G*****
--- 891
SBE9 CDNA             537 C*****GC*-
626
WHEAT BEIIB GENE      2000 AT*****G TACGTTCTTC TAACCATCTG ATCGTTTACC TGACTATACT
2089

BARLEY BEIIB CDNA      693 100 110 120 130 140 150 160 170 180
782
WHEAT BEIIB CDNA      892
981
SBE9 CDNA             627
716
WHEAT BEIIB GENE      2090 AATTCTATCT TTCAACTAAT TGTGAATAAT TACTGCTCAT CAGCTATCCT AAGGTTGGG ATTTGCACC TCCAGATGA ACAGCATATT
2179

BARLEY BEIIB CDNA      783 190 200 210 220 230 240 250 260 270
TGGGATCCAA 872
WHEAT BEIIB CDNA      982 *****
1071
SBE9 CDNA             717 *****A**T** *****A****G*
806
WHEAT BEIIB GENE      2180 AAGTCGCACA ACTAGCAITTA TTAAGAACTA ACTCCTGCTT CCAATTGCAG *****
2269

BARLEY BEIIB CDNA      873 CTGCAGACCA TATGAGCAAA 280 290 300 310 320 330 340 350 360
962
WHEAT BEIIB CDNA      1072 A*****
1161
SBE9 CDNA             807 A*****TAC *****C**G*
896

```

Figure 7

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WHEAT BEIIB GENE 2270 A***** GTATGCATGT AGTTCACAA ATATATCATA TTTCCTCTGT AGATTCTT TTTTAGATCG GCTTATCT
2359

BARLEY BEIIB CDNA 963 370 380 390 400 410 420 430 440 450
1052

WHEAT BEIIB CDNA 1162
1251

SBE9 CDNA CDNA 897
----- 986

WHEAT BEIIB GENE 2360 TTAATGTGG TTGAATATAC ACCTTATATG TACGTTGAGC TGTAATATA GTTGAAGTG TTAGGAGTA TTAATTCAC TGGACTCTAT
2449

BARLEY BEIIB CDNA 1053 460 470 480 490 500 510 520 530 540
1142

WHEAT BEIIB CDNA 1252
1341

SBE9 CDNA 987
-----

1076

WHEAT BEIIB GENE 2450 TCTTTCACCT GCCTGTTGCA CGAGCCCAT ACTAGATATC AATGTTGATG ATGCTTTTGT TGTATGAGGT CGAAGTGAAA CATGCATGTT
2539

BARLEY BEIIB CDNA 1143 550 560 570 580 590 600 610 620 630
1232

WHEAT BEIIB CDNA 1342
1431

SBE9 CDNA 1077
-----

1166

WHEAT BEIIB GENE 2540 ACCCTTTTAT ATAAGTAAGG TTGCACATGT ATTTTTTATG ATCTAAACAT TATTTACTGA TTTTGTCTT GCAAGACACT AAGCAGTTT
2629

BARLEY BEIIB CDNA 1233 640 650 660 670 680 690 700 710 720
1322

WHEAT BEIIB CDNA 1432
1521

SBE9 CDNA 1167
-----

1256

WHEAT BEIIB GENE 2630 ACATAATAAT GCGTTGGAG CAGGCCGACT GCACATCTGA ACTGTAGCTC CATGTGTTG ATATAGATTA CAAATGCTCA TATTCATATG
2719

730 740 750 760 770 780 790 800 810

```

Figure 7 (cont'd)

12/34

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BARLEY BEIIB CDNA 1323 ----- --AATGACTT GGGTATTG GAGATTTTC TGCCAAACAA TGCAGATGGT TCGCCGCCAA TTCTCATGG CTCACGGG
1412
WHEAT BEIIB CDNA 1522 ----- --*****C* T***G***** *****A***** *****C** *****
1611
SBE9 CDNA 1257 ----- --G***T*A T***G***** *C**T***** **T*****A **C**AG**T* *****T**A
1346
WHEAT BEIIB GENE 2720 AACTGTATTTC AG*****C* T***G***** *****A***** *****C** *****
2809

BARLEY BEIIB CDNA 1413 AAGGT----- 820 830 840 850 860 870 880 890 900
1502
WHEAT BEIIB CDNA 1612 *****----- 820 830 840 850 860 870 880 890 900
1701
SBE9 CDNA 1347 ***A*----- 820 830 840 850 860 870 880 890 900
1436
WHEAT BEIIB GENE 2810 *****TGTTT TCTTCTCCTT GCCAACGGTG TTAGGCTCAG GAACATGTCC TGTATTACTC AGAAGCTCTT TTGAACATCT AGGT**A**
2899

BARLEY BEIIB CDNA 1503 GGATACTCCA TCTGGGACAA AGGATTCAAT TCCTGCTTGG ATCAAGTACT CCGTGCAGAC TCCAGGAGAT ATACCATACA ATGGAATATA
1592
WHEAT BEIIB CDNA 1702 **G***** ***** ***** ***** ***** ***** ***** *****
1791
SBE9 CDNA 1437 *****TG* *****TG* *****T** *T*****G* *****T**A *****T** *****C*****
1526
WHEAT BEIIB GENE 2900 ***** ***** ***** ***** ***** ***** ***** *****
2989

BARLEY BEIIB CDNA 1593 TTATGACCCT CCTGAAGAGG 1000 1010 1020 1030 1040 1050 1060 1070 1080
1682
WHEAT BEIIB CDNA 1792 *****T*** *C***** 1000 1010 1020 1030 1040 1050 1060 1070 1080
1881
SBE9 CDNA 1527 *****T**A ***** 1000 1010 1020 1030 1040 1050 1060 1070 1080
1616
WHEAT BEIIB GENE 2990 *****T*** *C***** TATTTTACTT CATCTTCTGT GCTTTTATAGAT TTCAGATATT TTTATATAGAA GAAAAATTATG ATTTTTTCCC
3079

BARLEY BEIIB CDNA 1683 1090 1100 1110 1120 1130 1140 1150 1160 1170
1772
WHEAT BEIIB CDNA 1882 ----- 1090 1100 1110 1120 1130 1140 1150 1160 1170
1971

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Figure 7 (cont'd)

[illegible]

Partial Sequence of the *A. tauschii* Branching Enzyme Iib gene

```

1  GGATCCGATC CGGCTGCGGC GCGGCGGACG GGATGGCTGC GCCGGCATTC GCAGTTTCCG
61  CGGCGGGGCT GGCCCGGCCG TCGGCTCCTC GATCCGGCGG GGCAGAGCGG AGGGGGCGCG
121 GGGTGGAGCT GCAGTCGCCA TCGCTGCTCT TCGGCCGCAA CAAGGGCACC CGTTCACCCC
181 GTAATTATTT GCGCCACCTT TCTCACTCAC ATTCTCTCGT GTATTCTGTC GTGCTCGCCC
241 TTCGCCGACG ACGCGTGCCG ATTCCGTATC GGGCTGCGGT GTTCAGCGAT CTTACGTCGG
301 TTCCCTCCTG GTGTGGTGAT GTCTGTAGGT GCCGTCGGCG TCGGAGGTTC TGGATGGCGC
361 GTGGTCATGC GCGCGGGGGG GCCGTCCGGG GAGGTGATGA TCCCTGACGG CGGTAGTGGC
421 GGAACACCGC CTTCCATCGA CCGTCCCCTT CAGTTCGATT CTGATGATCT GAAGGTAGTT
481 TTTTTTTTGC ATCGATCTGA AGGTACTTGA CATATACTAC TGTATTACCC TGAGTAAATA
541 CTGCCACCAT ATTTTTATGG TTCGCTTGAA ATACCTGTFT ACTTGCTACG GTTTTCACTT
601 TCATTGAGAC GTCGGACGAA ATTCAGTGAA TTCCTATAAT TTGGTAGACA CCGAAATATA
661 TACTACTCCT TCCGTCCCAT AATATAAGAG CGTTTTTTGGC ACCTTATATT ATAGGGCGGA
721 GGGAGTACCT TTTAGGTCAA AATATTGTGG TAGTTTCAAT TGTATACAAG AATTCAAATA
781 TTTTTTTTAA AAAAAAATCA ACTAATTGGT TGAGTTTCAA GTGAAGCGTT TTGGTCCTTT
841 GGCTGAGATG TAAACCGAAA TCACTGAAAT TCATAGTAGC CGAAACTTTA ATAGAACTGA
901 AACTCAAAAT CTGCTATCCG GCGAAATTCT AAAGATTTGC TTATTTTACA CGTAGGTTGC
961 AGTACACCCT CTTTCTAATT TATTGGGGAA GGGGTATTAT TATCTTGTTA GTACCTGCCT
1021 GCATGACAAT TGAATCTAA GACAAAACAC CATATGCGAG GCCTACACAC GGTAGGTTGG
1081 TTTACAATA TGTGTGCCAC AGTTCGTCTG AACTTTTTGT CCTTCACATC GTGTTAGGTT
1141 CCATTCATTG ATGATGAAAC AAGCCTACAG GATGGAGGTG AAGATAGTAT TTGGTCTTCA
1201 GAGACAAATC AGGTTAGTGA AGAAATTGAT GCTGAAGACA CGAGCAGAAT GGACAAAGAA
1261 TCATCTACGA GGGAGAAATT ACGCATTCTG CCACCACCGG GAAATGGACA GCAAATATAC
1321 GAGATTGACC CAACGCTCCG AGACTTTAAG TACCATCTTG AGTATCGGTA TGCTTCGCTT
1381 CTATTGTGTG CACTTTAAAA ACAATTTACA GTCTTTGATA AGATGTGAAT GGCTGCTTGC
1441 TGTGACACGA AACTCTTGAA GTTCGTAGTC ACTCTTGTTG GTTCATGGTT CTGAGGTAAC
1501 ATGGTAACCG AACAAAAATA GGAAAGTGGC AAGCACTGCA ATGTGAGCTA CTGATAACCA
1561 CCCATTGTAA TTGGGTACAC TGATTAATAT ATATGTCTTC ATGGGCTCTA TTTTTTTTCA
1621 ATATCTATGC CAATTGAACA ACAATGCTTT GTGGACGGGT GTTCTTTTAC CCTCTTCTTC
1681 TATCAATAGA TGATATGCAT ACTCATGCGT ATCCTACAAA AAATTGAACA ACAATGCCAC
1741 TTTCCCCCGT GTTGCTTTTG TAAGGATGAA ACACATATGT CCAGATCAAA CTATACTAGC
1801 AGTCTAACTG TGCCTTAATG GATCAAAAAC AGATATAGCC TATACAGGAG AATACGTTCA
1861 GACATTGATG AACACGAAGG AGGCATGGAT GTATTTTCCC GCGGTTACGA GAAGTTTGGA
1921 TTTATGCGCA GGTGAAATTT CTTGACTAAA TAACTATGTA TCTACCTTTT CTTTGTACTC
1981 TATCAACATT CCTCTTCCCA TGCAGCGCTG AAGGTATCAC TTACCGAGAA TGGGCTCCTG
2041 GAGCAGATGT ACGTTCTTCT AACCATCTGA TCGTTTACCT GACTATACTA ATTCTATCTT
2101 TCAACTAATT GTGAATAATT ACTGCTCATC AGCTATCCTA AGGTTGGGGA TTTTGCACCT
2161 CCCAGATGAA CAGCATATTA AGTCGCACAA CTAGCATTAT TAAGAACTAA CTCCTGCTTC
2221 CAATTGCAGT CTGCAGCATT AGTTGGCGAC TTCAACAATT GGGATCCAAA TGCAGACCAT
2281 ATGAGCAAAG TATGCATGTA GTTTCACAAA TATATCATAT TTTCTTTGTA GATTTTTTTT
2341 TTTAGATCGG CTTATCTATT TAAATGTGGT TGAATATACA CCTTATATGT ACGTTGAGCT
2401 GTAAATATAG TTGGAAGTGT TTAGGAGTAT TAAATTCACT GGACTCTATT CTTTCACTTG
2461 CCTGTTGCAC GAGCCCATTA CTAGATATCA ATGTTGATGA TGCTTTTGTT GTATGAGGTC
2521 GAAGTGAAAC ATGCATGTTA CCCTTTTATA TAAGTAAGGT TGCACATGTA TTTTTTATGA
2581 TCTAAACATT ATTTACTGAT TTTGTTCTTG CAAGACACTA AGCAGTTTTA CATAATAATG
2641 GCGTTGGAGC AGGCCGACTG CACATCTGAA CTGTAGCTCC ATGTGGTTGA TATAGATTAC
2701 AAATGCTCAT ATTCAATGTA ACTGTTTTCA GAATGACCTT GGTGTTTGGG AGATTTTTCT
2761 GCCAAACAAT GCAGATGGTT CGCCACCAAT TCCTCACGGC TCACGGGTGA AGGTTGTTTT
2821 CTTCTCCTTG CCAACGGTGT TAGGCTCAGG AACATGTCCT GTATTACTCA GAAGCTCTTT
2881 TGAACATCTA GGTGAGAAATG GATACTCCAT CTGGGATAAA GGATTCAATT CCTGCTTGGA
2941 TCAAGTACTC CGTGCACTG CCAGGAGATA TACCATACAA TGGAATATAT TATGATCCTC
3001 CCGAAGAGGT ATTTTACTTC ATCTTCTGTG CTTTTAGATT TCAGATATTT TTATTAGAAG
3061 AAAATTATGA TTTTTTCCCT CACGAACCTT CCCAATTGCT ATTTCAAGCT GTCCTACTTA
3121 TTTGCTGCTG GCATCTTATT TTTCTATTCT CTAACCAGTT ATGAAATTCC TTACATGCAT
3181 ATGCAGGAGA AGTATGTATT CAAGCATCCT CAACCTAAAC GACCAAAATC ATTGCGGATA
3241 TATGAAACAC ATGTTGGCAT GAGTAGCCCG GTATTTTCATC TTTACCATGT ATTCCATAAA
3301 TGAAGTTAGC TATATGCAGT TCAAATTTAT TTACAGGTTG TTACAATGGT ATTTTTGTGT
3361 TGGTGCCCTT CTTTCGTTTT ATAAGTAAAA AACTTATCAT AAATTTATTT GTTATGCCGC
3421 TTGGTTAATA CAATCTGAAA AATGTAAGTG TGGACAATCT AGAACTAGAT AATACAAATC
3481 TGAAAAACA TGCTGGAATA GTGTCATTTT AGTCAACTAG GATGTTTTGA ATGCTCAAGA
3541 GAAGTACTAG TGTGTAGCAT CAAAAGCTGG TGTCCATTTG TTCAAATGTT TAATTAACAC
3601 TATAGTGAAA ACAAGTAATT GCACAAAGAA ACAAGTAATT GCCCAAGTTC ATATGTTTTT

```

Figure 8

3661 TCACTATATT ACATGTTTCA TCAACAATTT AATTAACCTC ATTCCTTACA AACATTTGTA
3721 TTTACATTTG TTCCTACATA TATAGTTATT TTATATATCA ACTTTATAAA TCATGACTGT
3781 TATAATTAAA ACCGATGGTA TATCAACGAT TGAGATAATT TGGCATATGT GGATGAATTT
3841 TGTGGCTTGT TATGCTCTTG TTTTAATAAC ATAATAAATA GATTATGCTT GTTGGTAGCC
3901 TTTTACATT AACACATGGG CAATTACTTG TTTCTTTGTG CAACCAGGAA CCAAAGATCG
3961 AG

//

Sequence of a wheat branching enzyme IIb cDNA

```

1  ATGGTCGACC TGCAGGCGGC CGCGAATGCA CTAGNGATTT TGACACCAGA
51  CCAACTGGTA ATGGTAGCGA CCGGCGCTCA GCTGGAATTC GCGGCCGCGT
101 CGACCGTGGG TTAAAGCAGG AGACGAGGCG GGGTCAGTTG GGCAGTTAGG
151 TTGGATCCGA TCCGGCTGCG GCGGCGGCGA CGGGATGGCT GCGCCGGCAT
201 TCGCAGTTTC CGCGGCGGGG CTGGCCCGGC CGTCGGCTCC TCGATCCGGC
251 GGGGCAGAGC GGAGGGGGCG CGGGGTGGAG CTGCAGTCGC CATCGCTGCT
301 CTTCCGGCCG AACAAGGGCA CCCGTTCCAC CCGTGCCGTC GCGGTCGGAG
351 GTTCTGGATG GCGCGTGGTC ATGCGCGCGG GGGGGCCGTC CGGGGAGGTG
401 ATGATCCCTG ACGGCGGTAG TGGCGGAACA CCGCCTTCCA TCGACGGTCC
451 CGTTCAGTTC GATTCTGATG ATCTGAAGGT TCCATTTCATT GATGATGAAA
501 CAAGCCTACA GGATGGAGGT GAAGATAGTA TTTGGTCTTC AGAGACAAAT
551 CAGGTTAGTG AAGAAATTGA TGCTGAAGAC ACGAGCAGAA TGGACAAAGA
601 ATCATCTACG AGGGAGAAAT TACGCATTCT GCCACCACCG GGAAATGGAC
651 AGCAAATATA CGAGATTGAC CCAACGCTCC GAGACTTTAA GTACCATCTT
701 GAGTATCGAT ATAGCCTATA CAGGAGAATA CGTTCAGACA TTGATGAACA
751 CGAAGGAGGC ATGGATGTAT TTTCCCGCGG TTACGAGAAG TTTGGATTTA
801 TGCGCAGCGC TGAAGGTATC ACTTACCGAG AATGGGCTCC TGGAGCAGAT
851 TCTGCAGCAT TAGTTGGCGA CTTCAACAAT TGGGATCCAA ATGCAGACCA
901 TATGAGCAA AATGACCTTG GTGTTTGGGA GATTTTCTG CCAAACAATG
951 CAGATGGTTC GCCACCAATT CCTCACGGCT CACGGGTGAA GGTGCGAATG
1001 GGTACTCCAT CTGGGACAAA GGATTCAATT CCTGCTTGGA TCAAGTACTC
1051 CGTGCAGACT CCAGGAGATA TACCATACAA TGGAATATAT TATGATCCTC
1101 CCGAAGAGGA GAAGTATGTA TTCAAGCATC CTCAACCTAA ACGACCAAAA
1151 TCATTGCGGA TATATGAAAC ACATGTTGGC ATGAGTAGCC CGGAACCAAA
1201 GATCAACACA TATGCAAAC TCAAGGATGA GGTGCTTCCA AGAATTAAAA
1251 GACTTGGATA CAATGCAGTG CAAATAATGG CAATCCAAGA GCACTCATA
1301 TATGGAAGCT TTGGGTACCA TGTTACCAAT TTCTTTGCAC CAAGTAGCCG
1351 TTTTGGGTCC CCAGAAGATT TAAATCTTT GATTGATAGA GCTCACGAGC
1401 TTGGCTTGGT TGTCTCATG GATGTTGTT ACAGTCACGC GTCAAATAAT
1451 ACCTTGGACG GGTGGAATGG TTTTGATGGC ACGGATACAC ATTACTTCCA
1501 TGGCGGTTC CCGGGCCATC ACTGGATGTG GGATTCCCGT GTGTTTAACT
1551 ATGGAATAA GGAAGTTATA AGGTTTCTAC TTTCCAATGC AAGATGGTGG
1601 CTAGAGGAGT ATAAGTTTGA TGGTTTCCGA TTCGATGGCG CGACCTCCAT
1651 GATGTATACC CATCATGGAT TACAAGTAAC CTTTACAGGA AGCTACCATG
1701 AATATTTTGG CTTTGCCACT GATGTAGATG CGGTCGTTTA CTTGATGCTG
1751 ATGAATGATC TAATTCATGG GTTTTATCCT GAAGCCGTAA CTATCGGTGA
1801 AGATGTTAGT GGAATGCCTA CATTTGCCCT TCCTGTTCAA GTTGGTGGGG
1851 TTGGTTTGA CTATCGCTTA CATATGGCTG TTGCCCAGCA ATGGATTGAA
1901 CTTCTCAAAG GAAACGATGA AGCTTGGGAG ATGGGTAATA TTGTGCACAC
1951 ACTAACAAC AGAAGGTGGC TGGAAAAGTG TGTTACTTAT GCTGAAAGTC
2001 ACGATCAAGC ACTTGTTGGA GACAAGACTA TTGCATTCTG GTTGATGGAC
2051 AAGGATATGT ATGATTTTCAT GCGCTGAAC GGACCTTCGA CGCCTAATAT
2101 TGATCGTGGA ATAGCACTGC ATAAAATGAT TAGACTTATC ACAATGGGTC
2151 TAGGAGGAGA GGGTTATCTT AACTTTATGG GAAATGAGTT CGGGCATCCT
2201 GAATGGATAG ACTTTCCAAG AGGCCACAA GTACTTCAA GTGGTAAGTT
2251 CATCCCAGGA AACAACAACA GTTACGACAA ATGCCGTCGA AGATTTGACC
2301 TGGGTGATGC AGAATTTCTT AGGTATCATG GTATGCAGCA GTTTGATCAG
2351 GCAATGCAGC ATCTTGAGGA AAAATATGGT TTTATGACAT CAGACCACCA
2401 GTACGTATCT CGGAAACATG AGGAAGATAA GGTGATCGTG TTTGAAAAG
2451 GGGACTTGGT ATTTGTGTTT AACTTCCACT GGAGTAGTAG CTATTTGAC
2501 TACCGGGTCG GCTGTTTAAA GCCTGGGAAG TACAAGGTGG TCTTAGACTC
2551 GGACGCTGGA CTCTTTGGTG GATTGGTAG GATCCATCAC ACTGCAGAGC
2601 ACTTCACTTC TGACTGCCAA CATGACAACA GGCCCCATTC ATTCTCAGTG
2651 TACACTCCTA GCAGAACCCTG TGTGTCTAT GCTCCAATGA ACTAACAGCA
2701 AAGTGCAGCA TACGCGTGCG CGCTGTTGTT GCTAGTAGCA AGAAAAATCG
2751 TATGGTCAAT ACAACCAGGT GCAAGGTTTA ATAAGGATTT TTGCTTCAAC
2801 GAGTCCTGGA TAGACAAGAC AACATGATGT TGTGCTGTGT GCTCCCAATC
2851 CCCAGGGCGT TGTGAAGAAA ACATGCTCAT CTGTGTTATT TTATGGATCA
2901 GCGACGAAAC CTCCCCCAA TACCCTTTT TTTTNNAAA GGAGGATAGG
2951 CCCCCGNCT TTGCNTNN

```

Figure 9

Alignment of Cereal Branching Enzyme Sequences

	1				50
Y11282	--MATFAVSG	ATLGVARPAG	AGGGLLPRSG	SERRGGVDLP	SLLLRKKDS.
sbe9	--MATFAVSG	ATLGVARPPA	A.....
barley BEIIa	-----	-----	-----	-----	-----
maize BEIIa	-----	-----	-----	-----DLP	SVLFRRKDAF
rice BEIV	--MASFAVSG	ARLGVVRAGG	GGGG..GGGP	AARSGGVDLP	SVLFRRKDSF
barley BEIIB	MAAPAFAV..	SAAGIARPSA	R...RSSGAE	PR.....	.SLLFGRNKG
wheat BEIIB	MAAPAFAV..	SAAGLARPSA	P...RSGGAE	RRGRGVELQS	PSLLFGRNKG
maize BEIIB	-----	-----	-----	-----	-----
rice BEIII	MAAPASAVPG	SAAGLRAGAV	RFPVPAGARS	WRAAAELPTS	RSLLSGR...
	51				100
Y11282	SR.....AVLSR	AASPGKVLVP	DGESDDLASP	A.....Q
sbe9	A.....Q
barley BEIIa	-----	-----	-----	-----	-----
maize BEIIa	SR.....TVLSC	AGAPGKVLVP	GGGSDDLSS	AEPVVD..Q
rice BEIV	SR.....GVVSC	AGAPGKVLVP	GGGSDDLSS	AEPDVETQEQ
barley BEIIB	TRFPRAVGVG	GSGWRVVMRA	GGPSGEVMIP	DGGSGGSGTP	PSIEGSVQFE
wheat BEIIB	TRSPRAVGVG	GSGWRVVMRA	GGPSGEVMIP	DGGSG..GTP	PSIDGPVQFD
maize BEIIB	-----	-----	-----KAVMVP	EGENDGL...	ASRADSAQFQ
rice BEIII	.RFPGA VRVG	GSGGRVAVRA	AGASGEVMIP	EGESDGM...	PVSAG.....
	101				150
Y11282	PEELQIPEDI	EEQ.....TAEV	NMTGGTAEKL
sbe9	PEELQIPEDI	EEQ.....TAEV	NMTGGTAEKL
barley BEIIa	-----	-GE.....MAEV	NMTGGAAEKL
maize BEIIa	PEELQIP...EAE	LTVEKTSSSP	TQTTSAAVEA	SSGVEAEERP
rice BEIV	PEESQIPDDN	KVKPFEEEE	I.....PAVEA	SIKVVAEDKL
barley BEIIB	SDDLVPFID	D.....EP	SLHDGGEDTI
wheat BEIIB	SDDLKVPFID	D.....ET	SLQDGGEDSI
maize BEIIB	SDELEVPDIS	E.....E.	.TTCGA....
rice BEIII	SDDLQLPALD	D.....EL	STEVGAEEVEI
	151				200
Y11282	ESSEPTQGIV	ETITDGV...T	KGVKELVVGE	KPRVVPKPGD
sbe9	ESSEPTQGIV	ETITDGV...T	KGVKELVVGE	KPRVVPKPGD
barley BEIIa	ESSEPTQGIA	ETITDGV...T	KGVKELVVGE	KPVVVPKPGD
maize BEIIa	ELSEVI....GVGGT	GGTKIDGAGI	K.AKAPLVEE	KPRVIPPPGD
rice BEIV	ESSEVIQDIE	ENVTEGV...I	KDADEPTVED	KPRVIPPPGD
barley BEIIB	RSSETYQVTE	EIDAEGVSRM	D.....	...KESSTVK	KIRIVPQPGN
wheat BEIIB	WSSETNQVSE	EIDAEDTSRM	D.....	...KESSTRE	KLRILPPPGN
maize BEIIB	...GVA....	..DAQALNRVRVVPPPSD
rice BEIII	ESSGAS....	..DVEGVKRV	V.....	...EELAAEQ	KPRVVPPTGD
	201				250
Y11282	GQKIYEIDPT	LKDFRSHLDY	RYSEYRRIRA	AIDQHEGGLE	AFSRGYEKL
sbe9	GQKIYEIDPT	LKDFRSHLDY	RYSEYRRIRA	AIDQHEGGLE	AFSRGYEKL
barley BEIIa	GQKIYEIDPT	LKDFRSHLDY	RYSEYKRIRA	AIDQHEGGLE	VFSRGYEKL
maize BEIIa	GQRIYEIDPM	LEGFRGHLDY	RYSEYKRLRA	AIDQHEGGLD	AFSRGYEKL
rice BEIV	GQKIYQIDPM	LEGFRNHLDY	RYSEYKRMRA	AIDQHEGGLD	AFSRGYEKL
barley BEIIB	GQKIYDIDPM	LRDFKYHLEY	RYSLYRRIRS	DIDEYDGGMD	VFSRGYEKFG
wheat BEIIB	GQKIYEIDPT	LRDFKYHLEY	RYSLYRRIRS	DIDEHEGGMD	VFSRGYEKFG
maize BEIIB	GQKIFQIDPM	LQGYKYHLEY	RYSLYRRIRS	DIDEHEGGLE	AFSRGYEKF
rice BEIII	GQKIFQMDSM	LNGYKYHLEY	RYSLYRLRS	DIDQYEGGLE	TFSRGYEKF
	251				300
Y11282	FTRSAEGITY	REWAPGAHSA	ALVGDFNNWN	PNADTMTRDD	YGVWEIFLPN
sbe9	FTRSAEGITY	REWAPGAHSA	ALVGDFNNWN	PNADTMTRDD	YGVWEIFLPN
barley BEIIa	FTRSAEGITY	REWAPGAHSA	ALVGDFNNWN	PNADTMTRDD	YGVWEIFLPN
maize BEIIa	FTRSAEGITY	REWAPGAYSA	ALVGDFNNWN	PNADAMARNE	YGVWEIFLPN
rice BEIV	FTRSAEGITY	REWAPGAQSA	ALVGDFNNWN	PNADTMTRNE	YGVWEISLPN
barley BEIIB	FVRSAGITY	REWAPGADSA	ALVGDFNNWD	PTADHMSKND	LGIWEIFLPN
wheat BEIIB	FMRSAEGITY	REWAPGADSA	ALVGDFNNWD	PNADHMSKND	LGVWEIFLPN
maize BEIIB	FNASAEGITY	REWAPGAFSA	ALVGDFNNWD	PNADRMSKNE	FGVWEIFLPN
rice BEIII	FNHSAEGITY	REWAPGAHSA	ALVGDFNNWN	PNADRMSKNE	FGVWEIFLPN

Figure 10

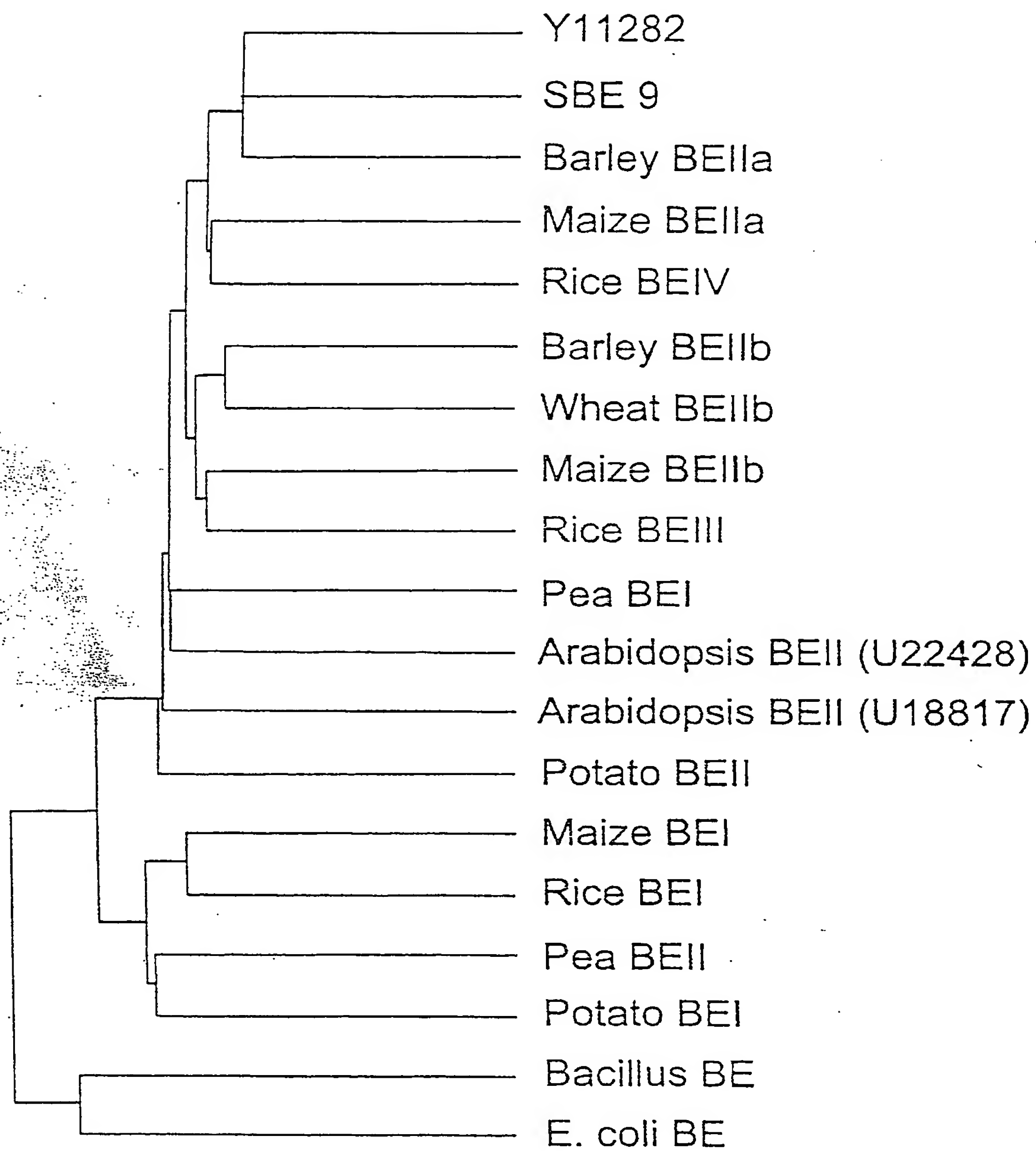
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sbe9	NADGSPAIPH	GSRVKIRMDT	PSGVKDSISA	WIKFSVQAPG	EIPFNGIYYD
barley BEIIa	NADGSPAIPH	GSRVKIRMDT	PSGVKDSISA	WIKFSVQAPG	EIPFNGIYYD
maize BEIIa	NADGSPAIPH	GSRVKIRMDT	PSGVKDSIPA	WIKFSVQAPG	EIPYNGIYYD
rice BEIV	NADGSPAIPH	GSRVKIRMDT	PSGVKDSIPA	WIKFAVQAPG	EIPYNGIYYD
barley BEIIb	NADGSPPIPH	GSRVKVRMDT	PSGTKDSIPA	WIKYSVQTPG	DIPYNGIYYD
wheat BEIIb	NADGSPPIPH	GSRVKVRMDT	PSGTKDSIPA	WIKYSVQTPG	DIPYNGIYYD
maize BEIIb	NADGTSPIPH	GSRVKVRMDT	PSGIKDSIPA	WIKYSVQAPG	EIPYDGIYYD
rice BEIII	NADGSSPIPH	GSRVKVRMET	PSGIKDSIPA	WIKYSVQAAG	EIPYNGIYYD
	351				400
Y11282	PPEEEKYVFQ	HPQPKRPESL	RIYESHIGMS	SPEPKINSYA	NFRDEVLPRI
sbe9	PPEEEKYVFQ	HPQPKRPESL	RIYESHIGMS	SPEPKINSYA	NFRDEVLPRI
barley BEIIa	PPEEEKYVFQ	HPQPKRPESL	RIYESHIGMS	SPEPKINSYA	NFRDEVLPRI
maize BEIIa	PPEEEKYVFK	HPQPKRPKSL	RIYESHVGMS	SPEPKINTYA	NFRDEVLPRI
rice BEIV	PPEEEKYVFQ	HPQPKRPNSL	RIYESHIGMS	SPEPKINTYA	NFRDEVLPRI
barley BEIIb	PPEEEKYVFK	HPQPKRPKSL	RIYETHVGMS	SPEPKINTYA	NFRDEVLPRI
wheat BEIIb	PPEEEKYVFK	HPQPKRPKSL	RIYETHVGMS	SPEPKINTYA	NFRDEVLPRI
maize BEIIb	PPEEVKYVFR	HAQPKRPKSL	RIYETHVGMS	SPEPKINTYV	NFRDEVLPRI
rice BEIII	PPEEEKYIFK	HPQPKRPKSL	RIYETHVGMS	STEPKINTYA	NFRDEVLPRI
	401				450
Y11282	KRLGYNAVQI	MAIQEHSSYYA	SFGYHVTNFF	APSSRFGTPE	DLKSLIDRAH
sbe9	KRLGYNAVQI	MAIQEHSSYYA	SFGYHVTNFF	APSSRFGTPE	DLKSLIDRAH
barley BEIIa	KRLGYNAVQI	MAIQEHSSYYA	SFGYHVTNFF	APSSRFGTPE	DLKSLIDRAH
maize BEIIa	KKLGYNAVQI	MAIQEHSSYYA	SFGYHVTNFF	APSSRFGTPE	DLKSLIDKAH
rice BEIV	KKLGYNAVQI	MAIQEHSSYYA	SFGYHVTNFF	APSSRFGTPE	DLKSLIDKAH
barley BEIIb	KRLGYNAVQI	MAIQEHSSYYG	SFGYHVTNFF	APSSRFGSPE	DLKSLIDRAH
wheat BEIIb	KRLGYNAVQI	MAIQEHSSYYG	SFGYHVTNFF	APSSRFGSPE	DLKSLIDRAH
maize BEIIb	KKLGYNAVQI	MAIQEHSSYYG	SFGYHVTNFF	APSSRFGTPE	DLKSLIDRAH
rice BEIII	KKLGYNAVQI	MAIQEHAYYG	SFGYHVTNFF	APSSRFGTPE	DLKSLIDKAH
	451				500
Y11282	ELGLLVLM DI	VHSHSSNNTL	DGLNGFDGTD	THYFHGGPRG	HHWMWDSRLF
sbe9	ELGLLVLM DI	VHSHSSNNTL	DGLNGFDGTD	THYFHGGPRG	HHWMWDSRLF
barley BEIIa	ELGLLVLM DI	VHSHSSNNTL	DGLNGFDGTD	THYFHGGPRG	HHWMWDSRLF
maize BEIIa	ELGLLVLM DI	VHSHSSNNTL	DGLNGFDGTD	THYFHGGPRG	HHWMWDSRLF
rice BEIV	ELGLLVLM DI	VHSHASNNTL	DGLNGFDGTD	THYFHGGPRG	HHWMWDSRLF
barley BEIIb	ELGLLVLM DV	VHSHASNNTL	DGLNGFDGTD	THYFHGGSRG	HHWMWDSRVF
wheat BEIIb	ELGLVLM DV	VHSHASNNTL	DGLNGFDGTD	THYFHGGSRG	HHWMWDSRVF
maize BEIIb	ELGLLVLM DV	VHSHASNNTL	DGLNGFDGTD	THYFHSGPRG	HHWMWDSRLF
rice BEIII	ELGLVLM DV	VHSHASNNTL	DGLNGFDGTD	THYFHSGSRG	HHWMWDSRLF
	501				550
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sbe9	NYGSWEVLR F	LLSNARWWLE	EYKFDGFRFD	GVTSM MYTHH	GLQMTFTGNY
barley BEIIa	NYGSWEVLR F	LLSNARWWLE	EYKFDGFRFD	GVTSM MYTHH	GLQMTFTGNY
maize BEIIa	NYGSWEVLR F	LLSNARWWLE	EYKFDGFRFD	GVTSM MYTHH	GLQVTFTGNY
rice BEIV	NYGSWEVLR Y	LLSNARWWLE	EYKFDGFRFD	GVTSM MYTHH	GLQVAF TGN Y
barley BEIIb	NYGNKEVIR F	LLSNARWWLE	EYKFDGFRFD	GATSM MYTHH	GLQVTFTGS Y
wheat BEIIb	NYGNKEVIR F	LLSNARWWLE	EYKFDGFRFD	GATSM MYTHH	GLQVTFTGS Y
maize BEIIb	NYGNWEVLR F	LLSNARWWLE	EYKFDGFRFD	GVTSM MYTHH	GLQVTFTGN F
rice BEIII	NYGNWEVLR F	LLSNARWWLE	EYKFDGFRFD	GVTSM MYTHH	GLQVAF TGN Y
	551				600
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sbe9	GEYFGFATDV	DAVVYLMLVN	DLIHGLHPDA	VSIGEDVSGM	PTFCIPVPDG
barley BEIIa	GEYFGFATDV	DAVVYLMLVN	DLIHGLYPDA	VSIGEDVSGM	PTFCIPVPDG
maize BEIIa	GEYFGFATDV	DAVVYLMLVN	DLIRGLYPEA	VSIGEDVSGM	PTFCIPVQDG
rice BEIV	GEYFGFATDV	DAVVYLMLVN	DLIHGLYPEA	VAIGEDVSGM	PTFCIPVQDG
barley BEIIb	HEYFGFATDV	DAVVYLMLVN	DLIHALYPEA	VTIGEDVSGM	PTFALPVQVG
wheat BEIIb	HEYFGFATDV	DAVVYLMLVN	DLIHGFYPEA	VTIGEDVSGM	PTFALPVQVG
maize BEIIb	NEYFGFATDV	DAVVYLMLVN	DLIHGLYPEA	VTIGEDVSGM	PTFALPVHDC
rice BEIII	SEYFGFATDA	DAVVYLMLVN	DLIHGLYPEA	ITIGEDVSGM	PTFALPVQDG

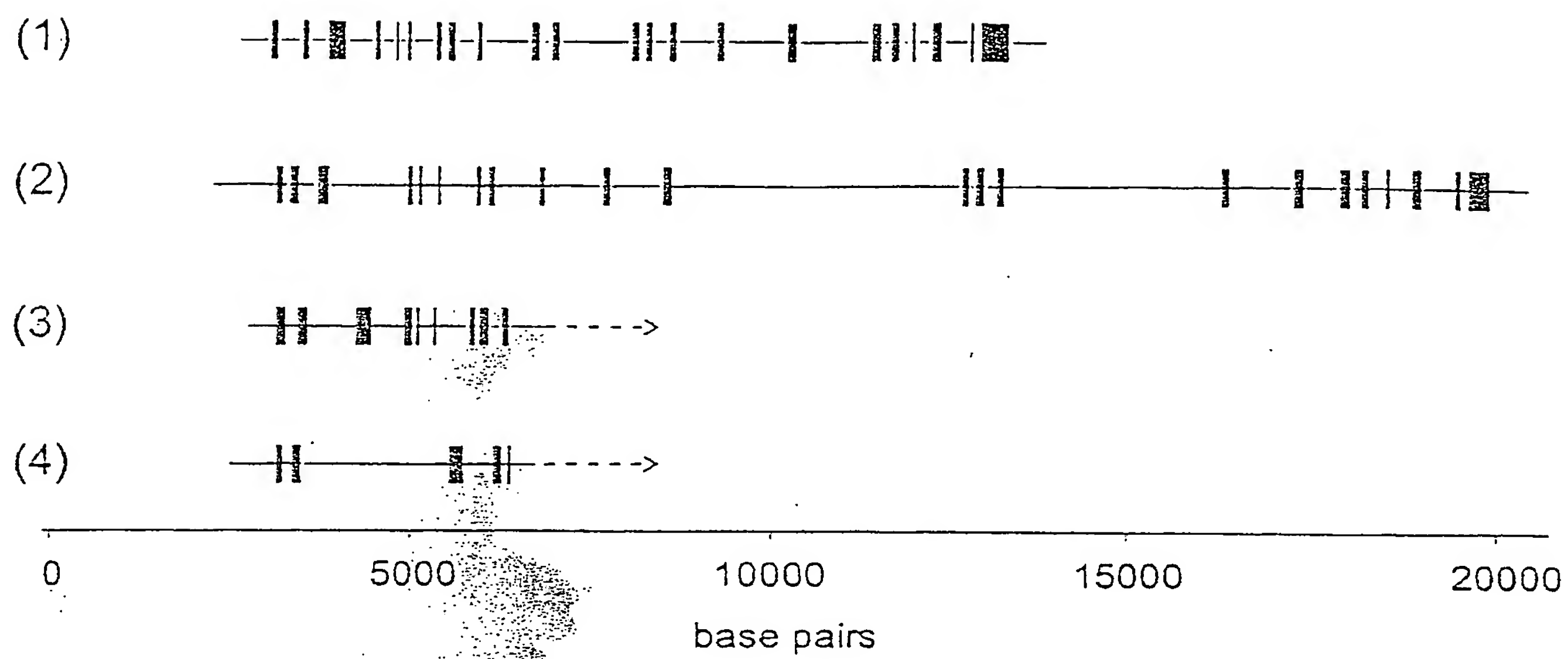
Figure 10 (cont'd)

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barley BEIIa	GVGFDYRLHM AVADKWIELL KQSDSWKMG	DIVHTLTNRR WLEKCVTYAE	
maize BEIIa	GVGFDYRLHM AVPDKWIELL KQSDSWKMG	DIVHTLTNRR WLEKCVTYAE	
rice BEIV	GVGFDYRLHM AVPDKWIELL KQSDSWKMG	DIVHTLTNRR WSEKCVTYAE	
barley BEIIb	GVGFDYRLHM AVADKWIELL KGSDEGWEMG	NIVHTLTNRR WLEKCVTYAE	
wheat BEIIb	GVGFDYRLHM AVARKWIELL KGNDEAWEMG	NIVHTLTNRR WLEKCVTYAE	
maize BEIIb	GVGFDYRMHM AVADKWIDLL KQSDSWKMG	DIVHTLTNRR WLEKCVTYAE	
rice BEIII	GVGFDYRLHM AVPDKWIELL KQSDSWKMG	DIVHTLTNRR WSEKCVTYAE	
	651		700
Y11282	SHDQALVGDK TIAFWLMDKD MYDFMALDRP	STPRIDRGIA LHKMIRLVTM	
sbe9	SHDQALVGDK TIAFWLMDKD MYDFMALDRP	STPRIDRGIA LHKMIRLVTM	
barley BEIIa	SHDQALVGDK TIAFWLMDKD MYDFMALDRP	STPRIDRGIA LHKMIRLVTM	
maize BEIIa	SHDQALVGDK TIAFWLMDKD MYDFMALDRP	STPRIDRGIA LHKMIRLVTM	
rice BEIV	SHDQALVGDK TIAFWLMDKD MYDFMALDRP	STPRIDRGIA LHKMIRLVTM	
barley BEIIb	SHDQALVGDK TIAFWLMDKD MYDFMALNGP	STPNIDRGIA LHKMIRLITM	
wheat BEIIb	SHDQALVGDK TIAFWLMDKD MYDFMALNGP	STPNIDRGIA LHKMIRLITM	
maize BEIIb	SHDQALVGDK TIAFWLMDKD MYDFMALDRP	STPTIDRGIA LHKMIRLITM	
rice BEIII	SHDQALVGDK TIAFWLMDKD MYDFMALDRP	ATPSIDRGIA LHKMIRLITM	
	701		750
Y11282	GLGGEGYLNFGMNEFGHPEW IDFPGRGPQTL	PTGKVLPGNN NSYDKCRRRF	
sbe9	GLGGEGYLNFGMNEFGHPEW IDFPGRGPQTL	PTGKVLPGNN NSYDKCRRRF	
barley BEIIa	GLGGEGYLNFGMNEFGHPEW IDFPGRGPQTL	PTGKVLPGNN NSYDKCRRRF	
maize BEIIa	GLGGEGYLNFGMNEFGHPEW IDFPGRGPQSL	PNGSVIPGNN NSFDKCRRRF	
rice BEIV	GLGGEGYLNFGMNEFGHPEW IDFPGRGPQSL	PNGSVLPGNN YSFDKCRRRF	
barley BEIIb	ALGGEGYLNFGMNEFGHPEW IDFPGRGPQVL	PTGKFIPGNN NSYDKCRRRF	
wheat BEIIb	GLGGEGYLNFGMNEFGHPEW IDFPGRGPQVL	PSGKFIPGNN NSYDKCRRRF	
maize BEIIb	GLGGEGYLNFGMNEFGHPEW IDFPGRGPQRL	PSGKFIPGNN NSYDKCRRRF	
rice BEIII	GLGGEGYLNFGMNEFGHPEW IDFPRAPQVL	PNGKFIPGNN NSYDKCRRRF	
	751		800
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sbe9	DLGDADFLRY HGMQEFDDQAM QHLEEKYGF	TSEHQYVSRK HEEDKVIIFE	
barley BEIIa	DLGDADFLRY RGMQEFDDQAM QHLEEKYGF	TSEHQYVSRK HEEDKVIIFE	
maize BEIIa	DLGDADFLRY RGMQEFDDQAM QHLEEKYGF	TSEHQYVSRK HEEDKVIIFE	
rice BEIV	DLGDADFLRY HGMQEFDDQAM QHLEEKYGF	TSEHQYVSRK HEEDKVIIFE	
barley BEIIb	DLGDADFLRY HGMQEFDDQAM QHLEEKYGF	TSEHQYVSRK HEEDKVIIFE	
wheat BEIIb	DLGDADFLRY HGMQEFDDQAM QHLEEKYGF	TSEHQYVSRK HEEDKVIIFE	
maize BEIIb	DLGDADFLRY HGMQEFDDQAM QHLEEKYGF	TSEHQYVSRK HEEDKVIIFE	
rice BEIII	DLGDADFLRY RGMLEFDDQAM QHLEEKYGF	TSEHQYVSRK HEEDKVIIFE	
	801		850
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barley BEIIa	RGDLVVFVNF HWSNSKXDYR VGCSRPGKYK	VALDSDDALF GGFSRLDHDV	
maize BEIIa	RGDLVVFVNF HWSNSYFDYR VGCSRPGKYK	IVLDSDDGLF GGFSRLDHDV	
rice BEIV	RGDLVVFVNF HWSNSYFDYR VGCSRPGKYK	IVLDSDDGLF GGFSRLDHDV	
barley BEIIb	KGDLVVFVNF HWSNSYFDYR VGCSRPGKYK	VVLDSDDGLF GGFSRLDHDV	
wheat BEIIb	KGDLVVFVNF HWSNSYFDYR VGCSRPGKYK	VVLDSDDGLF GGFSRLDHDV	
maize BEIIb	KGDLVVFVNF HWSNSYFDYR VGCSRPGKYK	VVLDSDDGLF GGFSRLDHDV	
rice BEIII	KGDLVVFVNF HWSNSYFDYR VGCSRPGKYK	VVLDSDDGLF GGFSRLDHDV	
	851		887
Y11282	DYFTTEHPHD NRPRSFSVYT PSRTAVVYAL	TE*----	
sbe9	DYFTTEHPHD NRPRSFSVYT PSRTAVVYAL	TE*----	
barley BEIIa	DYFTTEHPHD NRPRSFSVYT PSRTAVVYAL	TE*----	
maize BEIIa	EYFTADWPHD NRPCSFSVYA PSRTAVVYAP	AGAED*	
rice BEIV	EYFTADWPHD NRPCSFSVYT PSRTAVVYAL	..TED*	
barley BEIIb	EHFTNGCQHD NRPHSFSVYT PSRTCVCYAP	MN*----	
wheat BEIIb	EHFTSDCQHD NRPHSFSVYT PSRTCVCYAP	MN*----	
maize BEIIb	EHFTADCSHD NRPYSFSVYT PSRTCVCYAP	VE*----	
rice BEIII	EHFTADCSHD NRPYSFSVYS PSRTCVCYAP	AE*----	

Figure 10 (cont'd)

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*Figure 12*

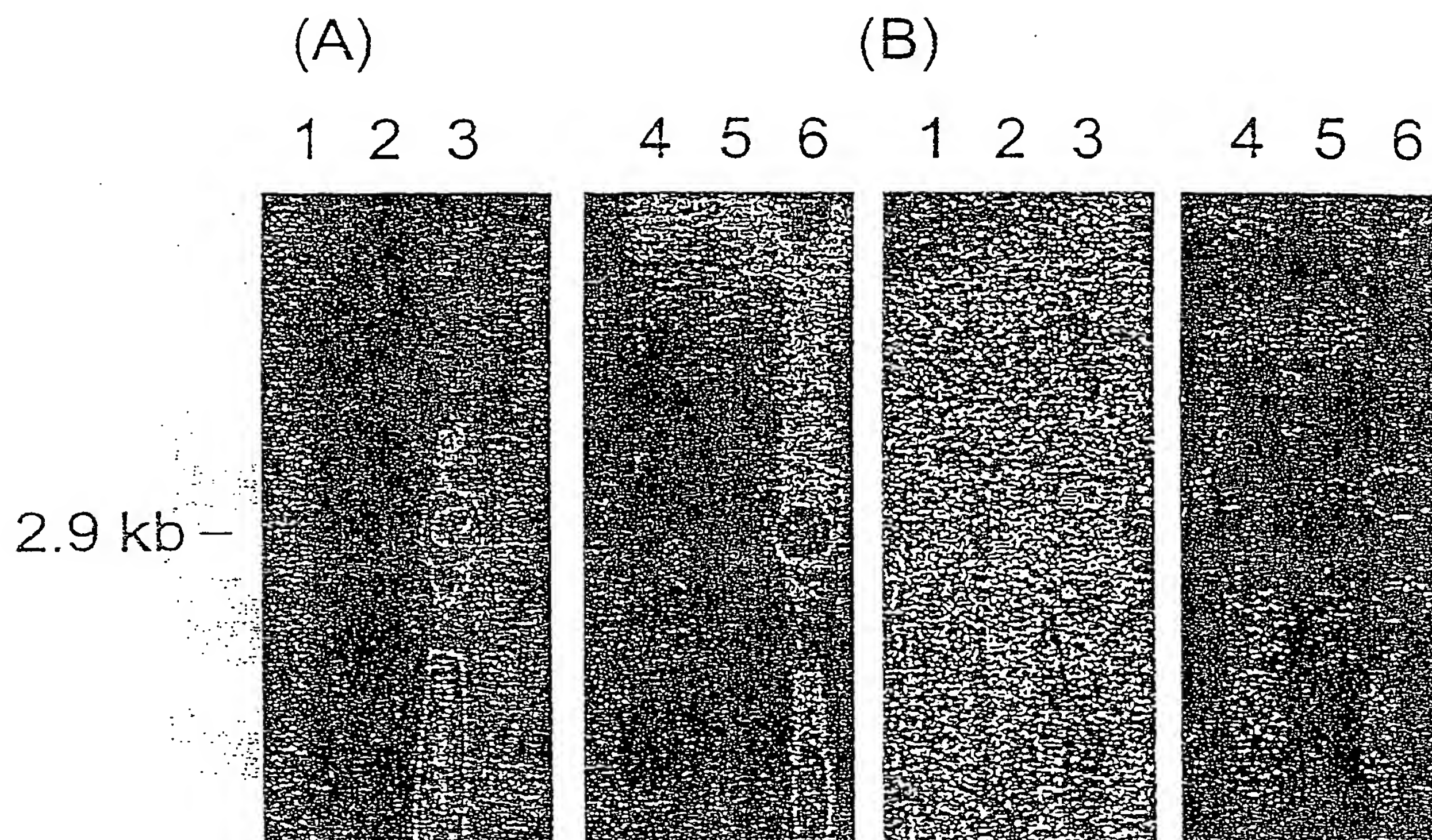


Figure 13

1 2

3 4 5

6 7 8

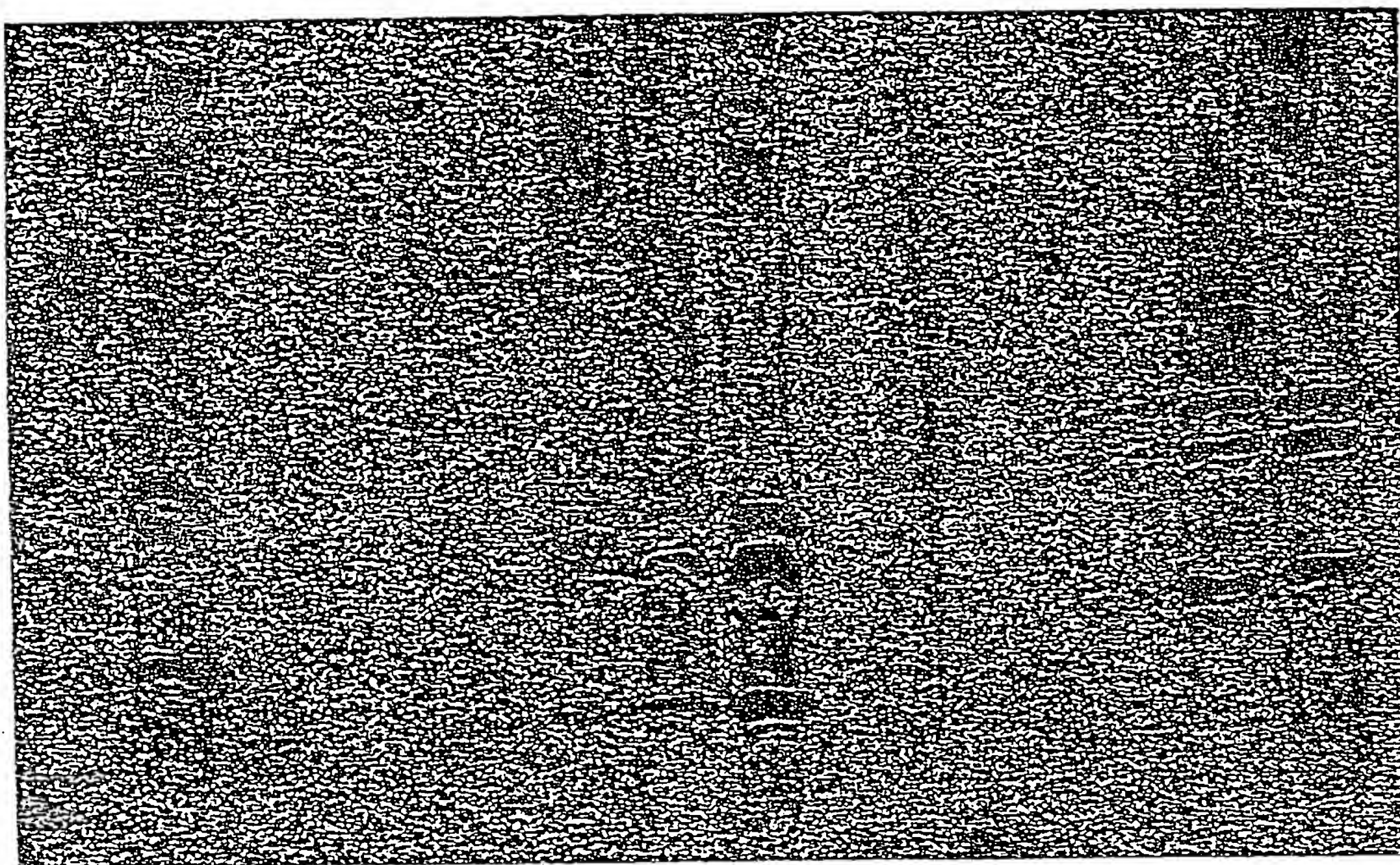
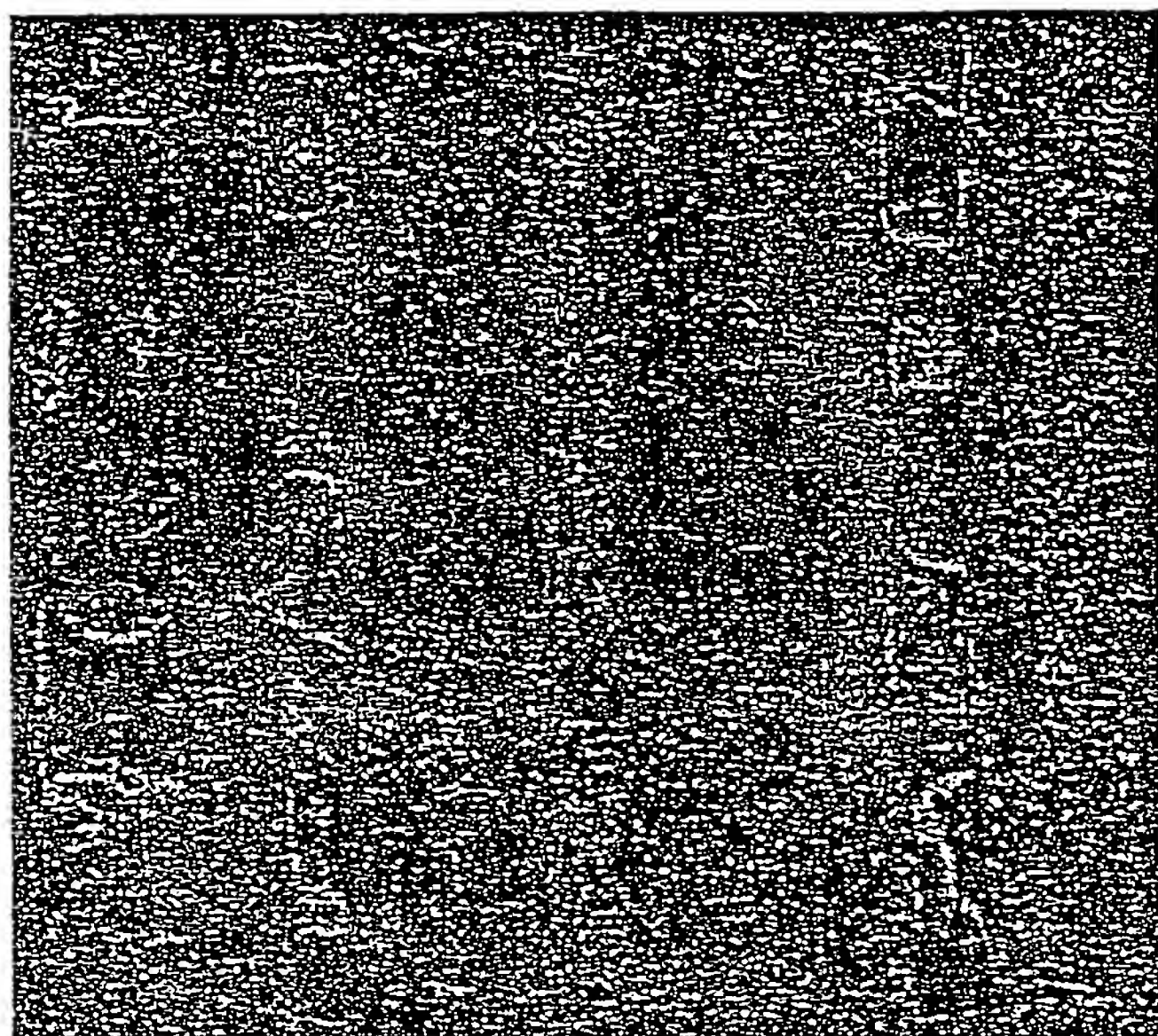


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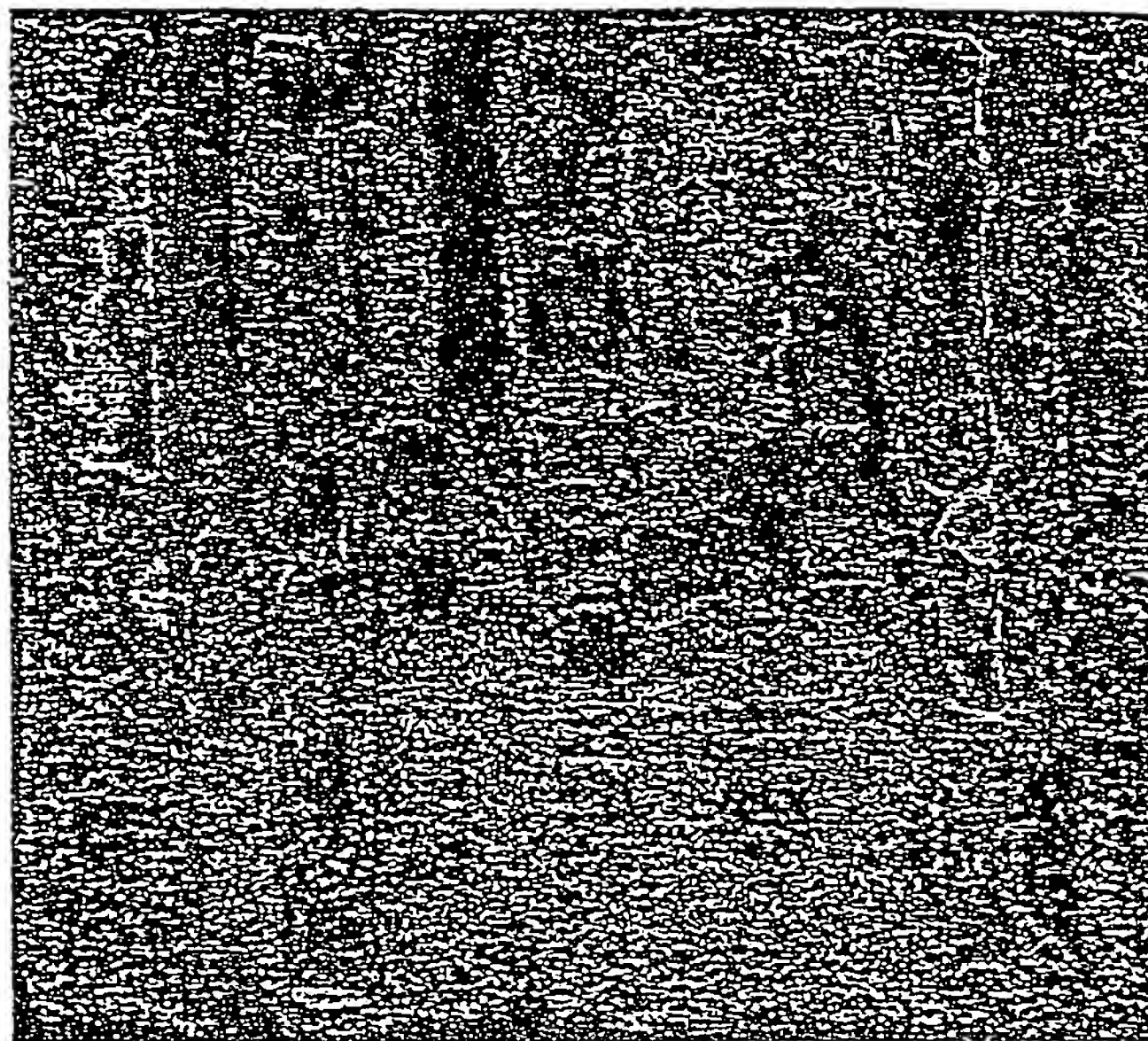
(A)

1 2 3 4 5 6 7 8 9 10 11 12

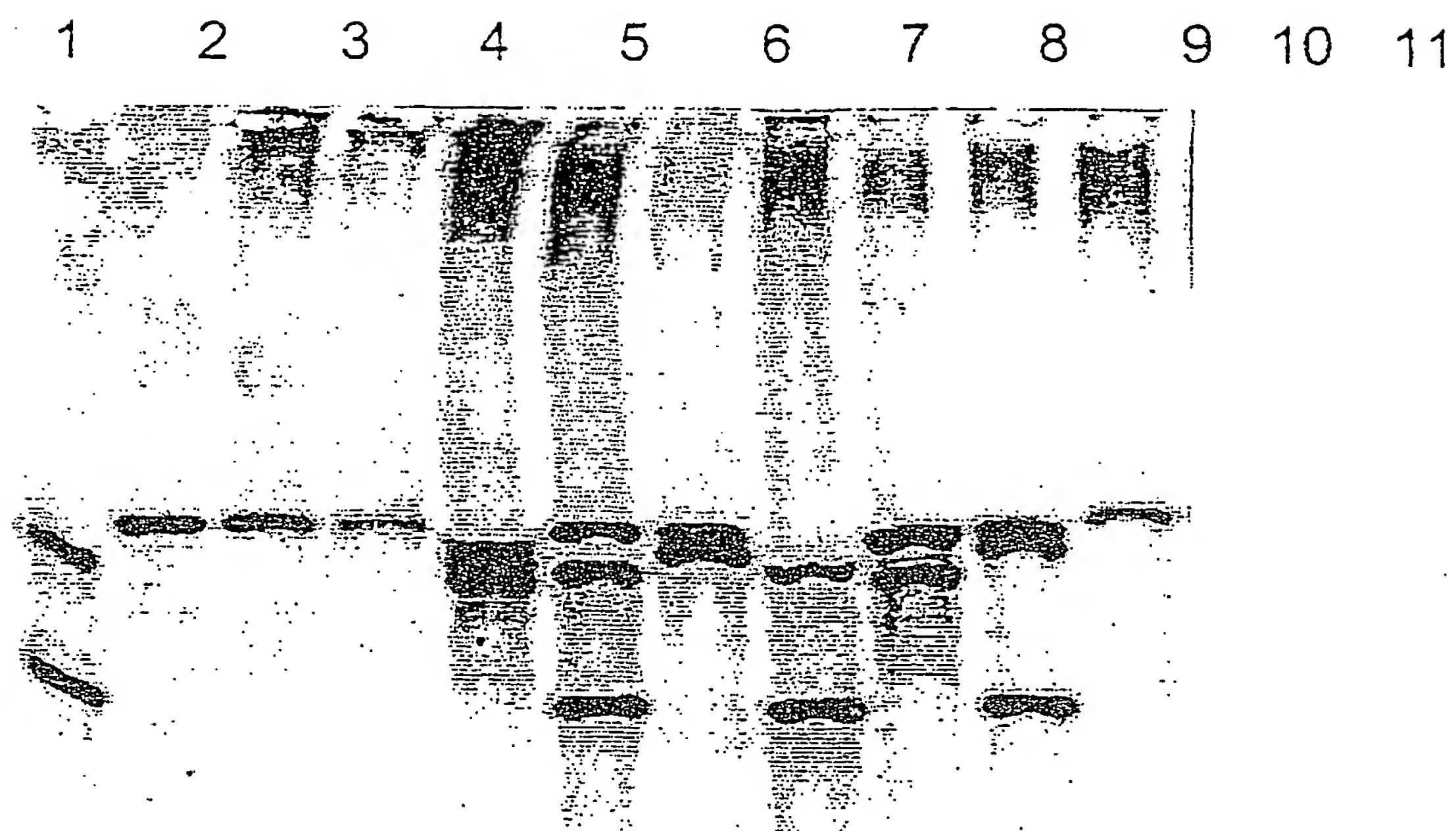


(B)

1 2 3 4 5 6 7 8 9 10 11 12

*Figure 15*

(A)



(B)

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19

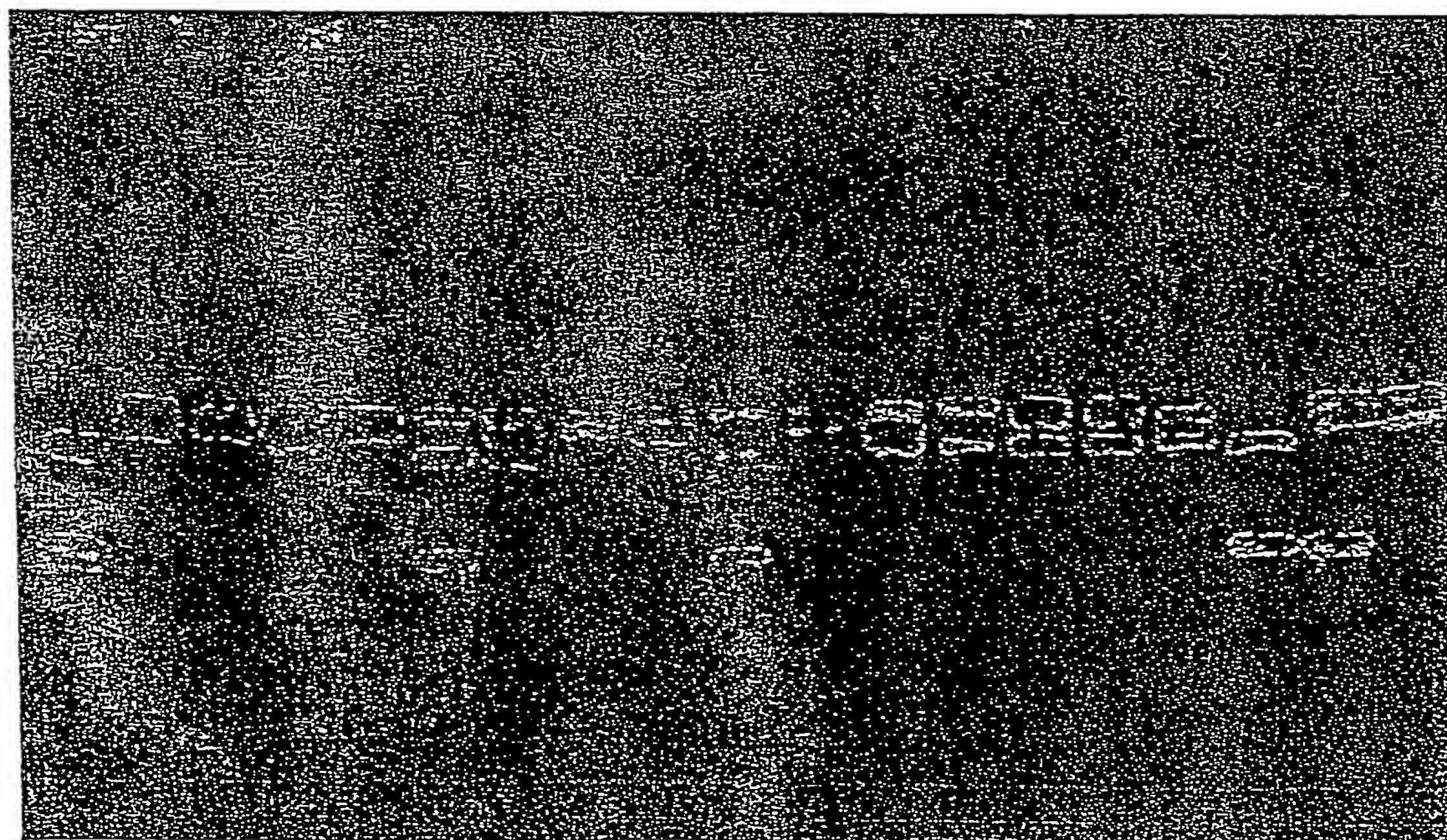
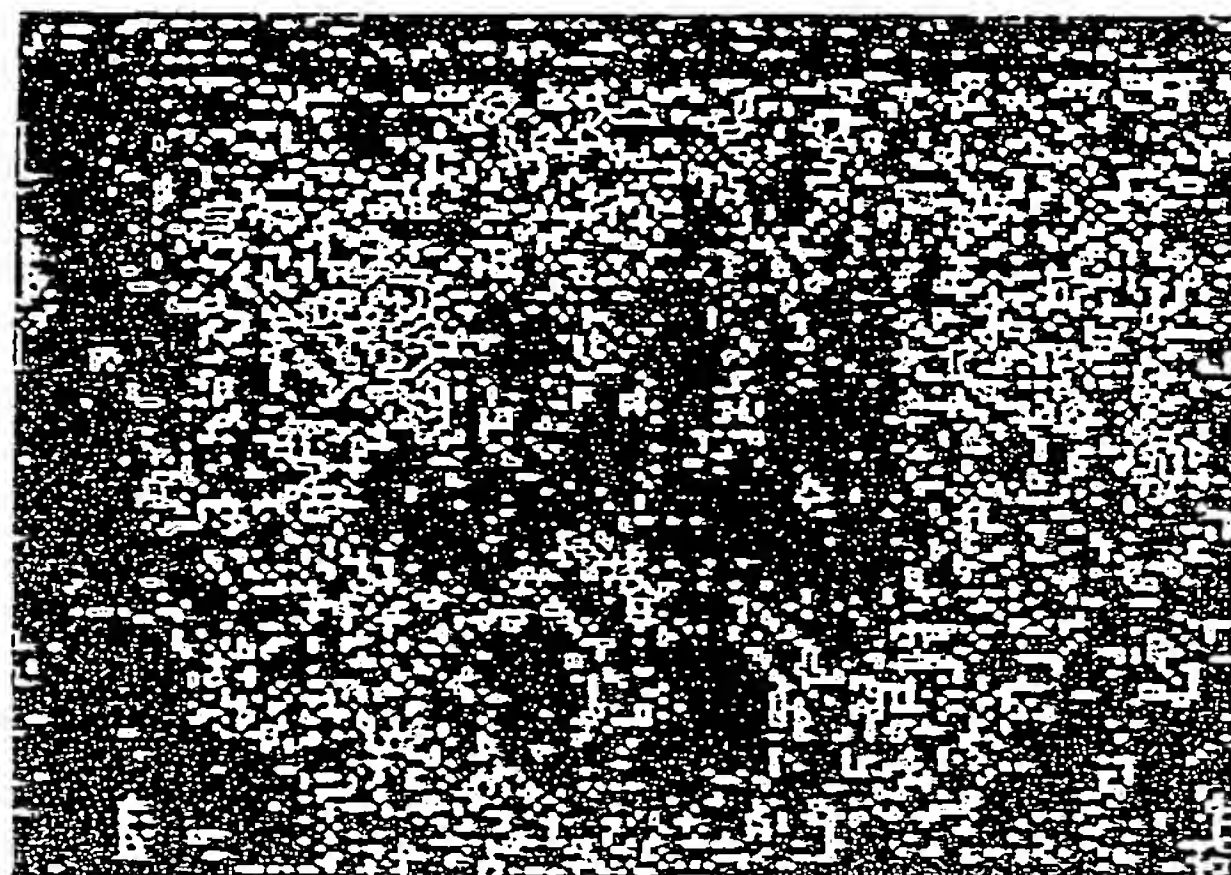


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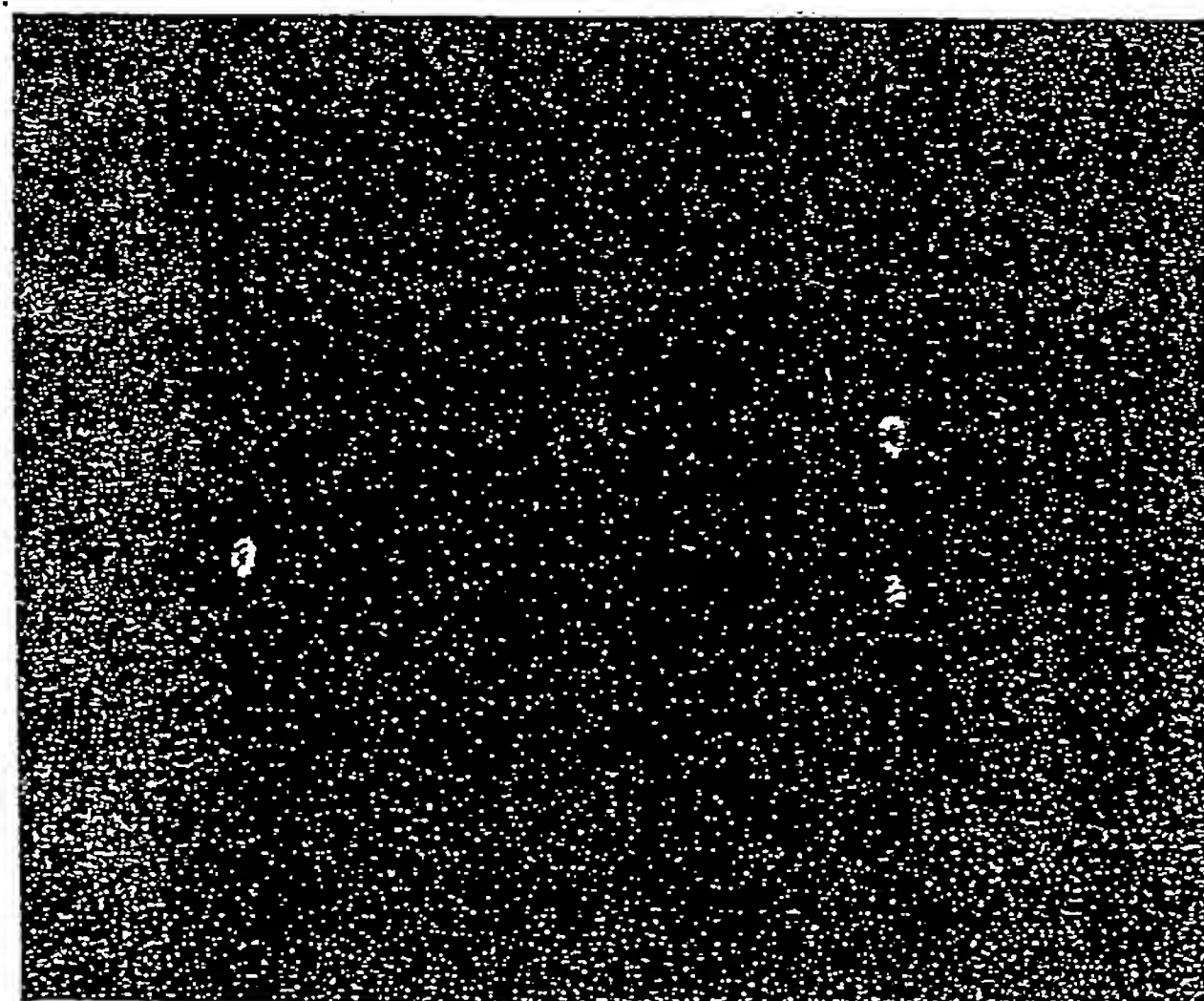
(A)



← 88 kD (U)

← 88 kD (L)

(B)



← 88 kD (U)

← 88 kD (L)

3KLH

R6

Figure 17

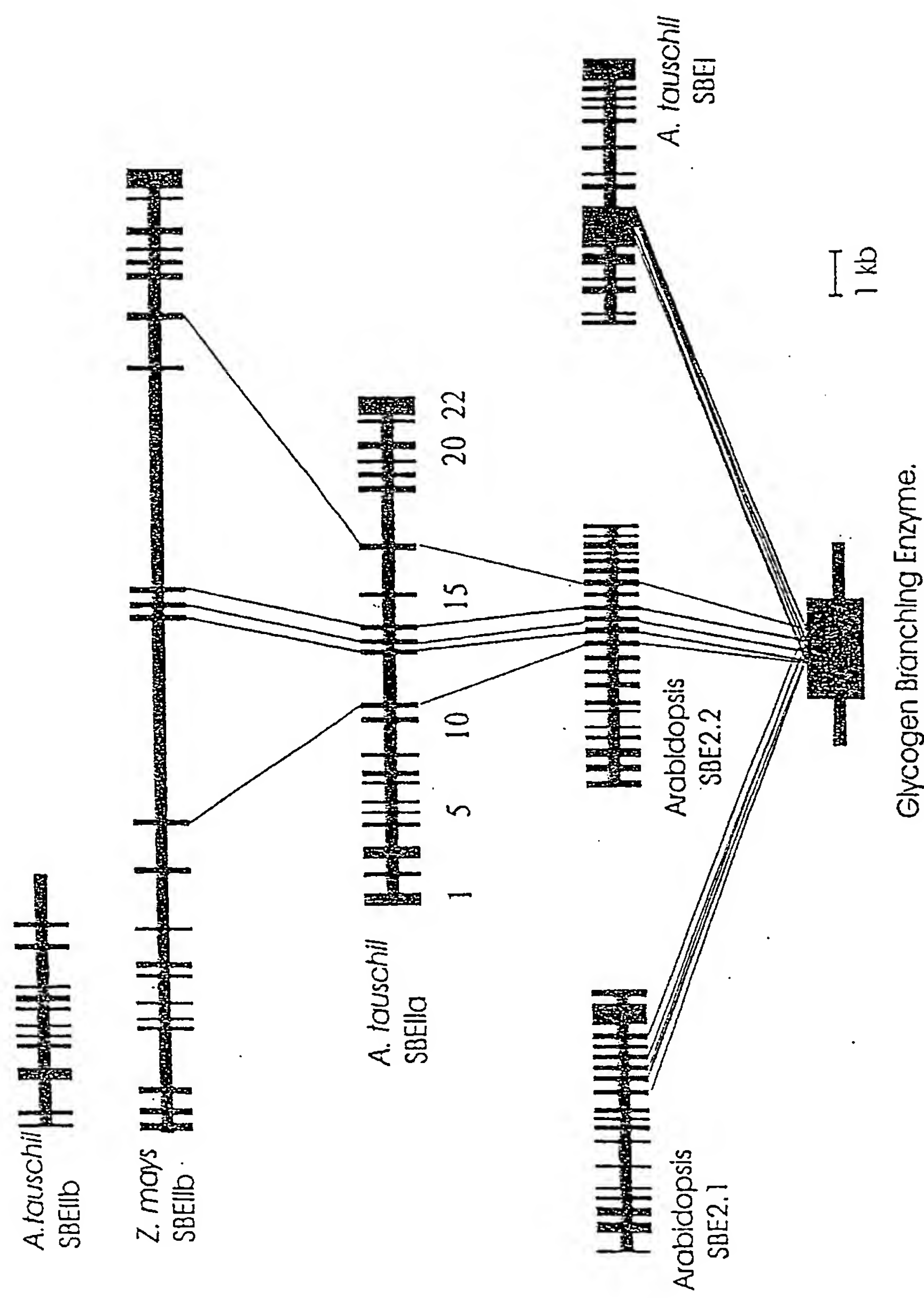


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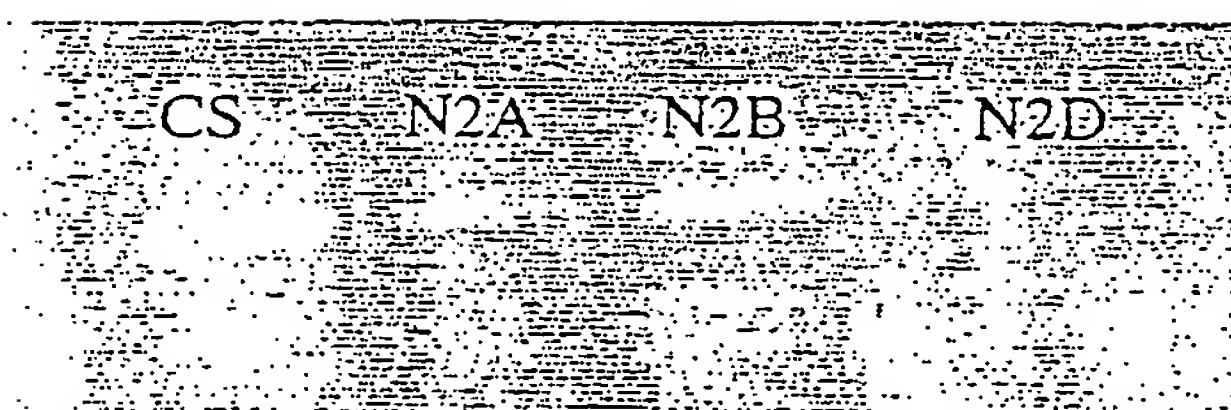


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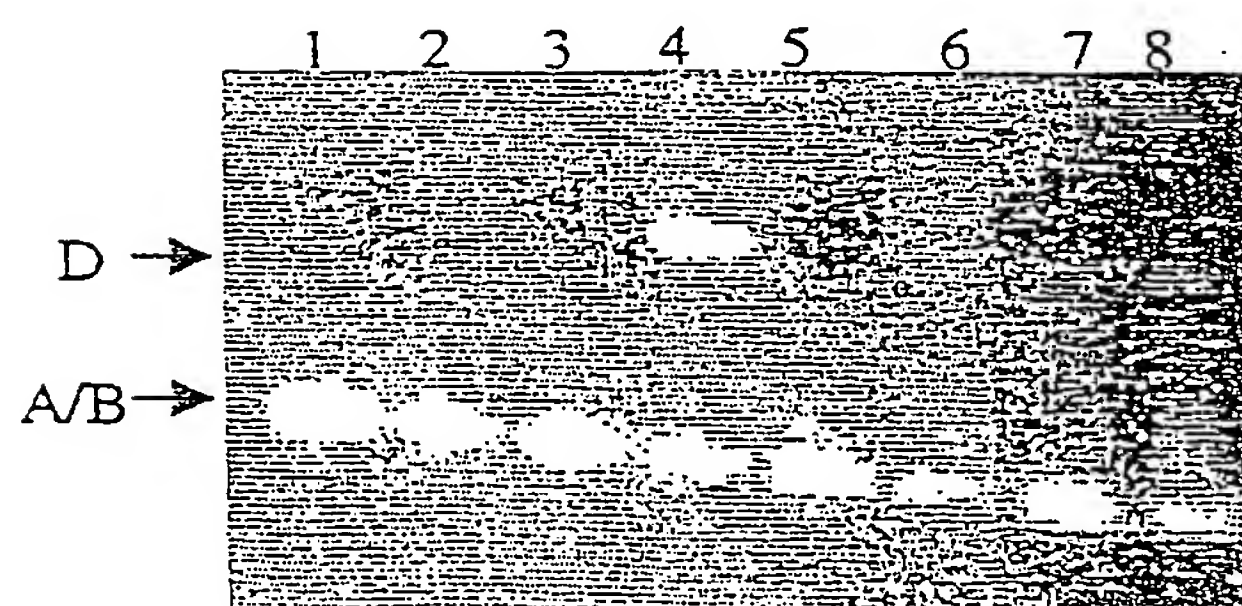


Figure 20

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	10	20	30	40	50	60
exon 1/2 A/B*	TGGCGGCGGCGACGGGATGGCTGCGCCGGCATTTCGCAGTTTCCGCGGCGGGGCTGGCCC					
exon 1/2 A/B	T-----GCGGCGACGGGATGGCTGCGCCGGCATTTCGCAGTTTCCGCGGCGGGGCTGGCCC					
exon 1/2 1 D	CGGCGGCGGCGACGGGATGGCTGCGCCGGCATTTCGCAGTTTCCGCGGCGGGGCTGGCCC					
exon 1/2 A/B	GGCCGTCGGCTCCTCGATCCGGCGGGGCAGAGCGGAGGGGGCGCGGGGTGGAGCTGCAGT					
exon 1/2 A/B	GGCCGTCGGCTCCTCGATCCGGCGGGGCAGAGCGGAGGGGGCGCGGGGTGGAGCTGCAGT					
exon 1/2 D	GGCCGTCGGCTCCTCGATCCGGCGGGGCAGAGCGGAGGGGGCGCGGGGTGGAGCTGCAGT					
exon 1/2 A/B	CGCCATCGCTGCTCTTCGGCCGCAACAAGGGCACCCGTTACCCC-----					
exon 1/2 A/B	CGCCATCGCTGCTCTTCGGCCGCAACAAGGGCACCCGTTACCCC-----					
exon 1/2 D	CGCCATCGCTGCTCTTCGGCCGCAACAAGGGCACCCGTTACCCCCTAATTATTTGCGCC					
exon 1/2 A/B	-----					
exon 1/2 A/B	-----					
exon 1/2 D	ACCTTTCTCACTCACATTCTCTCGTGTATTCTGTCTGCTCGCCCTTCGCCGACGACGC					
exon 1/2 A/B	-----					
exon 1/2 A/B	-----					
exon 1/2 D	GTGCCGATTCCGTATCGGGCTGCGGTGTTTCAGCGATCTTACGTCCGTTCCCTCCTGGTGT					
exon 1/2 A/B	-----GTGCCGTCGGCGTCGGAGGTTCTGGATGGCGCGTGGTCATGCGCGC					
exon 1/2 A/B	-----GTGCCGTCGGCGTCGGAGGTTCTGGATGGCGCGTGGTCATGCGCGC					
exon 1/2 D	GCTGATGTCTGTAGGTGCCGTCGGCGTCGGAGGTTCTGGATGGCGCGTGGTCATGCGCGC					
exon 1/2 A/B	GGGGGGCCGTCCGGGGAGGTGATGATCCCTGACGGCG					
exon 1/2 A/B	GGGGGGCCGTCCGGGGAGGTGATGATCCCTGACGGCG					
exon 1/2 D	GGGGGGCCGTCCGGGGAGGTGATGATCCCTGACGGCG					

Figure 21

SBE2_AL.DNA	1	TTC	10	20	30	40	50	60	70	80	90
SBE2_B.DNA	1	-----	-----	-----	-----	-----	-----	-----	-----	-----	CGGTATGCTT
SBE2_DL.DNA	1	TTC	10	20	30	40	50	60	70	80	90
	1	TTC	10	20	30	40	50	60	70	80	90
	1	TTC	10	20	30	40	50	60	70	80	90
	1	TTC	10	20	30	40	50	60	70	80	90
SBE2_AL.DNA	91	CGCT	100	110	120	130	140	150	160	170	180
SBE2_B.DNA	91	CGCT	100	110	120	130	140	150	160	170	180
SBE2_DL.DNA	91	CGCT	100	110	120	130	140	150	160	170	180
	91	CGCT	100	110	120	130	140	150	160	170	180
	91	CGCT	100	110	120	130	140	150	160	170	180
SBE2_AL.DNA	181	AAGT	190	200	210	220	230	240	250	260	270
SBE2_B.DNA	181	AAGT	190	200	210	220	230	240	250	260	270
SBE2_DL.DNA	181	AAGT	190	200	210	220	230	240	250	260	270
	181	AAGT	190	200	210	220	230	240	250	260	270
	181	AAGT	190	200	210	220	230	240	250	260	270
SBE2_AL.DNA	271	TACCGA	280	290	300	310	320	330	340	350	360
SBE2_B.DNA	271	TACCGA	280	290	300	310	320	330	340	350	360
SBE2_DL.DNA	271	TACCGA	280	290	300	310	320	330	340	350	360
	271	TACCGA	280	290	300	310	320	330	340	350	360
	271	TACCGA	280	290	300	310	320	330	340	350	360
SBE2_AL.DNA	361	CAACA	370	380	390	400	410	420	430	440	450
SBE2_B.DNA	361	CAACA	370	380	390	400	410	420	430	440	450
SBE2_DL.DNA	361	CAACA	370	380	390	400	410	420	430	440	450
	361	CAACA	370	380	390	400	410	420	430	440	450
	361	CAACA	370	380	390	400	410	420	430	440	450
SBE2_AL.DNA	451	-----	460	470	480	490	500	510	520	530	540
SBE2_B.DNA	451	-----	460	470	480	490	500	510	520	530	540
SBE2_DL.DNA	451	-----	460	470	480	490	500	510	520	530	540
	451	-----	460	470	480	490	500	510	520	530	540
	451	-----	460	470	480	490	500	510	520	530	540
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SBE2_B.DNA	541	TGGAT	550	560	570	580	590	600	610	620	630
SBE2_DL.DNA	541	TGGAT	550	560	570	580	590	600	610	620	630
	541	TGGAT	550	560	570	580	590	600	610	620	630
	541	TGGAT	550	560	570	580	590	600	610	620	630

Figure 22

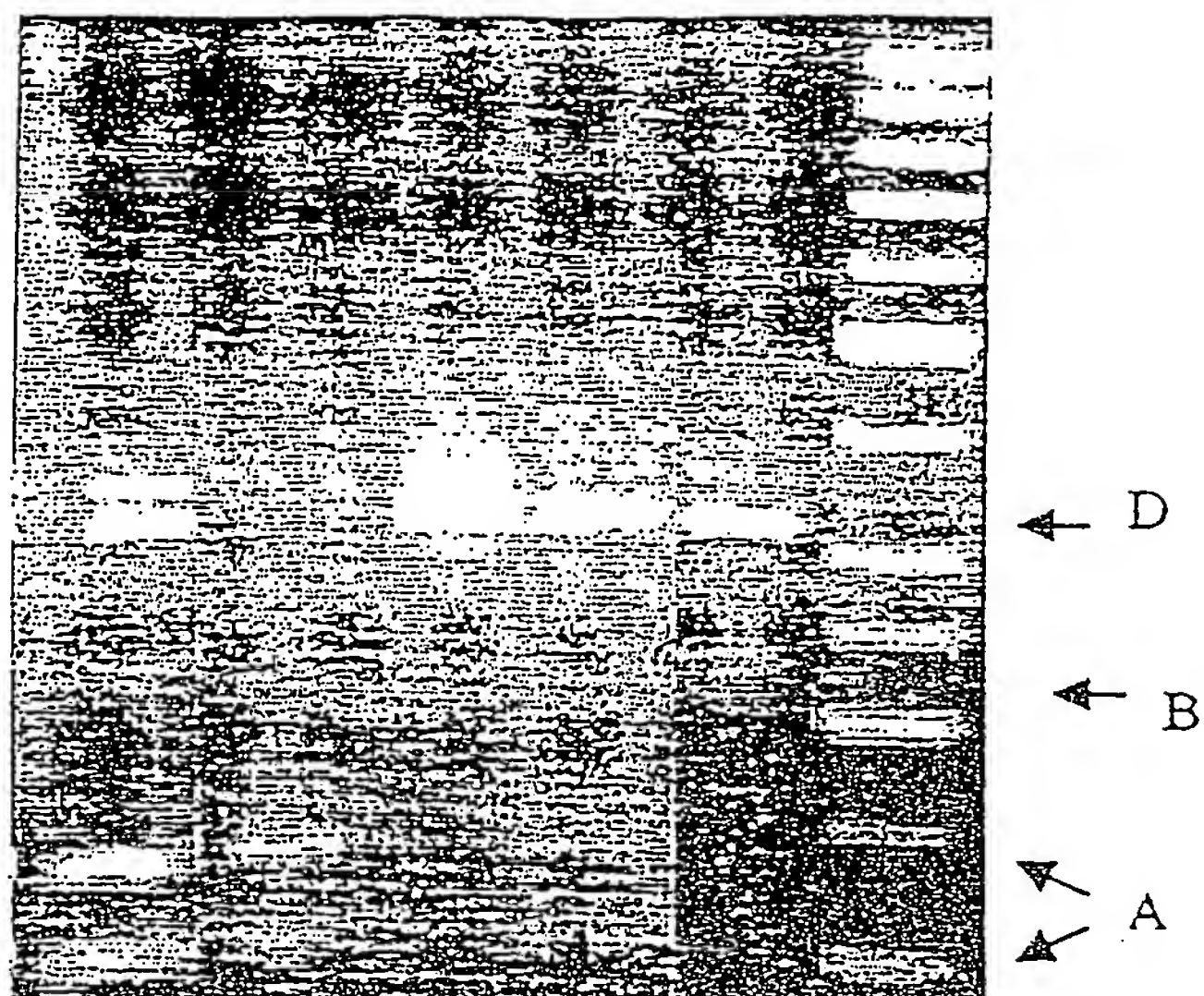


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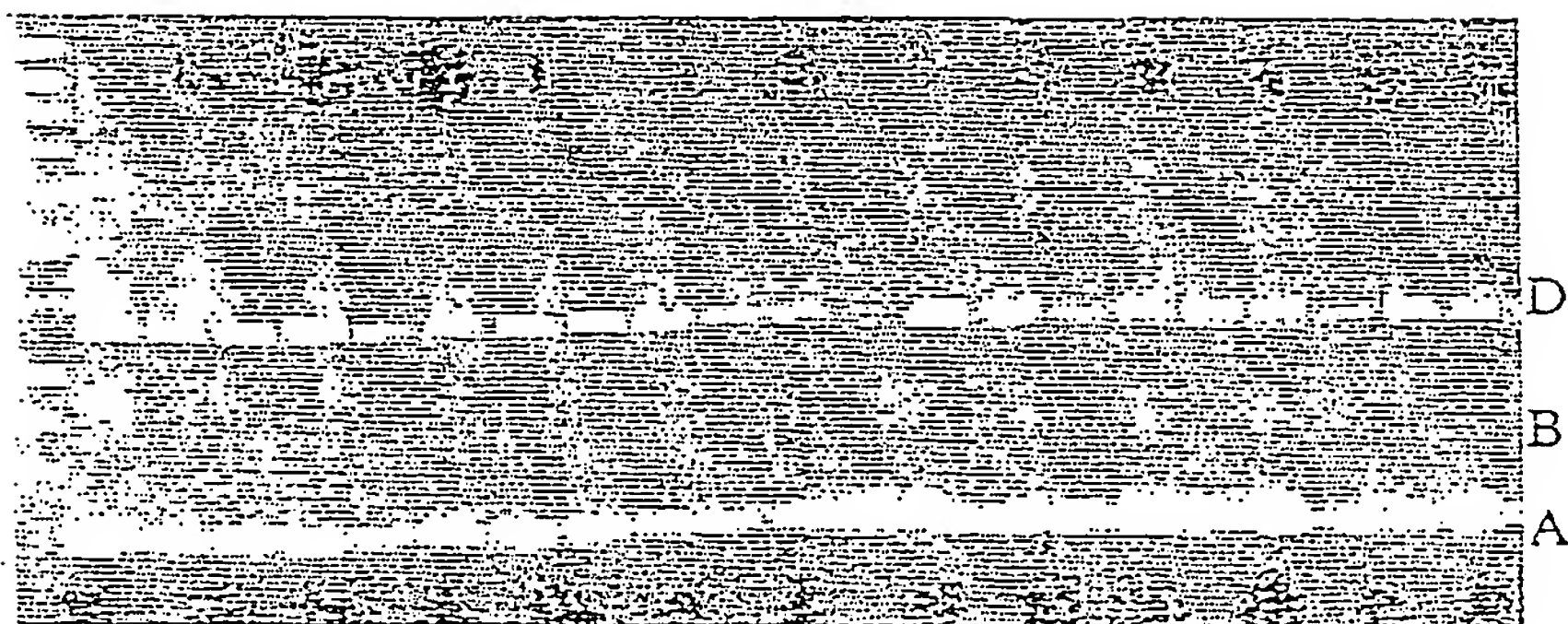


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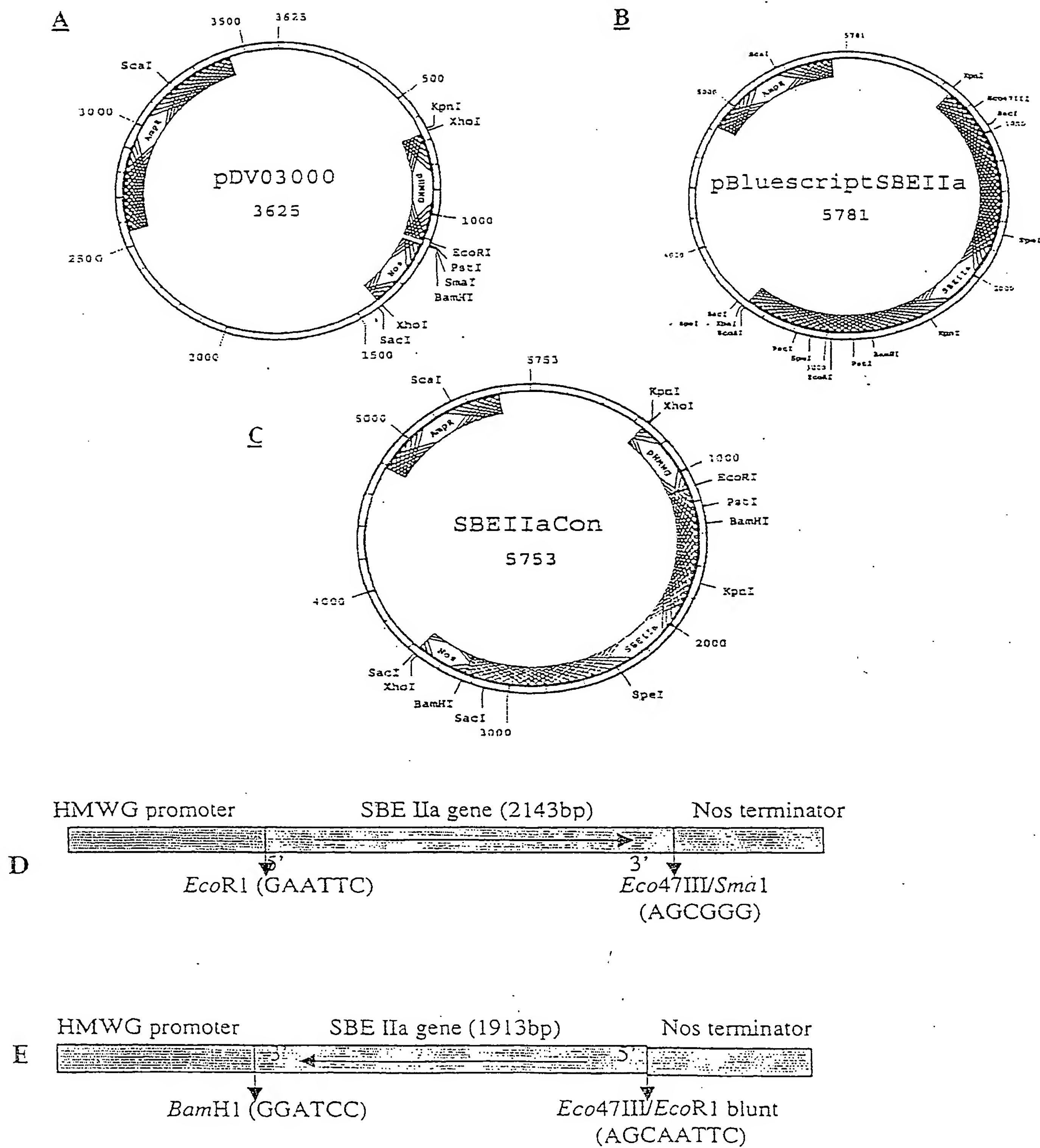


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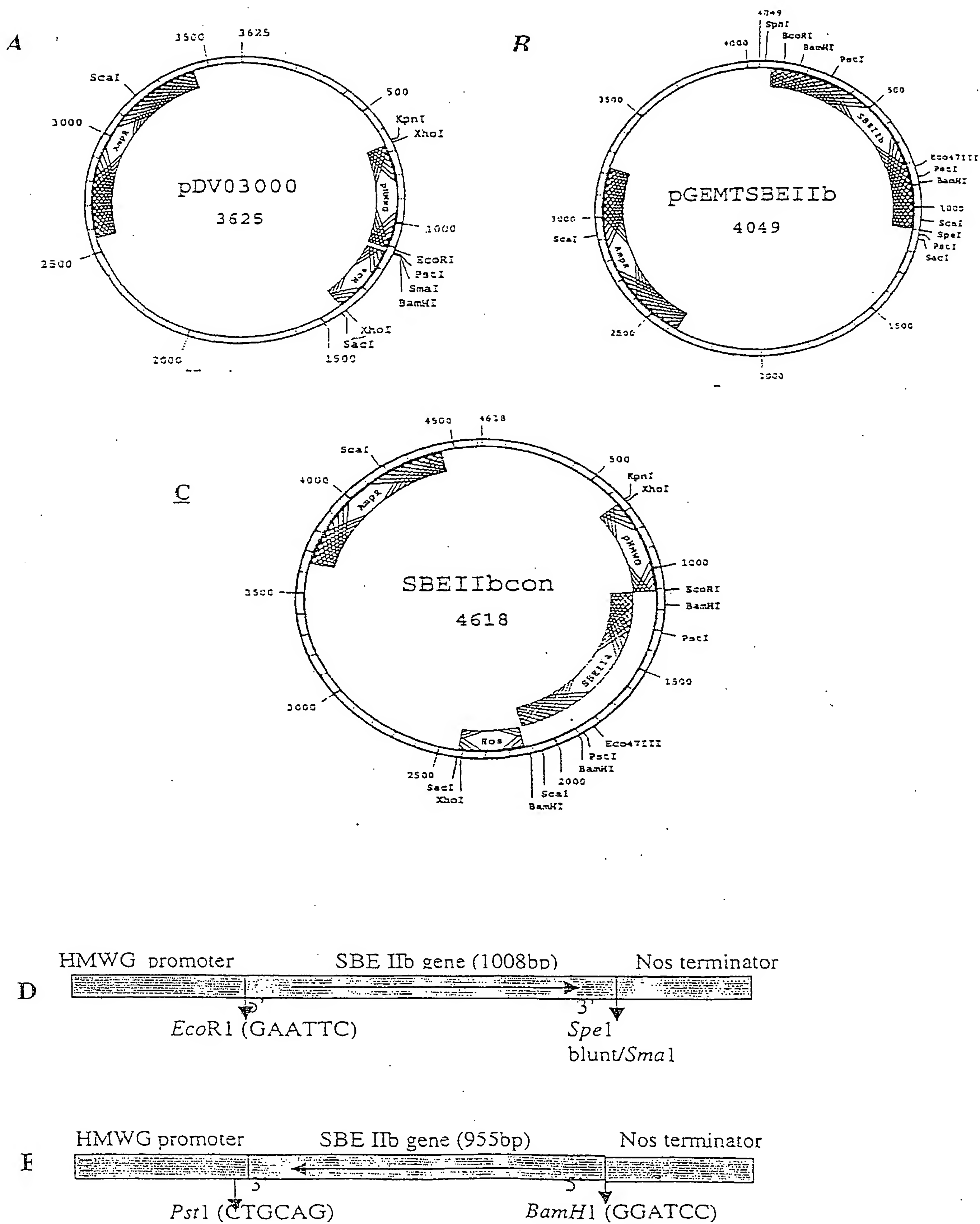


Figure 26

SEQUENCE LISTING

<110> CSIRO

Goodman Fielder Limited

Groupe Limagrain Pacific Pty Limited

<120> Starch Branching Enzymes

<130> FP14112

<140> PQ5742

<141> 2000-02-21

<150> PQ5742

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28

INTERNATIONAL SEARCH REPORT

International application No.

PCT/AU01/00175

A. CLASSIFICATION OF SUBJECT MATTER

Int. Cl. ⁷: C12N 15/29 A01H 5/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

DGENE: SEE ELECTRONIC DATA BASE BOX BELOW

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

DNA DATABASES: SEE ELECTRONIC DATA BASE BOX BELOW

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

GENBANK, EMBL: SEQ ID NOS 6 AND 10 DGENE: WHEAT BEW PEPTIDE SEQUENCE FROM FIGURE 10

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	SUN C et al "The two genes encoding starch-branching enzymes IIa and IIb are differentially expressed in barley" Plant Physiol (1998) 118, pages 37-49 See the entire document	1-52
X	GAO M et al "Evolutionary conservation and expression patterns of maize starch branching enzymes I and IIb genes suggests isoform specialization" Plant Mol Biol (1996) 30, pages 1223-32 See the entire document	1-52

☒ Further documents are listed in the continuation of Box C ☒ See patent family annex

* Special categories of cited documents:

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"&" document member of the same patent family

Date of the actual completion of the international search

6 April 2001

Date of mailing of the international search report

19 April 2001

Name and mailing address of the ISA/AU

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INTERNATIONAL SEARCH REPORT
Information on patent family members

International application No.
PCT/AU01/00175

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search
Report

Patent Family Member

WO	99 14314	AU	89670/98	EP	1012250
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